

## SUPPLEMENTARY FIGURES

### Supplementary Figure 1

**(A) DNA-sequence of mini-gp130-ELP:** normal print: cDNA coding for the LeB4-signal peptide, bold: cDNA coding for mini-gp130-ELP; Primer and restriction sites are indicated. The ATG start codon is underlined.

**(B) Protein sequence of mini-gp130-ELP.** Grey: LeB4-signal peptide; red: sgp130 domains 1-3; red, cursive: amino acid sequence determined by Edman-degradation of the mature purified mini-gp130-ELP; blue: c-myc-tag; green: 100 repeats of ELP; brown: ER-retention-signal.

### Supplementary Figure 2

#### Protein stability of purified mini-gp130-ELP at 37°C and 4°C.

**(A)** 1 µg purified mini-gp130-ELP was diluted in 1 ml PBS and incubated for 48 h at 37°C. Aliquots of originally 10 ng mini-gp130-ELP were separated by SDS/PAGE and blotted onto a PVDF membrane. Proteins were detected with the c-myc specific antibody 9E10 and visualized by ECL detection. Lane 1+2: 0 h; lane 3+4: 12 h; lane 5+6: 24 h; Lane 7+8: 48 h.

**(B)** 1 µg purified mini-gp130-ELP was diluted in 1 ml PBS and incubated for 26 d at 4°C. Aliquots of originally 10 ng mini-gp130-ELP were separated by SDS/PAGE and blotted onto a PVDF membrane. Proteins were detected with a c-myc specific antibody and visualized by ECL detection. Lane 1: 0 d; lane 2: 7 d; lane 3: 19 d; Lane 4: 26 d.

### Supplementary Figure 3

#### Expression of sgp130-variants in transgenic tobacco.

Leaves of transgenic lines were extracted in a mortar under liquid nitrogen in 50 mM Tris-HCl, 200 mM NaCl, 5 mM EDTA, 0.1% Tween 20, pH 8.0. The homogenate was centrifuged for 5 min at 4°C and 16,000 g. Proteins were separated on a 10% SDS polyacrylamide gel, blotted and c-myc-tag containing proteins were detected by Western blotting and ECL.

**(A)** Lane 1: 40 µg extract of sgp130Fc-ELP transgenic tobacco plants; lane 2: 40 µg extract of sgp130Fc transgenic tobacco plants; lane 3: 40 ng c-myc-tagged scFv control protein. M: molecular mass marker (kDa).

**(B)** Lane 1: 5 µg extract of mini-gp130-ELP transgenic tobacco plants; lane 2: 5 ng control protein; lane 3: 10 ng c-myc-tagged single chain Fv control protein; lane 4: 5 µg extract of mini-gp130-ELP transgenic tobacco plants. M: molecular mass marker (kDa).

**Supplementary Figure 4 Calibration of size exclusion chromatography.**

**(A)** For calibration the high molecular mass standard from Amersham Pharmacia Biotech was used. Peak 1: Thyroglobin 669 kD; Peak 2: Ferritin 490 kDa; Peak 3: Katalase 232 kDa; Peak 4: Aldolase 158 kDa.

**(B)** The known molecular masses of the purchased standard proteins and their respective elution volumes were subject to linear regression.

**Supplementary Figure 5 Absorption spectra of pure mini-gp130-ELP**

**(A)** The absorption spectrum of purified mini-gp130-ELP was recorded in the range of 240-320 nm.

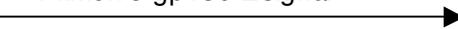
**(B)** The protein concentration of purified mini-gp130-ELP (after inverse transition cycling and size exclusion chromatography) was calculated as indicated from the absorption at 280 nm.

# Supplementary Figure 1

**A**

1 ATGGCTTCCA AACCTTTTCT ATCTTTGCTT TCAC~~TT~~TCCT TGCTTCTCTT

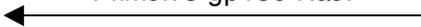
Primer: 5'gp130-Δ signal



AflIII

51 TACAAGCACA TGT~~TT~~AGCAG AGCTGCTGGA TCCTTGCGGC TATATCTCCC  
 101 CTGAGTCTCC TGTGGTGCAG CTGCATTCTA ACTTCACCGC CGTGTGTGTG  
 151 CTGAAGGAAA AGTGCATGGA C~~T~~ACTTCCAC GTGAACGCCA ACTACATCGT  
 201 GTGGAAAACC AACCACTTCA CCATCCCCAA GGAGCAGTAC ACCATCATCA  
 251 ACCGGACCGC TTCTTCTGTG ACCTTCACCG ATATCGCCTC CCTGAATATC  
 301 CAGCTGACCT GCAACATCCT GACCTTTGGA CAGCTGGAGC AGAATGTGTA  
 351 CGGCATCACC ATCATCTCTG GCCTGCCTCC AGAGAAGCCT AAGAACCTGT  
 401 CCTGCATCGT GAATGAGGGC AAGAAGATGA GGTGTGAGTG GGATGGCGGC  
 451 AGAGAGACAC ATCTGGAGAC CAACTTCACC CTGAAGTCTG AGTGGGCCAC  
 501 CCACAAGTTT GCCGACTGCA AGGCCAAGAG AGATACCCTT ACCTCTTGCA  
 551 CCGTGGACTA CTCCACCGTG TACTTCGTGA ACATCGAGGT GTGGGTGGAG  
 601 GCTGAGAATG CTCTGGGCAA GGTGACCTCT GACCACATCA ACTTCGACCC  
 651 CGTGTACAAG GTGAAGCCTA ACCCTCCTCA CAACCTGTCC GTGATCAACT  
 701 CTGAGGAGCT GTCCTCTATC CTGAAGCTGA CCTGGACCAA CCTTCCATC  
 751 AAGTCCGTGA TCATCTGAA GTACAACATC CAGTACAGGA CCAAGGATGC  
 801 TTCTACCTGG TCTCAGATCC CTCTGAGGA TACCGCTTCC ACCAGATCCA  
 851 GCTTCACAGT GCAGGACCTG AAGCCTTTTA CCGAGTACGT GTTCAGGATC  
 901 CGGTGCATGA AGGAGGATGG CAAGGGCTAT TGGTCTGACT GGTCTGAGGA

Primer: 3'gp130-NaeI



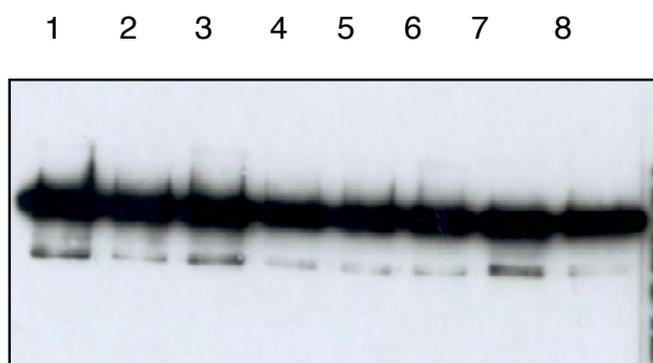
NaeI

951 GGCTTCTGGC ATCACCTACG AGGACAGAGC CCGCGGACAA GCGGCCGAG  
 1001 AACAAAACT CATCTCAGAA GAGGATCTGA ATGGGGCCGT CGAGATGGGC  
 1051 CACGGCGTGG GTGTTCCGGG CGTGGGTGTT CCGGGTGGCG GTGTGCCGGG  
 1101 CGCAGGTGTT CCTGGTGTAG GTGTGCCGGG TGTGGGTGTG CCGGGTGTG  
 1151 GTGTACCAGG TGGCGGTGTT CCGGGTGCAG GCGTTCGGG TGGCGGTGTG  
 1201 CCGGGCGTGG GTGTTCCGGG CGTGGGTGTT CCGGGTGGCG GTGTGCCGGG  
 1251 CGCAGGTGTT CCTGGTGTAG GTGTGCCGGG TGTGGGTGTG CCGGGTGTG  
 1301 GTGTACCAGG TGGCGGTGTT CCGGGTGCAG GCGTTCGGG TGGCGGTGTG  
 1351 CCGGGCGTGG GTGTTCCGGG CGTGGGTGTT CCGGGTGGCG GTGTGCCGGG  
 1401 CGCAGGTGTT CCTGGTGTAG GTGTGCCGGG TGTGGGTGTG CCGGGTGTG  
 1451 GTGTACCAGG TGGCGGTGTT CCGGGTGCAG GCGTTCGGG TGGCGGTGTG  
 1501 CCGGGCGTGG GTGTTCCGGG CGTGGGTGTT CCGGGTGGCG GTGTGCCGGG  
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 1951 CCGGGCGTGG GTGTTCCGGG CGTGGGTGTT CCGGGTGGCG GTGTGCCGGG  
 2001 CGCAGGTGTT CCTGGTGTAG GTGTGCCGGG TGTGGGTGTG CCGGGTGTG  
 2051 GTGTACCAGG TGGCGGTGTT CCGGGTGCAG GCGTTCGGG TGGCGGTGTG  
 2101 CCGGGCGTGG GTGTTCCGGG CGTGGGTGTT CCGGGTGGCG GTGTGCCGGG  
 2151 CGCAGGTGTT CCTGGTGTAG GTGTGCCGGG TGTGGGTGTG CCGGGTGTG  
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 2251 CCGGGCGTGG GTGTTCCGGG CGTGGGTGTT CCGGGTGGCG GTGTGCCGGG  
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 2351 GTGTACCAGG TGGCGGTGTT CCGGGTGCAG GCGTTCGGG TGGCGGTGTG  
 2401 CCGGGCGTGG GTGTTCCGGG CGTGGGTGTT CCGGGTGGCG GTGTGCCGGG  
 2451 CGCAGGTGTT CCTGGTGTAG GTGTGCCGGG TGTGGGTGTG CCGGGTGTG  
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 2551 CCGGGCGGGC TGGCGGCCGC AGAACCCAAA GACGAACTCT AG

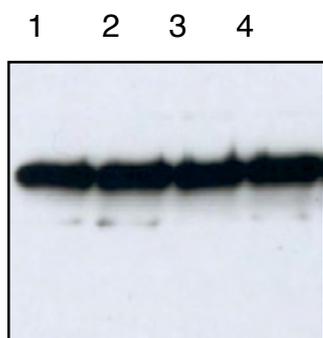


# Supplementary Figure 2

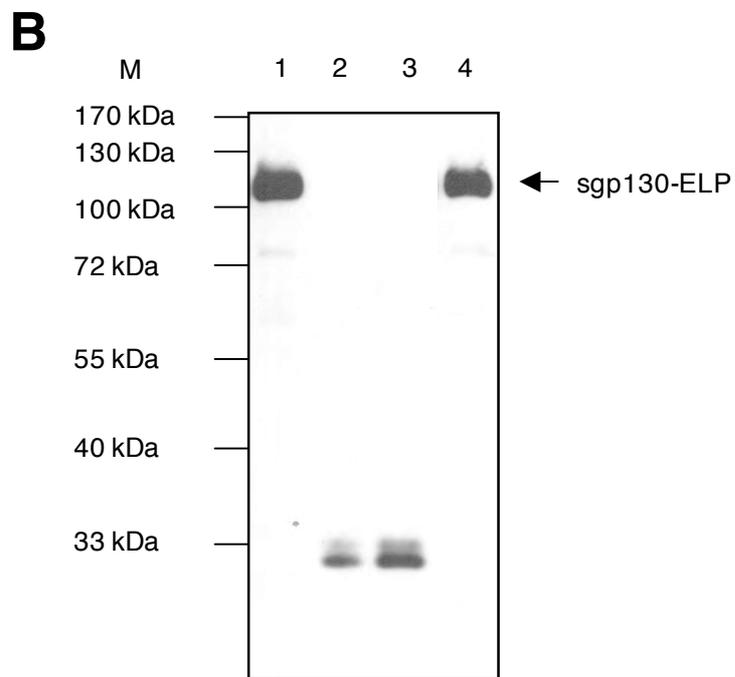
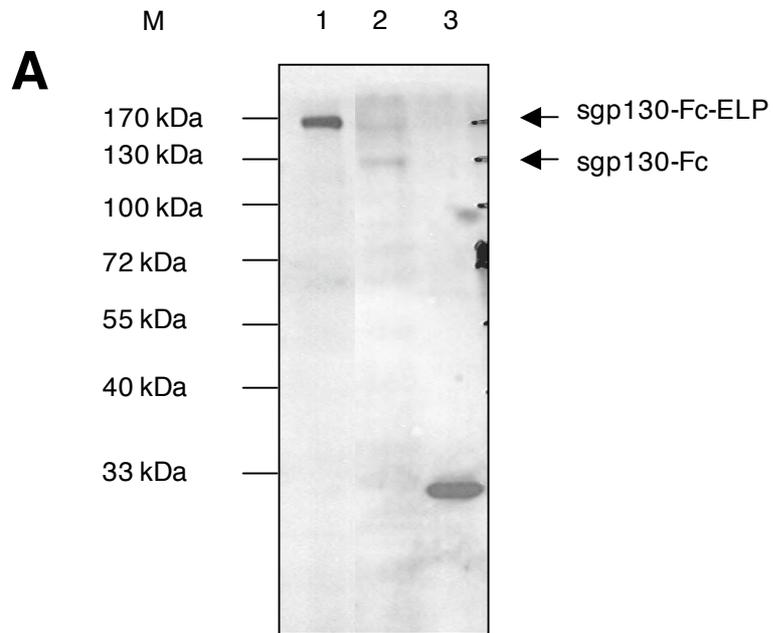
**A**



**B**

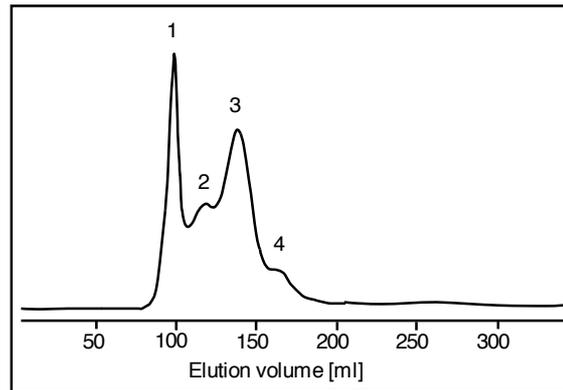


# Supplementary Figure 3

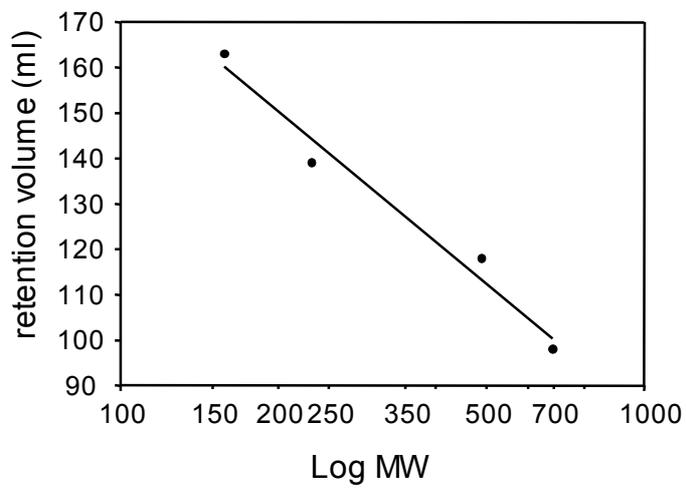


# Supplementary Figure 4

**A**



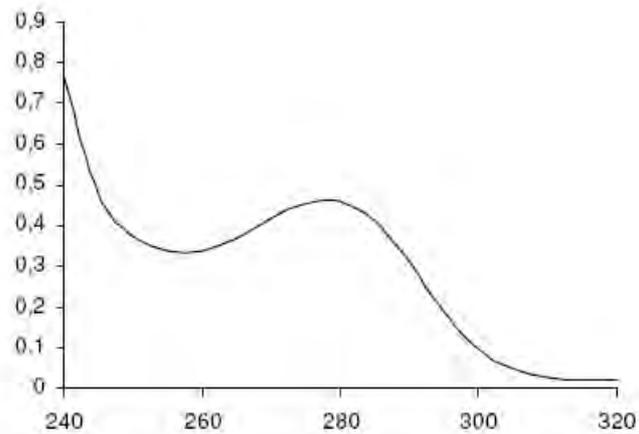
**B**



$$y = -95.5x + 370.1$$
$$r^2 = 0.973$$

# Supplementary Figure 5

**A**



nm	Abs
240	0,766
245	0,478
250	0,374
255	0,336
260	0,333
265	0,37
270	0,417
275	0,455
280	0,458
285	0,458
290	0,413
295	0,316
300	0,199
305	0,098
310	0,048
315	0,029
320	0,022

**B**

$$c = \frac{0.458 \times 100,000 \text{ g/mol}}{64,890 \text{ l/cm} \times \text{mol} \times 1 \text{ cm}} = 0.705 \text{ mg/ml}$$