

ELECTRONIC APPENDIX

This is the Electronic Appendix to the article

Impact of UV-B exposure on amphibian embryos; linking species physiology and oviposition
behavior

by

Wendy J. Palen, Craig E. Williamson, Aaron A. Clauser and Daniel E. Schindler

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Electronic appendices are refereed with the text; however, no attempt is made
to impose a uniform editorial style on the electronic appendices.

Section 1.

Solar phototron experiments, details by species.

Species	Start date	Stage ¹	Duration (days)	UV-B dose treatments (kJ·m ⁻²)	Collection area
<i>Rana cascadae</i>	July 19	8-10	3 *	0 119 204 264 426	Olympic National Park, WA
<i>Pseudacris regilla</i>	March 20	5-7	3 *	0 120 205 265 428	N. Cascade Mountains, WA
<i>Ambystoma gracile</i>	March 18	8-10	1	0 40 69 89 143	N. Cascade Mountains, WA
<i>Ambystoma macrodactylum</i>	July 17	10-13	1	0 50 86 111 179	Olympic National Park, WA

Notes: All phototron experiments were carried out at 15° C under a 12:12 photoperiod. Stage refers to the level of embryonic development at the start of the experiment. In addition to the 3-day exposure experiments reported here, preliminary one day experiments were carried out for both frog species (*), where no mortality was observed. Subsequent experiments for all species were designed to run until near 100% mortality was observed in the most UV-B exposed treatments. For each species at each level of UV-B exposure except 0 (dark control), we had treatments with and without photoreactivating radiation (PRR), resulting in 9 treatments per experiment. Differences in total UV-B dose are the result of different configurations of stainless-steel mesh, lamp outputs, and experiment duration. For comparison, the integrated daily dose of UV-B received at Olympic National Park (WA) during the summer solstice (June 21, 2000) at 1525 m was 129.7 kJ·m⁻².

¹ *R. cascadae* and *P. regilla* staged according to: Gosner, K.L., 1960. A simplified table for staging anuran embryos and larvae with notes on identification. *Herpetologica*. **16**, 183-190. *A. gracile* and *A. macrodactylum* staged according to: Harrison, R.G. 1969. *In* Organization and Development of the Embryo. Yale University Press. Pp. 44-66.

Section 2.

Non-linear model estimates of solar phototron experiments

Species	Photorepair	r^2	LD_{50} ($\text{kJ}\cdot\text{m}^{-2}$)	Steepness (a)
<i>Rana cascadae</i>	+ PRR	0.89	350 \pm 29	8.7 \pm 3.2
	- PRR	0.91	124 \pm 17	3.6 \pm 2.2
<i>Pseudacris regilla</i>	+ PRR	0.83	285 \pm 31	6.4 \pm 4.0
	- PRR	0.76	108 \pm 9	13.0 \pm 0
<i>Ambystoma gracile</i>	+ PRR	0.91	95 \pm 8	4.2 \pm 1.6
	- PRR	0.90	2 \pm 83	0.1 \pm 3.1
<i>Ambystoma macrodactylum</i>	+ PRR	0.87	100 \pm 6	7.6 \pm 3.3
	- PRR	0.92	24 \pm 58	1.0 \pm 3.3

Notes: Data are mean-corrected r^2 and parameter estimates (\pm SD), lethal dose corresponding to 50% survival (LD_{50}) and steepness (a), of Gauss-Newton non-linear least squares model fits for treatments with (+PRR) and without (-PRR) photorepair radiation.

Section 3.

Field survey details by species.

Species	Sites surveyed	Eggs/site	K_d (m^{-1})	Depth (cm)	% eggs shaded
<i>Rana cascadae</i>	30	3.8*	-0.223	5.5*	3.7
<i>Ambystoma gracile</i>	11	3.9*	-0.168	34.0*	0
<i>Ambystoma macrodactylum</i>	21	478	-0.136	18.2	51.0

Notes: Summary of amphibian egg surveys. Data are averages across breeding sites for the number of eggs or egg masses (*) per site, the diffuse attenuation coefficient (K_d , m^{-1}), and depth (cm) of eggs.