### Plant Gene Register

# Primary Structure and Characterization of a cDNA Clone of Fructokinase from Potato (Solanum tuberosum L. cv Record)<sup>1</sup>

#### Susan B. Smith\*, Mark A. Taylor, Lindsay R. Burch, and Howard V. Davies

Department of Cellular and Environmental Physiology, Scottish Crop Research Institute, Invergowrie, Dundee DD2 5DA, United Kingdom

The major source of carbon for starch synthesis in reserve tissues such as the potato (*Solanum tuberosum* L.) tuber is Suc. In the developing tuber, Suc is hydrolyzed principally by Suc synthase (EC 2.4.1.13) to UDP-Glc and Fru (Preiss, 1982). Fructokinase (EC 2.7.1.4) is believed to catalyze the phosphorylation of the Fru released to yield Fru-6-P. This sugar phosphate can then be used to support starch synthesis following further metabolism in the cytosol and amyloplast. There are some reports (Wolosiuk and Pontis, 1974) that Suc synthase activity can undergo feedback inhibition by free Fru. In this case fructokinase potentially plays an important role in maintaining the flux of carbon toward starch formation.

Fructokinase has been purified from a variety of plants including pea (Turner et al., 1977; Copeland et al., 1984), barley (Baysdorfer et al., 1989), avocado (Copeland and Tanner, 1988), and developing potato tubers (Gardner et al., 1992), but to date the corresponding gene has never been cloned from higher plants. A cDNA clone of the potato fructokinase gene has been isolated as a first step toward investigating more fully the role that fructokinase plays in carbohydrate metabolism in potato (Table I).

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The GenBank/EMBL accession number for the sequence reported in this article is Z12823.

#### LITERATURE CITED

Baysdorfer C, Kremer DF, Sicher RC (1989) Partial purification and characterisation of fructokinase activity from barley leaves. J Plant Physiol 134: 156–161

## **Table I.** Characteristics of a cDNA clone encoding potato fructokinase

#### Organism:

Potato (Solanum tuberosum L. cv Record).

Gene, Function, and Pathway:

Fructokinase (EC 2.7.1.4); phosphorylation of Fru to Fru-6-P in the starch biosynthetic pathway.

Techniques:

cDNA library screen using polyclonal antibody for pea fructokinase; dideoxy sequencing of both strands using successive oligonucleotide primers.

Method of Identification:

Amino acid sequence comparison with *scrK* genes of *Vibrio alginolyticus* (39% identity; 63% similarity using GAP alignment package); cross-reaction with both pea and potato polyclonal antibodies.

Expression Characteristics:

A 1.2-kb transcript detected in swelling stolons, which increased during tuber development. Low levels were detected in leaves and stems taken from tuberizing plants.

Gene Copy Numbers:

Restriction digest analysis of genomic DNA indicated that the fructokinase gene is present in low copy number.

Features of cDNA Structure:

Contains an open reading frame of 957 bp. There is an ATG initiation codon at nucleotide 11 and a TAG termination codon at 968. A polyadenylation signal is present at 1119.

Structural Features of Protein:

Copeland L, Stones SR, Turner JF (1984) Kinetic studies of fructokinase I of pea seeds. Arch Biochem Biophys 233: 748–760

- Copeland L, Tanner GJ (1988) Hexose kinases of avocado. Physiol Plant 74: 531-536
- Gardner A, Davies HV, Burch LR (1992) Purification and properties of fructokinases from developing tubers of potato (*Solanum tuberosum*). Plant Physiol 100: 178–183

Preiss J (1982) Regulation of the biosynthesis and degradation of starch. Annu Rev Plant Physiol 33: 431–454

Turner JF, Harrison DD, Copeland L (1977) Fructokinase (fraction IV) of peas. Plant Physiol 60: 666–669

Wolosiuk RA, Pontis HG (1974) Studies on sucrose synthetase. Arch Biochem Biophys 165: 140–145

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<sup>\*</sup> Corresponding author; fax 44-382-562426.

G+C Content:

<sup>50.4%.</sup> 

Polypeptide of 323 amino acids with a calculated mol wt of 34,392.