# Comparison of Methods to Estimate Dark Respiration in the Light in Leaves of Two Woody Species<sup>1</sup>

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Dark respiration in the light was estimated in leaves of two woody species (Heteromeles arbutifolia Ait. and Lepechinia fragans Greene) using two different approaches based on gas-exchange techniques: the Kok method and the Laisk method. In all cases, dark respiration in the light was lower (P < 0.05) than respiration in darkness, indicating that dark respiration was inhibited in the light. Rates of dark respiration in the light estimated by the Laisk method were 52% higher (P < 0.05) than those estimated by the Kok method. Differences between the methods could be explained by the low ambient CO<sub>2</sub> concentrations required by the Laisk approach. The mean value of the inhibition of respiration by light for the two species, corrected for the ambient CO2 concentration effect, was 55%. Despite the differences in leaf characteristics between the species, values of the CO<sub>2</sub> photocompensation point, at which the rate of photosynthetic CO2 uptake equaled that of photorespiratory CO2 evolution, were very constant, suggesting an excellent consistency in the results obtained with the Laisk approach.

In most studies of the carbon balance of plants or plant organs it is assumed that dark respiration in the light continues at the same rate as in darkness. However, there is evidence that light inhibits dark respiration in photosynthetic tissue (Kok, 1948; Sharp et al., 1984; Kirschbaum and Farquhar, 1987). This inhibition appears to be caused by metabolites from photosynthesis (ATP, NADPH) acting on the respiratory enzymes as respiratory regulators (Graham, 1980; McCashing et al., 1988). However, the mechanism of inhibition is not clear and appears to be complex (Gardeström and Wigge, 1988; Krömer and Heldt, 1991).

The instantaneous evolution of  $CO_2$  in the light is a result of at least four processes, which take place at different rates: photosynthesis, photorespiration, dark respiration in the light ( $R_d$ ), and refixing of  $CO_2$  from respiration (Graham, 1980). Thus, techniques for estimating  $R_d$  are complicated.

To estimate  $R_d$  by analysis of gas exchange, two main methods are used. The first, described by Kok (1948), analyzes the response of **A** to light at low intensities. The

response is linear at low levels of irradiation, but near the light compensation point there is a break in the linear response, increasing markedly the slope of the light curve due to a decrease in **A**. This has been interpreted as a result of an increase in the respiration rate due to a progressive disappearance of the light-induced inhibition of dark respiration (Kok, 1948; Sharp et al., 1984). Extrapolation of the linear section of the light curve before the change of slope to a light intensity of zero gives an estimate of  $R_d$ .

One of the main pitfalls of this method is that the decrease of PPFD during the construction of the light curves induces a gradual increase of  $c_i$  and in turn a relative increase in the value of **A**. As a result, the slope of the regression of **A** versus PPFD decreases. This underestimates the true value of  $R_{d}$ , yielding an  $R_d^a$ . To cope with this problem in the estimation of the true value of  $R_d$ , we corrected the value of  $R_d^a$  following the approach of Kirschbaum and Farquhar (1987). With this approach, the value of  $R_d^a$  obtained from the light-assimilation curve is corrected by considering the relationships between both the rate of ribulose-1,5-bisphosphate regeneration and the **A** with the PPFD over the range of the measurement and by assuming that the dark respiration in the light is independent of both  $c_i$  and PPFD.

The second method, described by Laisk (1977) and extended by Brooks and Farquhar (1985), analyzes **A** at low  $c_i$  values and varying light intensities. **A** can be expressed as:

$$\mathbf{A} = v_{\rm c} - 0.5 \ v_{\rm o} - R_{\rm d} \tag{1}$$

where  $v_c$  and  $v_o$  are the rate of carboxylation and oxygenation, respectively. The CO<sub>2</sub> concentration at which CO<sub>2</sub> uptake from carboxylation and CO<sub>2</sub> loss from photorespiration are equal is that at which  $v_c = 0.5 v_o$ . This CO<sub>2</sub> concentration has been called  $\Gamma_{\bullet}$ , which is related to the CO<sub>2</sub>/O<sub>2</sub> specificity of Rubisco. From Farquhar and von Caemmerer (1981), Equation 1 becomes:

$$\mathbf{A} = v_c \left(1 - \Gamma \cdot / [\mathrm{CO}_2]\right) - R_d \tag{2}$$

Equation 2 shows that, at a CO<sub>2</sub> concentration equal to  $\Gamma_{\bullet}$ , **A** is equal to  $-R_{d}$ . Analysis by the Laisk method is aimed at

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Abbreviations: **A**, net CO<sub>2</sub> assimilation rate;  $c_i$ , intercellular CO<sub>2</sub> concentration;  $\Gamma$ , CO<sub>2</sub> photocompensation point at which photosynthetic CO<sub>2</sub> uptake equals photorespiratory CO<sub>2</sub> evolution;  $R_d$ , rate of dark respiration in the light;  $R_d^a$ , apparent value of  $R_d$ ;  $R_n$ , rate of respiration in darkness.

finding the internal concentration of  $CO_2$  ( $\Gamma$ -) at which the rate of photosynthesis equals that of photorespiration. Given that at this internal  $CO_2$  concentration all of the  $CO_2$  from photorespiration is consumed in the photosynthetic process, the rate of  $CO_2$  evolution under these conditions represents  $R_d$ . The main disadvantage of this method is that the experiments must be performed at very low  $CO_2$  concentrations and are therefore under far from normal environmental conditions. In addition, this method assumes that the  $CO_2/O_2$  specificity of Rubisco and  $R_d$  are independent of the light intensity. However, changes in the value of  $R_d$  with light intensity have been reported (Brooks and Farquhar, 1985).

Because these methods are based on different approaches and use different experimental conditions, it may be presumed that they provide different estimates of  $R_d$ . As far as we know, there is no work comparing the results obtained by the two methods for analysis of the same plant material. The objective of this study was to compare the results of  $R_d$ obtained by these two methods.

#### MATERIALS AND METHODS

#### **Plant Material**

Two woody species, typical of the Californian chaparral, were chosen for the study: *Heteromeles arbutifolia* (Ait.), an evergreen shrub, and *Lepechinia fragans* (Greene), a deciduous shrub. Two-year-old plants were grown in pots in an experimental garden. During the 2 months previous to the experiments, temperature ranged from 20 to 25°C during the day and from 10 to 15°C at night, and the PPFD at midday was close to 600  $\mu$ mol photons m<sup>-2</sup> s<sup>-1</sup>. For the analysis, well-developed leaves with similar characteristics (age, size, aspect, and specific leaf area) of each species were used.

## **Gas-Exchange System**

Gas exchange in leaves was measured with an open gasexchange system as described by Held et al. (1991). Before the experiments, the IRGA (Analytical Development Co., Hoddesdon, United Kingdom; model 225 MK3) was calibrated at the low CO<sub>2</sub> concentrations used in the present study, as recommended by Bloom et al. (1980). IRGA sensitivity was constant over the range of CO2 concentrations used. The CO<sub>2</sub> concentration was measured in dry air by using a water trap filled with anhydrous magnesium perchlorate. The desired air humidity was obtained by mixing humidified and dry air. Air humidity was measured with RH sensors (Weathermeasure Corp., Sacramento, CA). The vapor pressure deficit during the experiments was approximately 1 kPa. The boundary layer conductance of the chamber was 2 mol  $m^{-2}$  s<sup>-1</sup>. The light source was a metal halide lamp (Sylvania GTE, Danvers, MA) suspended above the chamber. A tray with circulating water was placed between the lamp and the chamber to absorb IR radiation. Different PPFDs were obtained by covering the chamber with nylon filters of different extinction coefficients. The incident PPFD was measured inside the chamber with a gallium arsenide photocell (Hamamatsu, San Jose, CA) previously calibrated with a LI-190S sensor (Lambda Instruments, Lincoln, NE).

Gas-exchange calculations were made according to Ball

(1987), and photosynthetic and respiration rates were expressed per unit leaf dry weight. In the calculations, cuticular conductances were assumed to be zero because of the thick cuticle of leaves of the Mediterranean woody species (Lillis, 1991). All measurements were made on individual leaves attached to the plant at a leaf temperature of 20°C. The experiments were started at the same time each day (early in the morning) to avoid variations in the respiration rate due to changes in the concentration of carbohydrates (Azcón-Bieto et al., 1983).

## **Kok Method**

A values were obtained at low PPFD, decreasing in steps from 40 to 0  $\mu$ mol photons m<sup>-2</sup> s<sup>-1</sup>. The partial pressure of O<sub>2</sub> and CO<sub>2</sub> were 21 kPa and 35 Pa, respectively. For each PPFD, the leaf was allowed to stabilize for 1 h to reach a constant value of **A**. The photosynthetic rates recorded before the change in the slope of the light curve were regressed over the corresponding PPFD. The intercept of the regression line with the ordinate (light intensity zero) gave  $R_d^a$ . For each leaf,  $R_d^a$  was adjusted to a constant  $c_i$  using the program reported by Kirschbaum and Farquhar (1987) to obtain a corrected value of  $R_d$ .

 $R_n$  was measured at the beginning and end of each experiment by covering the chamber with a black cloth at an ambient CO<sub>2</sub> concentration of 350  $\mu$ L L<sup>-1</sup>. In most cases, it was unnecessary to wait more than 45 min to achieve stable  $R_n$  values. Measurements of **A** at low PPFDs are presented only in those cases in which  $R_n$  was similar at the beginning and end of the experiment, as recommended by Sharp et al. (1984).

## Laisk Method

On the day following the Kok experiments,  $R_d$  was estimated on the same leaves by the method of Laisk (1977). For each leaf, the experiments were repeated at three different low PPFDs (40, 75, and 145 µmol photons m<sup>-2</sup> s<sup>-1</sup>). After PPFD had been selected, **A** values were measured at decreasing  $c_i$  values (usually in the range 90–30 µL L<sup>-1</sup>). For each PPFD the linear regression of **A** over  $c_i$  was calculated. At the three light intensities considered, the linear regressions crossed at one point where the rate of photosynthesis equals that of photorespiration. The coordinates of this point of **A** and  $c_i$  represent  $R_d$  and  $\Gamma_{\bullet}$ , respectively, and were found graphically.

#### Effect of Light Intensity on R<sub>d</sub>

To explore the effect of light intensity on the value of  $R_d$ , the Laisk method was used for a medium-aged leaf of *H. arbutifolia*. However, in the present case, instead of a set of only three different low PPFDs (see the above paragraph), a wide array of PPFDs (550, 410, 300, 140, 75, 40, 20, and 9  $\mu$ mol photons m<sup>-2</sup> s<sup>-1</sup>) was considered in the analysis. For each of the selected PPFDs, *A* at different low  $c_i$  values was estimated, and the linear regressions of *A* versus  $c_i$  were calculated. Finally, for each neighboring pair of regression lines, their intersection point ( $\Gamma$ -,  $R_d$ ) was determined analytically and the value of  $R_d$  was retained. This procedure gave seven values of  $R_d$ , corresponding to the pairs of PPFD: 550–410, 410–300, 300–140, 140–75, 75–40, 40–20, and 20–9  $\mu$ mol photons m<sup>-2</sup> s<sup>-1</sup>.

## Effect of CO<sub>2</sub> Concentration on R<sub>n</sub>

To analyze the effect of low ambient  $CO_2$  concentration on respiration,  $R_n$  was measured in a set of leaves similar to those used in the previous experiments. A total of seven leaves was considered. Leaf temperature was kept at 20°C, and  $R_n$  was estimated first at an ambient  $CO_2$  concentration of 350  $\mu$ L L<sup>-1</sup> after a stabilization period of 45 min. Then ambient  $CO_2$  concentration in the cuvette was changed to 35  $\mu$ L L<sup>-1</sup>, and the respiration rate was estimated after allowing the leaf to stabilize for 1 h.

## RESULTS

## $R_{\rm d}$

Figures 1 and 2 show an example of the results obtained by both methods with a leaf of *L. fragans*. In the case of the Kok method (Fig. 1), there was a change in slope of the straight line at approximately 5  $\mu$ mol photons m<sup>-2</sup> s<sup>-1</sup>. The intercept of the regression line (open symbols) with the ordinate represents  $R_d^a$ . That value was 1.73 nmol CO<sub>2</sub> g<sup>-1</sup> s<sup>-1</sup> (arrow in the figure) for the leaf considered in the example.

Figure 2 shows the results obtained with the same leaf using the Laisk method. The regression lines of **A** over  $c_i$  at different PPFDs intercepted at one point, having the coordinates  $\Gamma$  and  $R_d$ , where  $R_d$  equals 2.87 nmol CO<sub>2</sub> g<sup>-1</sup> s<sup>-1</sup> (arrow in the figure).

In the two species the average values of  $R_d^a$  by the Kok method were lower (about 28%, P < 0.05, Student's *t* test) than those corrected following the method of Kirschbaum and Farquhar (1987) (Table I). The mean values of  $R_d$  for *H*.



**Figure 1.** A at limiting PPFDs of incident PAR (method of Kok) for a leaf of *L. fragans.* Lines represent linear regressions before (O) and after ( $\bullet$ ) the inflection point of the light curve. The arrow indicates the value of  $R_d^a$ .



**Figure 2. A** as a function of  $c_i$  ( $\mu$ L L<sup>-1</sup>) (Laisk method) for the same leaf as in Figure 1. Numbers in the figure indicate the incident PPFD under which measurements were made ( $\mu$ mol photons m<sup>-2</sup> s<sup>-1</sup>). Lines represent linear regressions at these PPFDs. The arrow indicates the value of  $R_d$  for that particular leaf.

*arbutifolia* obtained by the methods of Kok (after correcting for the  $c_i$  effect) and Laisk were 2.27 and 3.06 nmol CO<sub>2</sub> g<sup>-1</sup> s<sup>-1</sup>, respectively, and for *L. fragans* they were 2.49 and 3.79 nmol CO<sub>2</sub> g<sup>-1</sup> s<sup>-1</sup>, respectively (Table I).

Within each species, and comparing individual leaves, the values of  $R_d$  obtained by the Laisk method were always higher (about 52%, P < 0.01, Student's *t* test) than those obtained by the Kok method. In contrast, for each particular method, there were no significant differences between the mean values of  $R_d$  of the two species studied (Student's *t* test). Finally, it should be noted that the values of  $\Gamma$  were similar in the leaves of the two species (Table I).

## Effect of Light Intensity on R<sub>d</sub>

Figure 3 shows the relationship between the measured values of  $R_d$  at different PPFDs. The relationship was linear (r = -0.84, P < 0.05) and was defined by the equation:

$$R_{\rm d} = 2.3521 - 3.9568 \times 10^{-3} \times I \tag{3}$$

where I is the PPFD.

This equation can be used for predicting  $R_d$  at different PPFDs, and it shows that the value of  $R_d$  decreases from 2.05 nmol CO<sub>2</sub> g<sup>-1</sup> s<sup>-1</sup> at 75 µmol photons m<sup>-2</sup> s<sup>-1</sup> (the average PPFD used in the Laisk experiments) to values near zero at PPFD of approximately 500 µmol photons m<sup>-2</sup> s<sup>-1</sup>.

The average PPFD values used in the Kok and the Laisk experiments in this study were 20 and 75  $\mu$ mol photons m<sup>-2</sup> s<sup>-1</sup>, respectively. For those values, Equation 3 predicts the values of R<sub>d</sub> of 2.27 and 2.05 nmol CO<sub>2</sub> g<sup>-1</sup> s<sup>-1</sup>, respectively, for the leaf of *H. arbutifolia*. These values are quite close (difference less than 10%).

**Table I.** Mean values of  $R_d$  (methods of Kok and Laisk),  $R_n$ , and  $\Gamma_*$  in the two species In the case of the Kok method,  $R_d^a$  is before any correction for  $c_i$  and  $R_d$  is after this correction. See the text for explanation.

	R <sub>d</sub>				
	Kok			R <sub>n</sub>	$\Gamma_{\star}$
	$R_d^a$	R <sub>d</sub>	Laisk		
	nmol CO <sub>2</sub> g <sup>-1</sup> s <sup>-1</sup>				μL L <sup>-1</sup> CO <sub>2</sub>
H. arbutifolia					
Mean	1.63	2.27	3.06	4.40	35.7
$s_{D}(n=4)$	±0.68	±0.67	±0.74	±1.28	±0.8
L. fragans					
Mean	1.80	2.49	3.79	6.25	35.3
sD(n=5)	±0.44	±0.76	±0.78	±2.09	±1.7

## **R**<sub>n</sub>

Table I shows the values of  $R_n$  in the leaves of the two species in a normal CO<sub>2</sub> atmosphere (350 µL L<sup>-1</sup>). No significant differences were detected (Student's *t* test) between the mean values of  $R_n$  of the two species studied. The mean values of  $R_n$  in *H. arbutifolia* (4.40 nmol CO<sub>2</sub> g<sup>-1</sup> s<sup>-1</sup>) and *L.* fragans (6.25 nmol CO<sub>2</sub> g<sup>-1</sup> s<sup>-1</sup>) were higher than the values of  $R_d$ . There were significant differences between  $R_d$  obtained by the Kok method and  $R_n$  (P < 0.05, Student's *t* test).

Figure 4 shows the relationship between the measured values of  $R_n$  at ambient concentrations of CO<sub>2</sub> of 35 and 350  $\mu$ L L<sup>-1</sup>. The relationship was linear (r = 0.95, P < 0.01) and was defined by the equation:

$$R_{\rm n} = -0.17724 + 0.69639 \times R_{\rm n \ 35} \tag{4}$$

where  $R_n$  and  $R_{n 35}$  were estimated at ambient CO<sub>2</sub> concentrations of 350 and 35  $\mu$ L L<sup>-1</sup>, respectively.

This equation indicated that  $R_n$  at 350  $\mu$ L L<sup>-1</sup> CO<sub>2</sub> was lower (approximately 30%) than that at 35  $\mu$ L L<sup>-1</sup> CO<sub>2</sub>.



**Figure 3.** Relationship between  $R_d$  and incident PPFD in *H. arbuti-folia*. The line represents the regression  $y = 2.3521 - 3.9568 \times 10^{-3} \times x$  (r = 0.84, P < 0.05).

## DISCUSSION

In spite of the differences between the leaves of the two species, i.e. chemical composition, specific leaf area, leaf duration (Merino et al., 1984), the mean values of  $R_d$  for these two species were similar (Table I). However, there were strong, consistent differences depending on the method used to determine  $R_d$ . The method of Kok, used more commonly to separate respiration in light from that in darkness (Kok, 1948; Sharp et al., 1984; Kirschbaum and Farquhar, 1987), is more direct than that of Laisk.

It must be pointed out that, in spite of the laborious procedure of the Laisk method, the values of  $\Gamma$ - are practically identical in all leaves measured. This is as expected, since the value of  $\Gamma$ - is dependent mainly on the kinetic characteristics of Rubisco, which should be similar in leaves of the same plant population and possibly in leaves of species of similar ecology (Jordan and Ogren, 1981). In fact, the average values



**Figure 4.** Relationship between  $R_n$  measured at 350 and that at 35  $\mu$ L L<sup>-1</sup> CO<sub>2</sub> concentration in air ( $R_n$  and  $R_n$  <sub>35</sub>, respectively) in *H*. arbutifolia ( $\bullet$ ) and *L*. fragans (O) leaves. The line represents the regression  $R_n = -0.17724 + 0.69639 \times Rn$  <sub>35</sub> (r = 0.95, P < 0.01).

of  $\Gamma$ - found for the two species considered (35.7 and 35.3  $\mu$ L L<sup>-1</sup> CO<sub>2</sub>) are similar to the average value (35.5  $\mu$ L L<sup>-1</sup> CO<sub>2</sub>) found by Brooks and Farquhar (1985) for leaves of *Spinacia oleracea* in experiments conducted at the same temperature (20°C) as in the present study.

All of the above underline the high uniformity in the specificity of Rubisco and, in addition, indicate a high consistency in the results yielded by the Laisk method. The agreement between the published values of  $\Gamma$ - and those found in the present study suggests the existence of no significant errors in the application of the method.

The question is raised as to the reason for the higher  $R_d$  values obtained by the Laisk method (Table I). The difference observed between the average values of  $R_d$  obtained by the methods of Kok and Laisk (Table I) may not be explained as a consequence of the different average light intensity during the experiments (20 and 75  $\mu$ mol photons m<sup>-2</sup> s<sup>-1</sup>, respectively), since the values of  $R_d$  predicted for these light intensities using Equation 3 were very close (2.27 and 2.05 nmol CO<sub>2</sub> g<sup>-1</sup> s<sup>-1</sup>, a difference of less than 10%).

The high values of  $R_d$  obtained by the method of Laisk for the two species (Table I) may be related to the sensitivity of respiratory processes to the low concentration of CO<sub>2</sub> used for this experimental procedure. In fact, the results showed an obvious depressing effect of an increase in the ambient CO<sub>2</sub> concentration on  $R_n$  (Fig. 4). Such a depressing effect has been reported in other studies (Gifford et al., 1985; Reuveni and Gale, 1985; Amthor et al., 1992) and has been explained by CO<sub>2</sub> acting as an inhibitor of certain enzymes (e.g. succinate dehydrogenase; Shaish et al., 1987). However, other authors (Kirschbaum and Farquhar, 1987) did not find any response of  $R_d$  to an ambient CO<sub>2</sub> concentration increase.

Unfortunately, the experimental conditions required for the Laisk method do not allow direct exploration of the effect of CO<sub>2</sub> concentration on the value of  $R_d$ . Although the mechanism of the action of CO<sub>2</sub> on the respiration rate is not well known, there is no reason to suppose that the responses of  $R_n$  and  $R_d$  to CO<sub>2</sub> concentration will be different. In principle, the respiratory processes in the light do not seem essentially different from those taking place in darkness, since the physiological pathways are the same and the main difference appears to be related to the degree of activity of the enzymes involved (Graham, 1980). The similarity of the responses of  $R_d$  and  $R_n$  to a series of environmental factors (Brooks and Farquhar, 1985) agrees with this supposition.

If this is the case, the relationship between  $R_d$  estimated at ambient CO<sub>2</sub> concentrations of 35 and 350  $\mu$ L L<sup>-1</sup> should be similar to that for  $R_n$  at these same concentrations (Fig. 4) and expressed by Equation 4. Values of 1.96 and 2.46 nmol CO<sub>2</sub> g<sup>-1</sup> s<sup>-1</sup> are obtained for *H. arbutifolia* and *L. fragans*, respectively, using this equation to estimate the mean values of  $R_d$  (by the Laisk method) expected at an ambient CO<sub>2</sub> concentration of 350  $\mu$ L L<sup>-1</sup>. These new values for the Laisk method are closer to the values of  $R_d$  obtained by the Kok method in these two species (2.27 and 2.49 nmol CO<sub>2</sub> g<sup>-1</sup> s<sup>-1</sup>, respectively; Table I), making the differences between methods nonsignificant.

These results show that the  $R_d$  values in the two species are significantly lower (P < 0.05) than the  $R_n$  values (Table I). This indicates that dark respiration is partially inhibited in the light in the leaves of the two species. The average value for the degree of inhibition of dark respiration by light for the two species is approximately 55% (54% by the Kok method and 56% by the Laisk method). Results of other authors (Sharp et al., 1984; Brooks and Farquhar, 1985; Kirschbaum and Farquhar, 1987) show that the degree of inhibition in different species of agricultural or silvicultural interest ranges between 17 and 66%.

The nature of the inhibition may be due to the inhibiting effect of the light on the respiratory enzymes mediated by different cofactors, such as NADPH and ATP (Graham, 1980; McCashing et al., 1988). Although the ATP and reducing power generated in the photosynthetic process may follow many different pathways (Gardeström and Wigge, 1988; Krömer and Heldt, 1991), the utilization of ATP and NADPH produced directly from photosynthesis may decrease the requirements for ATP and NADH having catabolic origin and thus decrease respiration rates, resulting in a significant increase in growth efficiency (Raven, 1976).

The results show that during the day the average daytime rates of foliar respiration ( $R_d$ ) in woody species can be at least 55% lower than the rates during the night ( $R_n$ ) and even lower during a sunny day. This should be taken into account in the modeling of carbon balance.

In summary, the methods for estimating  $R_d$  considered in the present work are dependent on distinct factors; therefore, the results are not directly comparable when using distinct approaches. However, by correcting both **A** for  $c_i$  (Kok method) and  $R_d$  for the atmospheric CO<sub>2</sub> concentration (Laisk method), the agreement of the results by both methods should be quite acceptable.

The difficulty in detecting the change of slope in the light curves at low light intensities (Kok method) and the length of these experiments (usually a minimum of 5 h per leaf, in contrast to an average of 2 h in the case of the Laisk method) makes the latter more appropriate for cases in which the number of determinations is high and the  $R_n$  dependence on ambient CO<sub>2</sub> concentration is known.

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## LITERATURE CITED

- Amthor JS, Koch GW, Bloom AJ (1992) CO<sub>2</sub> inhibits respiration in leaves of *Rumex crispus* L. Plant Physiol **98**: 757–760
- Azcón-Bieto J, Lambers H, Day DA (1983) Effect of photosynthesis and carbohydrate status on respiratory rates and the involvement of the alternative pathway in leaf respiration. Plant Physiol 72: 598–603
- Ball JT (1987) Calculations related to gas exchange. In E Zeiger, GD Farquhar, IR Cowan, eds, Stomatal Function. Stanford University Press, Stanford, CA, pp 445–476
- Bloom AJ, Mooney HA, Björkman O, Berry JA (1980) Materials and methods for carbon dioxide and water exchange analysis. Plant Cell Environ 3: 371–376

- Brooks A, Farquhar GD (1985) Effect of temperature on the CO<sub>2</sub>/ O<sub>2</sub> specificity of ribulose-1,5-bisphosphate carboxylase/oxygenase and the rate of respiration in the light. Planta 165: 397–406
- Farquhar GD, von Caemmerer S (1981) Modelling of photosynthetic response to environmental conditions. In OL Lange, PS Nobel, CB Osmond, H Ziegler, eds, Encyclopedia of Plant Physiology, Vol 12B: Physiological Plant Ecology. Water Relations and Photosynthetic Productivity. Springer-Verlag, Berlin, pp 549–587
- Gardeström P, Wigge B (1988) Influence of photorespiration on ATP/ADP ratios in the chloroplasts, mitochondria, and cytosol, studied by rapid fractionation of barley (*Hordeum vulgare*) protoplasts. Plant Physiol 88: 69-76
- Gifford RM, Lambers H, Morison JIL (1985) Respiration of crop species under CO<sub>2</sub> enrichment. Physiol Plant 63: 351–356
- Graham D (1980) Effects of light and "dark" respiration. In DD Davies, ed, The Biochemistry of Plants. A Comprehensive Treatise, Vol 2. Academic Press, New York, pp 525–579
- Held AA, Mooney HA, Gorham JN (1991) Acclimation to ozone stress in radish: leaf demography and photosynthesis. New Phytol 118: 417-423
- Jordan DB, Ogren WL (1981) Species variation in the specificity of ribulose bisphosphate carboxylase/oxygenase. Nature 291: 513-515
- Kirschbaum MUF, Farquhar GD (1987) Investigation of the CO<sub>2</sub> dependence of quantum yield and respiration in *Eucalyptus pauciflora*. Plant Physiol 83: 1032–1036

- Kok B (1948) A critical consideration of the quantum yield of *Chlorella*-photosynthesis. Enzymologia 13: 1–56
- Krömer S, Heldt HW (1991) On the role of mitochondrial oxidative phosphorylation in photosynthesis metabolism as studied by the effect of oligomycin on photosynthesis in protoplasts and leaves of barley (*Hordeum vulgare*). Plant Physiol **95**: 1270–1276
- Laisk AK (1977) Kinetics of Photosynthesis and Photorespiration in C<sub>3</sub>-Plants. Nauka, Moscow
- Lillis M (1991) An ecomorphological study of the evergreen leaf. Braun-Blanquetia 7: 1-127
- McCashin BG, Cossins EA, Canvin DT (1988) Dark respiration during photosynthesis in wheat leaf slices. Plant Physiol 87: 155-161
- Merino JA, Field CB, Mooney HA (1984) Construction and maintenance costs of Mediterranean-climate evergreen and deciduous leaves. Oecol Plant 5: 211–229
- **Raven JA** (1976) The quantitative role of dark respiration processes in heterotrophic and photolithotrophic plant growth. Ann Bot **40**: 587–602
- **Reuveni J, Gale J** (1985) The effect of high levels of carbon dioxide on dark respiration and growth of plants. Plant Cell Environ 8: 623-628
- Shaish A, Roth-Dejerano N, Itai C (1989) The response of stomata to CO<sub>2</sub> relates to its effect on respiration and ATP level. Physiol Plant 76: 107-111
- Sharp RE, Matthews MA, Boyer JS (1984) Kok effect and the quantum yield of photosynthesis: light partially inhibits dark respiration. Plant Physiol 75: 95-101