

**Table 3. Summary of *in situ* hybridization**

Gene	Probe*	Distribution of hybridization signals in MBs†				No. of MBs	Total point‡	Rank‡
		(-)	(±)	(+)	(++)			
CG4842	GH15220	96.7	3.3	0.0	0.0	30	3.3	D
CG13244	GH27987	100.0	0.0	0.0	0.0	42	0.0	D
CG13297	RE64858	66.7	33.3	0.0	0.0	24	33.3	C
CG32473	RE62048	68.3	0.0	31.7	0.0	60	63.4	B
Rab10	LD39986	3.1	15.6	43.8	37.5	32	215.7	A
CG12866	Genomic PCR	83.3	16.7	0.0	0.0	24	16.7	D
CG5476	RE59605	23.3	10.0	46.7	20.0	30	163.4	A
CG18266	GH11218	63.3	23.3	6.7	6.7	30	56.8	B
Kif3C	GH04118	63.2	10.5	7.9	18.4	38	81.5	B
CG13312	Genomic PCR	60.7	28.6	10.7	0.0	28	50.0	B
CG2070	LP06328	35.3	26.5	26.5	11.8	34	114.9	A
CG2196	Genomic PCR	29.4	17.6	26.5	26.5	34	150.1	A
CG13350	SD03057	81.3	12.5	6.3	0.0	32	25.1	C
CG11164	RE50056	100.0	0.0	0.0	0.0	32	0.0	D
CG14313	Genomic PCR	87.5	0.0	12.5	0.0	56	25.0	C
CG13566	Genomic PCR	92.6	7.4	0.0	0.0	27	7.4	D
CG32676	GH03394	69.2	3.8	15.4	11.5	26	69.1	B
Hr46	GH21112	36.0	0.0	20.0	44.0	50	172.0	A
Dhod	GH24729	81.2	0.0	18.8	0.0	80	37.6	C
CG14446	GM27257	62.9	0.0	37.1	0.0	62	74.2	B
CG2267	GH11073	28.9	10.5	13.2	47.4	38	179.1	A
CG3994	RE54080	90.0	10.0	0.0	0.0	30	10.0	D
CG15185	Genomic PCR	56.8	27.3	6.8	9.1	44	68.2	B

**Table 3. Summary of *in situ* hybridization**

CG14014	RE73577	45.5	0.0	54.5	0.0	22	109.0	A
CG14628	Genomic PCR	94.7	5.3	0.0	0.0	38	5.3	D
CG3371	GH13810	85.5	0.0	14.5	0.0	62	29.0	C
CG3818	Genomic PCR	76.7	23.3	0.0	0.0	30	23.3	C
CG14911	Genomic PCR	38.5	26.9	34.6	0.0	26	96.1	B
CG4575	Genomic PCR	100.0	0.0	0.0	0.0	32	0.0	D
CG14460	Genomic PCR	100.0	0.0	0.0	0.0	30	0.0	D
CG10205	LP09070	100.0	0.0	0.0	0.0	26	0.0	D
fkh	RE03865	41.2	0.0	38.2	20.6	68	138.2	A
trio	GH01987	50.0	21.9	28.1	0.0	32	78.1	B
CG33143	Genomic PCR	80.0	6.7	3.3	10.0	30	43.3	C
CG13444	RE33441	82.1	0.0	3.6	14.3	28	50.1	B
CG7945	LD01950	37.5	0.0	62.5	0.0	32	125.0	A
jing	LD36562	45.3	0.0	54.7	0.0	86	109.4	A
CG6974	Genomic PCR	94.1	5.9	0.0	0.0	34	5.9	D
CG7846	LD24980	96.4	3.6	0.0	0.0	28	3.6	D
CG13716	Genomic PCR	84.4	12.5	3.1	0.0	32	18.7	D
CG12301	LD31322	73.8	0.0	26.2	0.0	42	52.4	B
CG4940	Genomic PCR	79.2	16.7	4.2	0.0	24	25.1	C
CG10505	Genomic PCR	96.9	0.0	0.0	3.1	32	9.3	D
CG31559	RE15313	33.3	66.7	0.0	0.0	30	66.7	B
CG6372	GH13022	80.6	0.0	19.4	0.0	36	38.8	C
CG31530	RE24196	9.4	0.0	15.6	75.0	32	256.2	A
Brf	LD32109	50.0	0.0	50.0	0.0	12	100.0	A
CG12744	GH03826	100.0	0.0	0.0	0.0	48	0.0	D

**Table 3. Summary of *in situ* hybridization**

CG13604	LD48066	65.4	3.8	23.1	7.7	26	73.1	B
klg	GH05072	74.0	0.0	24.0	2.0	50	54.0	B
CG15057	Genomic PCR	93.3	6.7	0.0	0.0	30	6.7	D
RluA-1	RH09496	66.7	33.3	0.0	0.0	30	33.3	C
CG13159	RE25177	96.4	3.6	0.0	0.0	28	3.6	D
ey	pBSEyE10.1.1	33.3	0.0	31.3	35.4	48	168.8	A
CG17319	Genomic PCR	15.8	3.7	57.9	2.6	38	127.3	A
CG11178	LD19061	61.8	0.0	29.4	8.8	34	85.2	B
CG8412	RE17513	0.0	10.0	13.3	76.7	30	266.7	A
CG17221	RH24774	69.2	7.7	23.1	0.0	26	53.9	B
CG13176	RH05781	75.0	3.6	21.4	0.0	28	46.4	C
CG9296	RE06748	10.7	35.7	42.9	10.7	28	153.6	A
Hrb87F	GH05625	76.7	0.0	23.3	0.0	86	46.6	C
CG12253	LD32974	97.6	2.4	0.0	0.0	41	2.4	D
CG15579	Genomic PCR	97.1	2.9	0.0	0.0	34	2.9	D
CG13879	Genomic PCR	70.4	18.5	0.0	11.1	27	51.8	B
CG1234	LD35257	71.9	18.8	9.4	0.0	32	37.6	C
ltp-r83A	Genomic PCR	70.6	8.8	8.8	11.8	34	61.8	B
CG3983	AT23067	20.0	26.7	23.3	30.0	30	163.3	A
CG6083	AT02556	97.2	2.8	0.0	0.0	36	2.8	D
PP2A-B'	LD29902	38.9	0.0	19.4	41.7	36	163.9	A
Sema-1a	RE36155	41.2	8.8	11.8	38.2	34	147.0	A
elk	HL01061	56.7	0.0	43.3	0.0	30	86.6	B
Atg5	LD34980	94.1	5.9	0.0	0.0	34	5.9	D
CG4853	GH27815	92.9	0.0	0.0	7.1	28	21.3	C

**Table 3. Summary of *in situ* hybridization**

<b>RN-tre</b>	<b>LD31492</b>	<b>66.7</b>	<b>0.0</b>	<b>33.3</b>	<b>0.0</b>	<b>96</b>	<b>99.9</b>	<b>B</b>
<b>Pkn</b>	<b>RH55776</b>	<b>7.7</b>	<b>0.0</b>	<b>11.5</b>	<b>80.8</b>	<b>26</b>	<b>265.4</b>	<b>A</b>
<b>CG10948</b>	<b>RE29010</b>	<b>81.0</b>	<b>0.0</b>	<b>19.0</b>	<b>0.0</b>	<b>58</b>	<b>38.0</b>	<b>C</b>
<b>Nep1</b>	<b>GH03315</b>	<b>50.0</b>	<b>0.0</b>	<b>36.7</b>	<b>13.3</b>	<b>60</b>	<b>113.3</b>	<b>A</b>
<b>CG5669</b>	<b>LD04007</b>	<b>58.8</b>	<b>0.0</b>	<b>11.8</b>	<b>29.4</b>	<b>34</b>	<b>111.8</b>	<b>A</b>
<b>CG10880</b>	<b>LD25288</b>	<b>94.1</b>	<b>5.9</b>	<b>0.0</b>	<b>0.0</b>	<b>34</b>	<b>5.9</b>	<b>D</b>
<b>CG13213</b>	<b>SD01170</b>	<b>40.9</b>	<b>0.0</b>	<b>59.1</b>	<b>0.0</b>	<b>22</b>	<b>118.2</b>	<b>A</b>
<b>CG10083</b>	<b>SD28142</b>	<b>100.0</b>	<b>0.0</b>	<b>0.0</b>	<b>0.0</b>	<b>32</b>	<b>0.0</b>	<b>D</b>
<b>CG14483</b>	<b>Genomic PCR</b>	<b>93.8</b>	<b>6.3</b>	<b>0.0</b>	<b>0.0</b>	<b>32</b>	<b>6.3</b>	<b>D</b>
<b>Rpb11</b>	<b>GM15177</b>	<b>92.6</b>	<b>0.0</b>	<b>7.4</b>	<b>0.0</b>	<b>54</b>	<b>14.8</b>	<b>D</b>
<b>Dp1</b>	<b>SD20908</b>	<b>58.6</b>	<b>0.0</b>	<b>35.7</b>	<b>5.7</b>	<b>70</b>	<b>88.5</b>	<b>B</b>
<b>mod(mdg4)</b>	<b>SD03001</b>	<b>57.9</b>	<b>0.0</b>	<b>42.1</b>	<b>0.0</b>	<b>76</b>	<b>84.2</b>	<b>B</b>
<b>CG5384</b>	<b>LD26251</b>	<b>43.3</b>	<b>16.7</b>	<b>10.0</b>	<b>30.0</b>	<b>30</b>	<b>126.7</b>	<b>A</b>
<b>tra2</b>	<b>RH39422</b>	<b>64.8</b>	<b>0.0</b>	<b>35.2</b>	<b>0.0</b>	<b>88</b>	<b>70.4</b>	<b>B</b>
<b>usp</b>	<b>LD09973</b>	<b>72.7</b>	<b>0.0</b>	<b>27.3</b>	<b>0.0</b>	<b>22</b>	<b>54.6</b>	<b>B</b>
<b>fra</b>	<b>RE61427</b>	<b>42.3</b>	<b>38.5</b>	<b>11.5</b>	<b>7.7</b>	<b>26</b>	<b>84.6</b>	<b>B</b>
<b>CG7328</b>	<b>RE49431</b>	<b>60.0</b>	<b>0.0</b>	<b>40.0</b>	<b>0.0</b>	<b>30</b>	<b>80.0</b>	<b>B</b>
<b>CG3305</b>	<b>RE72002</b>	<b>71.4</b>	<b>21.4</b>	<b>3.6</b>	<b>3.6</b>	<b>28</b>	<b>39.4</b>	<b>C</b>
<b>CG13004</b>	<b>AT03755</b>	<b>92.9</b>	<b>7.1</b>	<b>0.0</b>	<b>0.0</b>	<b>28</b>	<b>7.1</b>	<b>D</b>
<b>CG2915</b>	<b>SD12095</b>	<b>36.7</b>	<b>10.0</b>	<b>53.3</b>	<b>0.0</b>	<b>30</b>	<b>116.6</b>	<b>A</b>
<b>CG9839</b>	<b>LD40589</b>	<b>100.0</b>	<b>0.0</b>	<b>0.0</b>	<b>0.0</b>	<b>30</b>	<b>0.0</b>	<b>D</b>
<b>CG4287</b>	<b>GH08991</b>	<b>81.8</b>	<b>18.2</b>	<b>0.0</b>	<b>0.0</b>	<b>22</b>	<b>18.2</b>	<b>D</b>
<b>apt</b>	<b>LD45581</b>	<b>86.8</b>	<b>0.0</b>	<b>13.2</b>	<b>0.0</b>	<b>76</b>	<b>26.4</b>	<b>C</b>
<b>PHDP</b>	<b>RE20490</b>	<b>78.6</b>	<b>10.7</b>	<b>3.6</b>	<b>7.1</b>	<b>28</b>	<b>39.2</b>	<b>C</b>
<b>CG12252</b>	<b>SD01014</b>	<b>100.0</b>	<b>0.0</b>	<b>0.0</b>	<b>0.0</b>	<b>26</b>	<b>0.0</b>	<b>D</b>

**Table 3. Summary of *in situ* hybridization**

CG9248	Genomic PCR	75.0	22.5	2.5	0.0	40	27.5	C
Sara	LD33044	93.3	0.0	6.7	0.0	30	13.4	D

\*Probe: cDNA clones used to generate complementary RNA probes for *in situ* hybridization. For genes with no EST clone, DNA template for *in vitro* transcription was directly generated by PCR from the genomic sequence (Genomic PCR). Existence-uncertain genes (CG14350 and CG16803) were not tested and excluded from the list. Kif3C was tested with a full-length EST clone.

† Percent distribution of signal intensities in MBs. Hybridization signals in MBs are scanned by confocal microscopy and categorized into four grades: (-), no signal; (±), weak; (+), moderate; (++), strong. Right and left hemispheres are independently assessed. Note that hybridization signals were amplified with tyramin and exhibited variation among brains.

‡ To evaluate expression levels and reflect the variation in signal intensities among samples, points were given based on signal intensities: zero for (-), one for (±), two for (+), and three for (++). Total point was calculated as the sum of (point x % value). Expression level of each gene was then ranked in four groups according to its total point. Rank A, ≥100; rank B, 50.0-99.9; rank C, 20.0-49.9; and rank D, 0-19.9. Because of signal variation, weak-to-moderate signals were occasionally detected even for D-rank genes. Conversely, negative brains were observed for A-rank genes at varying percentages.