- <sup>7</sup> Howie PW. The hemostatic mechanisms in pre-eclampsia. Clin Obstet Gynecol 1977;4:595-611.
- <sup>8</sup> Howie PW, Prentice CRM, McNicol GP. Coagulation, fibrinolysis and platelet function in pre-eclampsia, essential hypertension and placental insufficiency. *Journal of Obstetrics and Gynaecology of the British Commonwealth* 1971;**78**:992-1003.
- <sup>9</sup> Robertson WB, Brosens I, Dixon HG. The pathological response of the vessels of the placental bed to hypertensive pregnancy. *J Pathol Bacteriol* 1967;**93**:581-92.
- <sup>10</sup> Kitzmiller JL, Benirschke K. Immunofluorescent study of placental bed vessels in pre-eclampsia of pregnancy. Am J Obstet Gynecol 1973;115: 248-51.
- <sup>11</sup> Petrucco OM, Thomson NM, Lawrence JR, Weldon MW. Immunofluorescent studies in renal biopsies in pre-eclampsia. Br Med J 1974;i: 473-6.
- <sup>12</sup> Tribe CR, Smart GE, MacKenzie JC. Pre-eclampsia and the kidney. Br Med J 1974;ii:335.
- <sup>13</sup> Demers LM, Gabbe SG. Placental prostaglandin levels in pre-eclampsia. Am J Obstet Gynecol 1976;126:137-9.
- <sup>14</sup> Pipkin FB, Symonds EM. The renin-angiotensin system in the maternal and fetal circulation in pregnancy hypertension. *Clin Obstet Gynecol* 1977;4:651-64.

- <sup>15</sup> Scott JS, Jenkins DM, Need JA. Immunology of pre-eclampsia. Lancet 1978;i:704-6.
- <sup>16</sup> Petrucco OM, Seamark RF, Holmes K, Forbes IJ, Symonds RG. Changes in lymphocyte function during pregnancy. Br J Obstet Gynaecol 1976; 83:245-50.
- <sup>17</sup> Need JA, Jenkins DM, Scott JS. The response of lymphocytes to phytohaemagglutinin in women with pre-eclampsia. Br J Obstet Gynaecol 1976;83:438-40.
- <sup>18</sup> Jenkins DM, Need JA, Rajah SM. Deficiency of specific HLA antibodies in severe pregnancy pre-eclampsia/eclampsia. *Clin Exp Immunol* 1977; 27:485-6.
- <sup>19</sup> Jenkins DM, Need JA, Scott JS, Morris H, Pepper M. Human leucocyte antigens and mixed lymphocyte reaction in severe pre-eclampsia. Br Med J 1978;i:542-4.
- <sup>20</sup> Redman CWG, Bodmer JG, Bodmer WF, Beilin LJ, Bonnar J. HLA antigens in severe pre-eclampsia. *Lancet* 1978;ii:397-9.
- <sup>21</sup> Cooper DW, Liston WA. Genetic control of severe pre-eclampsia. J Med Genet 1979;16:409-16.
- <sup>22</sup> Feeney JG, Tovey LAD, Scott JS. Influence of previous blood-transfusion on incidence of pre-eclampsia. *Lancet* 1977;i:874-5.
- <sup>23</sup> Department of Health and Social Security. Report on confidential enquiries into maternal deaths in England and Wales 1973-1975. London: HMSO, 1979.

## Interferon: therapeutic fact or fiction for the '80s?

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Regular Review

For many years it had been known that infection with one virus could protect animals against infection with another when, in 1957, Isaacs and Lindenmann<sup>1</sup> discovered that medium from tissue cultures challenged with a killed virus could protect other cells against infection. The substances producing these effects are highly active glycoproteins known as interferons. They are released (in conjunction with many other unidentified molecules) from cells infected with virus or exposed to stimuli which mimic virus infection. Interferons probably act on cell membrane receptor sites causing intracellular production of proteins which mainly inhibit the translation of viral m-RNA.

Interferons are relatively species-specific, having maximal activity in cells from the same or closely related species. Furthermore, there are three major types of human interferon with different molecular structures and different physicochemical and antigenic properties. Preparations are arbitrarily defined by comparison with international standard preparations, and one unit of interferon is roughly the amount which reduces viral replication in tissue culture by half.

Sources of interferon—Most clinical studies have been performed using human leucocyte interferon (HuIFN $\alpha$ ). This is made by exposing pooled buffy-coat lymphocytes to a parainfluenza virus which may be inactivated later by acidification. The interferon thus obtained can be purified and concentrated by simple methods to 10<sup>6</sup>-10<sup>7</sup> units per ml/mg protein for clinical use.<sup>2</sup> The specific activity of pure human leucocyte interferon is about 10<sup>9</sup> U/mg protein, so clinical material is only about 0.1% pure. Most of the contaminating protein is probably albumin, but other proteins are present which may be biologically active. Interferon preparations have effects other than the inhibition of viral replication; for example, they inhibit cell growth and multiplication,<sup>3</sup> enhance the expression of cell-surface antigens,<sup>4</sup> suppress some functions of T and B lymphocytes,<sup>5</sup> and enhance the activity of natural killer cells.<sup>6</sup> <sup>7</sup> Any of these functions may be responsible for the antineoplastic activity<sup>8</sup> of interferons under current scrutiny.

Preliminary clinical studies have also been performed with interferon derived from human fibroblasts induced with a synthetic double-stranded RNA (poly I:C). The yields are enhanced by the judicious use of metabolic inhibitors which suppress synthesis initially of cellular protein and then of RNA.<sup>9</sup> Fibroblast interferon (HuIFN $\beta$ ) has also been purified to homogeneity (>10<sup>8</sup> U/mg protein), but the clinical material has about the same activity as clinical leucocyte interferon. Fibroblast interferon is less stable than human leucocyte interferon, and when injected intramuscularly achieves considerably lower blood concentrations of interferon,<sup>10</sup> probably because it is bound to tissue at the site of injection; whether this affects its therapeutic action is not known.

The antiviral activities of the two interferons have been compared directly against herpes simplex virus in monkeys' eyes,<sup>11</sup> in a clinical trial of herpes keratitis,<sup>12</sup> and against vaccinia in monkey skin.<sup>13</sup> Used topically or intradermally they do not appear to differ appreciably in efficacy. The major advantages of using fibroblasts over buffy-coat leucocytes as a source of interferon are that they may be well characterised and the production of interferon is limited only by resources for bulk cell cultivation. There are, however, technical problems in growing surface-adherent cells on a large enough scale to make production of interferon worth while, and as yet the manufacture of fibroblast interferon has provided sufficient material for only limited clinical studies.

Another approach<sup>14 15</sup> has been to grow lymphoblastoid cells in suspension cultures; in response to challenge with parainfluenza virus these produce mainly leucocyte and a small amount of fibroblast type interferons.<sup>16</sup> The cell lines which appear to produce interferons most efficiently came originally from patients with Burkitt's lymphoma, and each cell contains the genome for Epstein-Barr virus. All interferons for clinical use and the cells which produce them are extensively tested for safety. Though lymphoblastoid interferon is apparently free of virus and oncogenic activity, its use is being restricted to patients with life-threatening disease or neoplasia.

A third type of human interferon  $(HuIFN\delta)$  is derived from T lymphocytes by exposure to non-specific immune stimulants

or to antigens to which the lymphocytes have been sensitised.<sup>17</sup> This "immune" or "type II" interferon is produced early in acute viral infections and may be important in the initial host response. It is less thermostable and is inactivated at pH 2, distinguishing it from classical viral or type I interferon. Type II interferon has not yet been synthesised and purified in bulk for testing in clinical trials.

Finally, several groups are working on more exotic methods of interferon production: when an interferon protein molecule has been sequenced (and this has been achieved for parts of the molecules of fibroblast and lymphoblast interferons), maybe a small but highly active part of it could be synthesised. Alternatively, the genetic instruction for human interferon protein may be inserted into *Escherichia coli*, which could be grown on an almost unlimited scale; and several groups have now reported success with this technique (see *Nature* 1950;

## Details of clinical trials with exogenous interferon in virus diseases

Reference No	Virus	Year	Interferon type	Dose schedule	Result (effect of interferon compared with controls)
				As prophylaxis against virus disease	
18	Vaccinia	1962	Monkey kidney	Intradermal 10 <sup>3</sup> -10 <sup>4</sup> U/0·1 ml, 24 hours before	Protected skin sites
24	Vaccinia	1978	Human fibroblast	vaccination Intradermal $5 \times 10^4$ U/0·1 ml, 24 hours before vaccination	Protected skin sites
32 36 19	Respiratory virus Influenza Influenza	1965 1969 1973	Monkey kidney Human leucocyte Human leucocyte	Intranasal 10 <sup>4</sup> U/ml Intranasal 10 <sup>3</sup> -10 <sup>4</sup> U/ml during epidemic Intranasal 8 × 10 <sup>5</sup> U total on day before virus	No protection Reduced rate of infection Delay in symptoms and virus excretion
19	Rhinovirus	1973	Human leucocyte	challenge Intranasal $1.4 \times 10^6$ U total in four days, virus challenge on second day	Significant reduction in cold severity, virus
39	Herpes simplex virus keratitis	1976	Human leucocyte	Eye-drops $6.4 \times 10^6$ U/ml twice daily to prevent recurrence	excretion No reduction in rate of recurrence
37	Herpes simplex virus keratitis	1976	Human leucocyte	Eye-drops $11-31 \times 10^6$ U/ml daily after debridement	Reduced rate of local recurrence of dendritic ulcer
62 27	Rubella Herpes labialis	1975 1979	Human leucocyte Human leucocyte	Intramuscular $6.5 \times 10^5$ U twice before vaccination Intramuscular $7 \times 10^4$ U/kg/day for five days, trigeminal surgery on second day	Delay in virus excretion and antibody production Reduced shedding of herpes simplex virus and
25	Renal transplantation	1975	Human fibroblast	Intramuscular $3 \times 10^6$ U twice weekly for three months	cold sores No reduction in rate of virus infections
26	Renal transplantation	1979	Human leucocyte	Intramuscular $3 \times 10^6$ U/day for six weeks	Delay in cytomegalovirus viraemia. No effect on herpes simplex virus
38	Herpes simplex virus	1976	Human leucocyte	Eye-drops $6.3 \times 10^4$ U/ml three times a day after	No effect on rate of healing
20	keratitis Herpes simplex virus		Human leucocyte	thermocautery Eye-drops $2 \times 10^6$ U/ml three times a day after	Significantly accelerated healing
12	keratitis Herpes simplex virus	1978	Human fibroblast	thermocautery Eye-drops 10 <sup>6</sup> U/ml once daily	No difference between interferons in efficacy
21	keratitis Herpes simplex virus	1978	or leucocyte Human leucocyte	Eye-drops $10^6$ or $30 \times 10^6$ U/ml daily with	Higher dose accelerated healing
42 43 22	keratitis Herpes zoster Herpes zoster Herpes zoster	1975 1978 1978	Human leucocyte Human leucocyte Human leucocyte	trifluorothymidine Intramuscular 10 <sup>6</sup> U/day for five days (not blind) Intramuscular 8 × 10 <sup>4</sup> -5 × 10 <sup>6</sup> U/kg/day Intramuscular 4 × 10 <sup>4</sup> -5 × 10 <sup>6</sup> U/kg/day	Faster lesion crusting and resolution of lesion pain Non-significant reduction in rate of complications Highest dose prevented spread and reduced
				In treatment of chronic virus infection	complications
44	Genital warts	1975	Human leucocyte	Cream 4000 $U/g$ applied five times daily (females)	Resolution of warts in all treated patients by 12 weeks
45 53	Genital warts Hepatitis B chronic active hepatitis	1979 1980	Human fibroblast Human leucocyte	Intralesional 300 U/0·1 ml (males) Intramuscular $12 \times 10^6$ U daily for seven days, then reducing	Minor but significant reduction in wart growth Transient fall in HBV-DNAP (also in one control)
			Uncontroll	ed observations in treatment of chronic viral disease	
46	Hepatitis B chronic active hepatitis	1976	Human fibroblast	Intramuscular 107 U on alternate days, seven doses (one patient)	Reduction in nucleocapsid HBcAg staining on biopsy
47	Hepatitis B chronic active hepatitis	1976	Human leucocyte	Intramuscular $6-170 \times 10^3$ U/kg/day, variable courses (four patients)	Fall in HBV-DNAP (and in HBsAg later)
48	Hepatitis B chronic active hepatitis	1978	Human leucocyte	Intramuscular $6 \times 10^3$ U/kg/day (13 patients)	Permanent fall in HBV-DNAP in three patients (two patients cleared HBsAg)
49	Hepatitis B chronic active hepatitis	1978	Human fibroblast	Intramuscular 10 <sup>7</sup> U daily for 14 days (two patients)	Fall in serum HBcAb titre
50	Hepatitis B chronic active hepatitis	1977	Human fibroblast	Intramuscular $3 \times 10^6$ U daily for 14 days (two patients)	No effect. HBV-DNAP fell in one untreated control
51	Hepatitis B chronic active hepatitis	1979	Human fibroblast	Intramuscular 10 <sup>6</sup> U daily for 82 days (one patient)	Fall in HBsAg, HBcAg, HBV-DNAP
52	Hepatitis B chronic active hepatitis	1979	Human leucocyte	Intramuscular 10 <sup>6</sup> U daily (four patients)	Fall in DNAP and eAg, improved liver function tests, and liver biopsy appearances (two patients) Temporary fall in HBV-DNAP, improved liver
53	Hepatitis B chronic active hepatitis	1979	Human leucocyte	Intramuscular 2-5 × 10 <sup>4</sup> U/kg/day for 5 weeks to 5 months (seven patients)	biopsy appearances in one patient. eAg cleared in two
54 55	Cytomegalovirus Cytomegalovirus	1966 1976	Human leucocyte Human leucocyte	Intramuscular about $10^3$ U/ml (four infants) Intramuscular $2.5 \times 10^{5}$ - $10^{6}$ U daily (variable courses)	No effect Viruria cleared in two out of four patients with congenital and one out of five with acquired viruria
56	Cytomegalovirus	1976	Human leucocyte	Intramuscular 1·7-3·5 × 10 <sup>s</sup> U/kg/day for 7-14 days (six patients)	Transient suppression of viruria in one patient
57	Cytomegalovirus	1976	Human leucocyte	Intramuscular 10 <sup>6</sup> U daily (three bone-marrow recipients)	Apparent benefit with decreased viraemia in one
58	Chronic rubella	1976	Human leucocyte	Intramuscular $3 \times 10^6$ U daily for 14 days (one patient)	Regression of acute vasculitis, resolution of viraemia

HBV-DNAP = Hepatitis - B - virus DNA polymerase. HBcAg = Hepatitis B core antigen. HBsAg = Hepatitis B surface antigen. HBcAb = Hepatitis B core antibody. eAg = e Antigens.

**283**:323). Interferon protein produced in this way will have to be separated from the products of bacterial fermentation, and if the carbohydrate component of this interferon is necessary for stability or activity, glycosylation of the molecule will need to be performed after fermentation.

Clinical evaluation of interferon and inducers against infections-Clinical studies with interferon have gone through three phases. Apart from a well-controlled study published in 1962,18 which showed that interferon prepared from monkey kidney cells and given intradermally could protect human skin sites against subsequent vaccination, the early studies (1957-66) using relatively low potency material were either uncontrolled or gave negative results. As difficulties became apparent in manufacturing the large amounts of interferon which appeared to be necessary attention turned to developing and testing a range of interferon inducers. While these were active in studies on animals, they were generally unsuccessful in clinical trials. In the '70s, interest in exogenous interferon was revived by the pioneering effort of Mogensen and Cantell, who used buffy coats from Finnish Red Cross blood units for the mass production of human leucocyte interferon.<sup>2</sup> Research with this material and smaller quantities of fibroblast interferon from several groups has been concentrated on infections with herpes viruses, respiratory viruses, and hepatitis B virus. Production limitations have restricted the numbers of clinical trials of interferon: the important ones are summarised in the table. Several poorly controlled or inconclusive studies are excluded from this list, but uncontrolled observations on the effect of interferon in chronic viral infections are included. When well-conducted studies have failed this has generally been attributed to the use of too little interferon-a view confirmed in some diseases by the success of later studies with higher dose schedules.<sup>19-22</sup> Nevertheless, an important caveat to these studies (and even more so to the trials in cancer) must be that the preparations given were composed almost entirely of material other than interferon and some of the effects may have been due to impurities.

Study of the interferon system in relation to natural or experimental virus infections has indicated the clinical circumstances in which exogenous interferon could be used.<sup>23</sup> Studies on animals have shown that interferon is most active when given before or with the virus challenge.

Interferon in prophylaxis-Studies of the clinical use of interferon in prophylaxis have given conflicting results. Whereas skin sites can be protected against challenge with vaccinia virus by intradermal inoculations of interferon,18 24 fibroblast interferon given by intramuscular injection three times a week to recipients of renal transplants failed to reduce the frequency of naturally acquired virus infections.<sup>25</sup> A more recent study employing daily human leucocyte interferon delayed the onset of cytomegalovirus viraemia in seropositive transplant recipients.<sup>26</sup> There was no effect on the reactivation of herpes simplex virus infection nor on the eventual outcome of transplantation in these patients. Reactivation of viral illness may also be expected (though there is no immunosuppression) after surgery to the trigeminal ganglia for tic douloureux, and in one study<sup>27</sup> leucocyte interferon given from one day before operation for five days reduced both the frequency and duration of shedding of herpes simplex virus from the oropharynx. Herpes labialis also occurred less often in the patients treated with interferon.

*Respiratory diseases*—Interferon may be detected in the nasal washings and serum of volunteers infected with influenza.<sup>28 29</sup> Peak concentrations of interferon in nasal washings occur around the time of the fall in virus shedding

before antibodies are found, suggesting a cause-and-effect relation. Whether or not interferon has a key role in limiting acute virus infections of man is still not clear. In some diseases, such as respiratory syncytial virus infection in children, interferon is rarely detectable in serum or nasal washings,<sup>30</sup> virus excretion is protracted, and the fall in the virus titre is associated with a rise in concentration of virusspecific IgA antibody,<sup>31</sup> suggesting that resolution of the disease (at least in children in hospital) is not entirely mediated by endogenous interferon.

In another study human leucocyte interferon  $(14 \times 10^7 \text{ U})$ total) given as frequent nasal sprays over four days significantly reduced symptoms and shedding of virus after an experimental challenge with rhinovirus on the second day of treatment with interferon.<sup>19</sup> Material of lower potency has proved ineffective.<sup>19 32</sup> While interferon,<sup>33</sup> like any intranasal liquid,<sup>34</sup> has a short half life in the nasal cavity, it needs to be in contact with nasal epithelial cells for a long period to make them resistant to viral infection.<sup>35</sup> It is therefore difficult to understand how, in Russia, nose drops containing a low titre of human leucocyte interferon could cause the degree of protection indicated by the results of a large clinical study on children during an influenza epidemic.36 Experiments are needed to find a method of application of interferon that ensures prolonged contact with the mucosa and to find how late in the course of acute respiratory infection exogenous interferon may be given and still have a clinical effect.

Viral diseases of the eye-Eye-drops containing concentrated human leucocyte interferon  $(11-31 \times 10^6 \text{ U/ml})$  have been shown to reduce the rate of recurrence of herpetic dendritic ulcers after minimal wiping debridement.<sup>37</sup> Dilute eye-drops  $(6.25 \times 10^4 \text{ U/ml})$  given after thermocautery for herpetic keratitis were no more effective than placebo in accelerating healing or reducing viral shedding.38 More concentrated interferon (10<sup>6</sup> U/ml), however, was effective in a further similar trial.<sup>20</sup> Without local physical treatment, topical interferon given at low concentrations seems an inefficient treatment of herpetic eye disease, but highly concentrated interferon has not yet been evaluated alone. Nevertheless, the combination of high-dose interferon  $(30 \times 10^6 \text{ U/ml})$  with trifluorothymidine (an antiherpetic nucleoside analogue, not generally available in Britain), enhanced the rapid healing of herpetic keratitis.<sup>21</sup> In contrast, regular treatment over several months with low-potency ( $6.4 \times 10^4$  U/ml) eye-drops did not prevent recurrent herpetic keratitis when compared with placebo<sup>39</sup>; this study is being repeated with more-concentrated material. Both experiments on animals<sup>40</sup> and these clinical studies show that the concentration of interferon eyedrops is crucial and that the efficacy of low-titre material cannot be enhanced by giving it more often. A recent controlled study in Israel<sup>41</sup> of fibroblast interferon against placebo in epidemic adenovirus conjunctivitis has shown promising results.

Herpes zoster—Three controlled trials have been published of the treatment of herpes zoster infection with intramuscular human leucocyte interferon. The first was not double blind and the placebo-treated group was small,<sup>42</sup> but the proportion of patients with persisting pain in the dermatome beyond 15 days was greater in those given placebo than in those treated with interferon, in whom crusting of the lesions also occurred earlier. One group of workers has examined the effect of interferon on herpes zoster infection complicating neoplastic diseases under placebo-controlled double-blind conditions. In children<sup>43</sup> there was a trend towards fewer complications of infection in the interferon-treated group. In the adults<sup>22</sup> increasing dose schedules were studied and the highest  $(5\cdot1\times10^5~U/kg/day)$  inhibited the formation of fresh vesicles in the primary dermatome, prevented dissemination, and reduced complications, including postherpetic neuralgia.

*Chronic viral infections*—A human leucocyte interferon cream used for several weeks cured vulval warts,<sup>44</sup> while intralesional injections of very small amounts of fibroblast interferon into penile warts had a small but statistically significant inhibitory effect on their growth.<sup>45</sup>

Chronic infections with, for example, rubella, cytomegalovirus, herpes simplex virus, or hepatitis B virus may be influenced by treatment with interferon. Since, however, large numbers of cells may be infected in these conditions, the effect might be expected to wear off when treatment was stopped unless the infected cells had been destroyed. This may explain why, in several uncontrolled studies, high doses of both human leucocyte and fibroblast interferons only temporarily reduced the circulating markers of viral infection in chronic active hepatitis associated with hepatitis B virus.46-53 Infection may apparently be abolished by treatment in a few of these patients, but in most the fall in viral markers is only temporary. One recently reported placebo-controlled trial showed that leucocyte interferon could consistently lower circulating hepatitis-B-virus-associated DNA polymerase in eight patients but that the effect was only temporary.<sup>54</sup> One patient in the placebo group had spontaneous clearing of detectable DNA polymerase three weeks after completing treatment. Spontaneous cure may occur occasionally in chronic active hepatitis. The beneficial effects seen with interferon in some patients may have been due to an antiviral effect or to an effect on the immune system. Further work is needed to identify the patients who may respond to treatment with interferon and to evaluate the efficacy of higher dose schedules or combination treatment with other promising antiviral agents such as adenine arabinoside.48

Chronic infection with cytomegalovirus has also been treated<sup>55-58</sup>; virus was cleared from the urine in a few patients (four out of 25 in published reports), and the virus titre in the urine fell temporarily in some of the remainder. Whether a more favourable result could be obtained with higher doses of interferon is not yet known, but it is not surprising that prophylactic treatment at the doses used did not reverse reactivated infection<sup>58</sup> when higher doses had only a minor delaying effect.<sup>26</sup> In one child with congenitally acquired chronic rubella,<sup>59</sup> infection was no longer detectable after a two-week course of interferon.

*Toxicity*—Larger doses of interferon have not been used clinically for two reasons: insufficient supplies and dose-related (but reversible) toxicity. The major unwanted effects (with both leucocyte and fibroblast interferons) appear to be bone-marrow suppression, an inhibition of growth in young children,<sup>57</sup> and temporary loss of hair.<sup>60</sup> Mild side effects (fever, headache, myalgia, and malaise) tend to wear off after about a week's continuous treatment and do not occur in patients on corticosteroids.<sup>26</sup>

Interferon and cancer—Mouse interferon has a beneficial effect on some experimental tumours in mice.<sup>61</sup> When human leucocyte interferon was given for 18 months to patients with osteogenic sarcoma who had received standard primary treatment interim analysis showed that they had survived longer without metastases than patients treated concurrently at other centres. Because the design of that trial is open to criticism, control groups should be chosen very carefully in future studies. In uncontrolled studies some remarkable responses have been reported in the size of the tumour in small numbers of patients with juvenile laryngeal papilloma, multiple myeloma, metastasising breast tumours, non-Hodgkin's lymphoma, and some leukaemias.<sup>62</sup> These apparent benefits may have been due, at least in part, to non-specific effects. Placebo-controlled trials are needed to prove without doubt that interferon has activity, with long-term follow-up to evaluate the possible benefit to treated patients.

The temptation to prescribe new agents (particularly new antiviral and antineoplastic drugs) on an uncontrolled basis often seems overwhelming but early favourable results may prevent later, adequate evaluation. So far with interferon a remarkable amount of knowledge has been gained using small amounts of material in well-controlled studies. These have shown that human interferon has activity in some established viral illnesses (notably herpetic eye disease, herpes zoster in malignancy, and genital warts). If hopes for interferon are fulfilled it may become widely available in perhaps 10 years' time. Present supplies should be concentrated in further controlled studies aimed at discovering how best to use it and deciding who most needs it. Many years seem likely to elapse before interferon is used (in the West) to treat common colds.

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- <sup>1</sup> Isaacs A, Lindenmann J. Virus interference. 1. The interferon. Proceedings of the Royal Society; Series B: Biological Sciences 1957;147:258-67.
- <sup>2</sup> Mogensen KE, Cantell K. Production and preparation of human leukocyte interferon. *Pharmacol Ther* 1977;1:369-81.
- <sup>3</sup> Strander H. Anti-tumour effects of interferon and its possible use as an anti-neoplastic agent in man. *Tex Rep Biol Med* 1977;**35**:429-35.
- <sup>4</sup> Lindahl P, Gresser I, Leary P, Tovey M. Enhanced expression of histocompatibility antigens of lymphoid cells treated with interferon. *J Infect Dis* 1976;133, suppl:A66-8.
   <sup>5</sup> Brodeur BR, Merigan TC. Mechanism of the suppressive effect of
- <sup>o</sup> Brodeur BR, Merigan TC. Mechanism of the suppressive effect of interferon on antibody synthesis in vivo. J Immunol 1975;114:1323-8.
- <sup>6</sup> Einhorn S, Blomgren H, Strander H. Interferon and spontaneous cytotoxicity in man. 1. Enhancement of the spontaneous cytotoxicity of peripheral lymphocytes by human leukocyte interferon. Int J Cancer 1978;22:405-12.
- <sup>7</sup> Zarling JM, Eskra L, Borden EC, Horoszewicz J, Carter WA. Activation of human natural killer cells cytotoxic for human leukaemia cells by purified human interferon. *J Immunol* 1979;**123**:63-70.
- <sup>8</sup> Gresser I, Tovey MG. Antitumour effects of interferon. *Biochim Biophys* Acta 1978;516:231-47.
- <sup>9</sup> Havell EA, Vilček J. Production of high-titred interferon in cultures of human diploid cells Antimicrob Agents Chemother 1972;2:476-84.
   <sup>10</sup> Billiau A, De Somer P, Edy VG, De Clercq E, Heremans H. Human
- <sup>10</sup> Billiau A, De Somer P, Edy VG, De Clercq E, Heremans H. Human fibroblast interferon for clinical trials: pharmacokinetics and tolerability in experimental animals and humans. *Antimicrob Agents Chemother* 1979;16:56-63.
- <sup>11</sup> Neumann-Haefelin D, Sundmacher R, Skoda R, Cantell K. Comparative evaluation of human leukocyte and fibroblast interferon in the prevention of herpes simplex virus keratitis in a monkey model. *Infect Immun* 1977;**17**:468-70.
- <sup>12</sup> Sundmacher R, Cantell K, Skoda R, Hallermann C, Neumann-Haefelin D. Human leucocyte and fibroblast interferon in a combination therapy of dendritic keratitis. *Albrecht von Graefes Arch Klin Ophthalmol* 1978; 208:229-33.
- <sup>13</sup> Scott GM, Cartwright T, Tyrrell DAJ, Butler JK, Porteous M, Stevens RM. Preliminary experience with fibroblast interferon against vaccinia virus in human and monkey skin. In: Ikić D, ed. Proceedings of the symposium on preparation, standardisation, clinical use of interferon. Zagreb: Yugoslav Academy of Sciences and Arts, 1977:135-42.
   <sup>14</sup> Bridgen PJ, Anfinsen CB, Corley L, et al. Human lymphoblastoid
- <sup>14</sup> Bridgen PJ, Anfinsen CB, Corley L, et al. Human lymphoblastoid interferon: large scale production and partial purification. J Biol Chem 1977;252:6585-7.
- <sup>15</sup> Finter NB. Large scale production of lymphoblastoid interferon. In: National Organising Committee, ed. International virology IV. Wageningen: Centre for Agricultural Publiching and Decumeratering. 1079-00.
- ingen: Centre for Agricultural Publishing and Documentation, 1978:98.
  <sup>16</sup> Havell EA, Yip YK, Vilček J. Characteristics of human lymphoblastoid (Namalva) interferon. *J Gen Virol* 1977;**38**:51-9.

- <sup>17</sup> Epstein LB. Effects of interferon on the immune response in vitro and in vivo. In: Stewart WE II, ed. *Interferons and their actions*. Cleveland, Ohio: CRC Press Inc, 1977:91-132.
- <sup>18</sup> A report to the Medical Research Council from the Scientific Committee on Interferon. Effect of interferon on vaccination in volunteers. *Lancet* 1962;i:873-5.
- <sup>19</sup> Merigan TC, Reed SE, Hall TS, Tyrrell DAJ. Inhibition of respiratory virus infection by locally applied interferon. *Lancet* 1973;i:563-7.
- <sup>20</sup> Sundmacher R, Neumann-Haefelin D, Cantell K. Successful treatment of dendritic keratitis with human leukocyte interferon. A controlled clinical study. *Albrecht von Graefes Arch Klin Ophthalmol* 1976;201: 39-45.
- <sup>21</sup> Sundmacher R, Cantell K, Neumann-Haefelin D. Combination therapy of dendritic keratitis with trifluorothymidine and interferon. *Lancet* 1978;ji:687.
- <sup>22</sup> Merigan TC, Rand KH, Pollard RB, Abdallah PS, Jordan GW, Fried RP. Human leukocyte interferon for the treatment of herpes zoster in patients with cancer. N Engl J Med 1978;298:981-7.
- <sup>23</sup> Stewart WE II. Antiviral actions of interferons in animals. The interferon system. Wien, New York: Springer Verlag, 1979:266-91.
- <sup>24</sup> Scott GM, Cartwright T, LeDu G, Dicker D. Effect of human fibroblast interferon on vaccination in volunteers. *J Biol Stand* 1978;**6**:73-6.
- <sup>25</sup> Weimar W, Schellekens H, Lameijer LDF, et al. Double-blind study of interferon administration in renal transplants. Eur J Clin Invest 1978; 8:255-8.
- <sup>26</sup> Cheeseman SH, Rubin RH, Stewart JA, et al. Controlled clinical trial of prophylactic human-leukocyte interferon in renal transplantation: effects on cytomegalovirus and herpes simplex virus infections. N Engl J Med 1979;300:1345-9.
- <sup>27</sup> Pazin GJ, Armstrong JA, Lam MT, Tarr GC, Jannetta PJ, Ho M. Prevention of reactivated herpes simplex infection by human leukocyte interferon after operation on the trigeminal root. N Engl J Med 1979; 301:225-30.
- <sup>28</sup> Gresser I, Dull HB. A virus inhibitor in pharyngeal washings from patients with influenz. Proc Soc Exp Biol Med 1964;115:192-6.
- <sup>29</sup> Jao RL, Wheelock EF, Jackson GG. Production of interferon in volunteers infected with Asian influenza. J Infect Dis 1970;121:419-26.
- <sup>30</sup> Hall CB, Douglas RG, Simons RL, Geiman JM. Interferon production in children with respiratory syncytial, influenza and parainfluenza virus infections. *J Pediatr* 1978;93:28-32.
- <sup>31</sup> McIntosh K. Interferon in nasal secretions from infants with viral respiratory infections. J Pediatr 1978;93:33-6.
- <sup>32</sup> A report to the Medical Research Council from the Scientific Committee on Interferon. Experiments with interferon in man. *Lancet* 1965;i: 505-6.
- <sup>33</sup> Johnson PE, Greenberg SB, Harmon MW, Alford BR, Couch RB. Recovery of applied human leucocyte interferon from the nasal mucosa of chimpanzees and humans. *J Clin Microbiol* 1976;4:106-7.
- <sup>34</sup> Aoki FY, Crawley JCW. Distribution and removal of human serum albumin-technetium 99m instilled intranasally. Br J Clin Pharmacol 1976;3:869-78.
- <sup>35</sup> Greenberg SB, Harmon MW, Johnson PE, Couch RB. Antiviral activity of intranasally applied human leukocyte interferon. Antimicrob Agents Chemother 1978;14:596-600.
- <sup>36</sup> Solov'ev VD. The results of controlled observations on the prophylaxis of influenza with interferon. Bull WHO 1969;41:683-8.
- <sup>37</sup> Jones BR, Coster DJ, Falcon MG, Cantell K. Topical therapy of ulcerative herpetic keratitis with human interferon. *Lancet* 1976;ii:128.
- <sup>38</sup> Sundmacher R, Neumann-Haefelin D, Manthey KF, Müller O. Interferon in treatment of dendritic keratitis in humans: a preliminary report. *J Infect Dis* 1976;**133**, suppl:A160-4.
- <sup>39</sup> Kaufman HE, Meyer RF, Laibson PR, Waltman SR, Nesburn AB, Shuster JJ. Human leukocyte interferon for the prevention of recurrences of herpetic keratitis. *J Infect Dis* 1976;133, suppl:A165-8.
- <sup>40</sup> McGill JI, Collins P, Cantell K, Jones BR, Finter NB. Optimal schedules for the use of interferon in the corneas of rabbits with herpes simplex keratitis. *J Infect Dis* 1976;**133**, suppl:A13-7.

28 JUNE 1980

<sup>41</sup> Revel M. Interferon treatment of adenovirus eye infection. 2nd international workshop on interferons, New York, April 1979. In press.

BRITISH MEDICAL JOURNAL

- <sup>42</sup> Emödi G, Rufli T, Just M, Hernandez R. Human interferon therapy for herpes zoster in adults. Scand J Infect Dis 1975;7:1-5.
- <sup>43</sup> Arvin AM, Feldman S, Merigan TC. Human leukocyte interferon in the treatment of varicella in children with cancer: a preliminary controlled trial. Antimicrob Agents Chemother 1978;13:605-7.
- <sup>44</sup> Ikić D, Bosnić N, Smerdel S, Jusić D, Soos E, Delimar N. Double blindclinical study with human leukocyte interferon in the therapy of condylomata accuminata. In: Ikić D, ed. *Proceedings of a symposium on* the clinical use of interferon. Zagreb: Yugoslav Academy of Sciences and Arts, 1975:229-33.
- <sup>45</sup> Scott GM, Csonka GW. Effect of injections of small doses of human fibroblast interferon into genital warts. Br *J Vener Dis* 1979;55:442-5.
- <sup>46</sup> Desmyter J, Ray MB, Degroote J, et al. Administration of human fibroblast interferon in chronic hepatitis-B infection. Lancet 1976;ii: 645-7.
- <sup>47</sup> Greenberg HB, Pollard RB, Lutwick LI, Gregory PB, Robinson WS, Merigan TC. Effect of human leucocyte interferon on hepatitis B virus infection in patients with chronic active hepatitis. N Engl J Med 1976;295:517-22.
- <sup>48</sup> Merigan TC, Robinson WS, Gregory PB, et al. Antiviral therapy in hepatitis Binfection. In National Organising Committee, ed. International virology IV. Wageningen: Centre for Agricultural Publishing and Documentation, 1978:125.
- <sup>49</sup> Kingham JGC, Ganguly NK, Shaari ZD, et al. K treatment of HBsAgpositive chronic active hepatitis with human fibroblast interferon. Gut 1978;19:91-4.
- <sup>50</sup> Weimar W, Heijtink RA, Schalm SW, et al. Fibroblast interferon in HBsAg-positive chronic active hepatitis. Lancet 1977;ii:1282.
- <sup>51</sup> Dolen JG, Carter WA, Horoszewicz JZ, Vladutiu AO, Leibowitz AI, Nolan JP. Fibroblast interferon treatment of a patient with chronic active hepatitis. Increased number of circulating T lymphocytes and elimination of rosette-inhibitory factor. Am J Med 1979;67:127-31.
- <sup>52</sup> Kato Y, Kobayashi K, Suyama T, Hattori N. Effects of human leucocyte interferon therapy on hepatitis B virus in patients with chronic active hepatitis. *Gastroenterology* 1979;**77**:A21.
- <sup>53</sup> Scullard GH, Alberti A, Wansbrough-Jones MH, et al. Effects of human leucocyte interferon on hapatitis B virus replication and immune responses in patients with chronic hepatitis B infection. J Clin Lab Immunol 1979;1:277-82.
- <sup>54</sup> Weimar W, Heijtink RA, Ten Kate FJP, et al. Double-blind study of leucocyte interferon administration in chronic HBsAg-positive hepatitis. Lancet 1980;i:336-8.
- <sup>55</sup> Falcoff E, Falcoff R, Fournier F, Chany C. Production en masse, purification partielle et caractérisation d'un interféron destiné à des essais thérapeutiques humains. Annales de l'Institut Pasteur 1966;111:562-84.
- <sup>56</sup> Emödi G, O'Reilly R, Müller A, Everson LK, Binswanger U, Just M. Effect of human exogenous leukocyte interferon in cytomegalovirus infections. *J Infect Dis* 1976;**133**, suppl:A199-204.
- <sup>57</sup> Arvin AM, Yeager AS, Merigan TC. Effect of leukocyte interferon on urinary excretion of cytomegalovirus by infants. *J Infect Dis* 1976; 133, suppl:A205-10.
- <sup>58</sup> O'Reilly RJ, Everson LK, Emödi G, et al. Effects of exogenous interferon in cytomegalovirus infections complicating bone marrow transplantation. *Clin Immunol Immunopathol* 1976;**6**:51-61.
- <sup>59</sup> Larsson A, Forsgren M, Hard AF, Segerstad S, Strander H, Cantell K. Administration of interferon to an infant with congenital rubella syndrome involving persistent viremia and cutaneous vasculitis. Acta Paediatr Scand 1976;65:105-10.
- <sup>60</sup> Merigan TC, Sikora K, Breeden JH, Levy R, Rosenberg SA. Preliminary observations on the effect of interferon in non-Hodgkin's lymphoma. N Engl J Med 1978;299:1449-53.
- <sup>61</sup> Gresser I. Antitumour effects of interferon. Adv Cancer Res 1972;16: 97-140.
- <sup>62</sup> Anonymous. Can interferons cure cancers? Lancet 1979;i:1171-2.
- <sup>63</sup> Best JM, Banatvala JE. The effect of a human interferon preparation on vaccine-induced rubella infection. J Biol Stand 1975;3:107-12.