

Survey of In Vitro Susceptibilities of *Vibrio cholerae* O1 and O139 to Antimicrobial Agents

TATSUO YAMAMOTO,^{1*} G. BALAKRISH NAIR,² M. JOHN ALBERT,³
CARLOS CARRILLO PARODI,⁴ AND YOSHIFUMI TAKEDA⁵

Department of Bacteriology, School of Medicine, Juntendo University,¹ and Research Institute, International Medical Center of Japan,⁵ Tokyo, Japan; National Institute of Cholera and Enteric Diseases, Calcutta, India²; International Center for Diarrheal Disease Research, Bangladesh, Dhaka, Bangladesh³; and National Institute of Health, Lima, Peru⁴

Received 17 March 1994/Returned for modification 16 June 1994/Accepted 29 September 1994

***Vibrio cholerae* O139 (173 strains) and O1 (221 strains) were tested for their in vitro susceptibilities to 39 antimicrobial agents. Both O139 and O1 strains were highly susceptible to azithromycin, cepheids, minocycline, penems, and newer fluoroquinolones. O139 strains (94.8%), O1 Indian El Tor strains (97%), and Bangladeshi El Tor strains (50%) were highly resistant to streptomycin, sulfamethoxazole, and trimethoprim and moderately resistant to chloramphenicol and furazolidone, in sharp contrast to O1 Peruvian El Tor and O1 classical strains. Some Bangladeshi El Tor strains (43.3%) showed tetracycline resistance as well.**

Vibrio cholerae O1 biotype El Tor, the causative agent of the seventh cholera pandemic (6), was first recorded in 1961 in Indonesia and continues to possess the capacity of causing cholera epidemics. For instance, in 1991, El Tor strains appeared in South and Central America, starting in Peru, where no large cholera epidemics had been recorded in this century (31, 32). In 1982, the classical biotype of *V. cholerae* O1 (the causative agent of the previous cholera pandemics) reappeared in Bangladesh and continues to persist (16). Large explosive cholera epidemics due to *V. cholerae* non-O1 strains that elaborate cholera toxin occurred in India and Bangladesh from October 1992 (1, 10, 24, 33). *V. cholerae* O139 Bengal, which is currently classified as the epidemic non-O1 strain, has spread to neighboring countries (e.g., Thailand [4], Pakistan [7], and Singapore [5]), and imported cases associated with *V. cholerae* O139 have now been reported from the United States (28), Japan (17), and Switzerland (3).

In addition to being resistant to streptomycin (24) and the vibriostatic agent O/129 (1), *V. cholerae* O139 strains are usually resistant to sulfamethoxazole-trimethoprim (co-trimoxazole) (1, 24, 28) and to furazolidone (24); the last two are often recommended for the treatment of cholera (9, 23). Moreover, most of the *V. cholerae* O1 El Tor strains in Bangladesh are resistant to tetracycline (1). In this study, we investigated the in vitro susceptibilities of *V. cholerae* O139 strains to 39 antimicrobial agents and compared them with those obtained for *V. cholerae* O1 strains belonging to the classical and El Tor biotypes.

All *V. cholerae* strains used in this study were of clinical origin. The 173 O139 strains examined were isolated during the period 1992 to 1993 and included 134 strains from India, 30 from Bangladesh, and 9 from Thailand. Of the 72 O1 classical strains examined, 66 were isolated from 1960 to 1965 in India while 6 were isolated in 1982 in Bangladesh. A total of 149 O1 El Tor strains examined in this study included 67 from India isolated from 1985 to 1990, 52 from Peru isolated in 1991, and

30 from Bangladesh isolated in 1994. Isolates were stored frozen at -80°C .

The antimicrobial agents were gifts from their manufacturers. Amoxicillin was used in combination with clavulanic acid (a β -lactamase inhibitor). Penems used were FCE22101 (8) and SY5555 (11a) (alternatively named SUN5555 [21], ALP201 [2a], and WY-49,605). Azithromycin is a 15-membered ring macrolide (25). Newer quinolones included BAYy3118 (2) and DU6859a (27). Furazolidone, 3-(5-nitrofurfurylideneamino)-2-oxazolidinone, was also used. Sulfamethoxazole and trimethoprim were used alone and in combination at ratios of 5:1 (as in the combination drug) and 20:1 (the expected ratio in the human body). O/129 (2,4-diamino-6,7-diisopropylpteridine phosphate) was purchased from Sigma Chemical, St. Louis, Mo.

Susceptibility testing of bacterial strains was done by the agar dilution method with Mueller-Hinton agar (Difco Laboratories, Detroit, Mich.) according to standard procedures (14, 26). The final concentrations of antimicrobial agents were from 0.004 to 128 $\mu\text{g/ml}$. The test bacteria were grown for 18 h at 37°C with agitation in L broth (18) and diluted to approximately 10^6 CFU/ml. Aliquots of the bacterial suspension (approximately 10^4 CFU per spot) were inoculated on the surface of antimicrobial agent-containing agar plates. Incubation was for 20 h at 37°C . The MIC was determined as previously described (14, 26). *Escherichia coli* NIHJ JC-2 was used as a reference strain for quality control (12, 14, 19, 20, 22). When the susceptibility to sulfamethoxazole or trimethoprim was tested, Mueller-Hinton agar supplemented with 7.5% (vol/vol) defibrinated horse blood (frozen and thawed) was also used, in addition to Mueller-Hinton agar alone (13).

The MICs of the antimicrobial agents against clinical isolates of O139 are summarized in Table 1. Among antimicrobial agents tested, the newer quinolones (norfloxacin, ofloxacin, tosfloxacin, ciprofloxacin, sparfloxacin, BAYy3118, and DU6859a) showed the greatest activity (MICs, ≤ 0.06 $\mu\text{g/ml}$). Six (I-2, I-7, I-14, I-15, I-64, and I-72) of the 173 O139 strains (3.5%) were resistant to ampicillin (MIC, ≥ 256 $\mu\text{g/ml}$), tetracycline (MIC, 8 to 16 $\mu\text{g/ml}$), chloramphenicol (MIC, 32 $\mu\text{g/ml}$), kanamycin (MIC, ≥ 256 $\mu\text{g/ml}$), and gentamicin (MIC, 128 to ≥ 256 $\mu\text{g/ml}$). With the exception of one strain (B20), the remaining 172

* Corresponding author. Mailing address: Department of Bacteriology, School of Medicine, Juntendo University, 2-1-1 Hongo, Bunkyo-ku, Tokyo, Japan. Fax: 81-3-3814-9300.

TABLE 1. MICs of antimicrobial agents for 173 clinical isolates of *V. cholerae* O139

Antimicrobial agent	MIC ($\mu\text{g/ml}$)		
	50%	90%	Range
Penicillins			
Ampicillin	2	4	2– ≥ 256
Amoxicillin-clavulanic acid (2:1)	8	8	4–32
Penems			
FCE22101	2	2	1–8
SY5555	1	2	1–4
Broad-spectrum cepheps			
Cefoperazone	0.06	0.12	0.03–16
Cefixime	0.06	0.06	0.03–1
Tetracyclines			
Tetracycline	0.25	0.5	0.25–16
Doxycycline	0.25	0.25	0.12–2
Minocycline	0.12	0.12	0.06–0.25
Chloramphenicol	8	8	1–32
Aminoglycosides			
Streptomycin	≥ 256	≥ 256	8– ≥ 256
Kanamycin	8	8	4– ≥ 256
Gentamicin	1	2	0.25– ≥ 256
Macrolides			
Spiramycin	128	128	64–128
Oleandomycin	64	128	64–128
Midecamycin	32	32	16–64
Josamycin	16	16	8–32
Rokitamycin	16	16	8–32
Kitasamycin	16	16	8–32
Roxithromycin	8	16	4–32
Clarithromycin	8	8	4–16
Erythromycin	4	8	1–8
Azithromycin	0.5	0.5	0.12–1
Lincomycin	≥ 256	≥ 256	64– ≥ 256
Clindamycin	64	64	16–64
Nalidixic acid (older class of quinolones)	0.25	0.5	0.12–16
Others			
Polymyxin B	128	128	8– ≥ 256
Colistin	≥ 256	≥ 256	≥ 256
SMX ^a	≥ 256	≥ 256	8– ≥ 256
TMP ^b	≥ 256	≥ 256	0.5– ≥ 256
SMX-TMP (20:1)	≥ 256	≥ 256	1– ≥ 256
SMX-TMP (5:1)	≥ 256	≥ 256	0.5– ≥ 256
Furazolidone	4	8	0.12–8
O/129	≥ 256	≥ 256	4– ≥ 256

^a SMX, sulfamethoxazole.^b TMP, trimethoprim.

strains were all highly resistant to streptomycin (MIC, ≥ 64 $\mu\text{g/ml}$), sulfamethoxazole (MIC, ≥ 256 $\mu\text{g/ml}$), trimethoprim (MIC, ≥ 128 $\mu\text{g/ml}$), and O/129 (MIC, ≥ 256 $\mu\text{g/ml}$).

The MICs at which 50% of the isolates were inhibited (MIC₅₀s) and MIC₉₀s of the antimicrobial agents for O1 clinical isolates were very similar to the MIC₅₀s and MIC₉₀s for O139 (Table 1), except those of tetracycline, chloramphenicol,

furazolidone, streptomycin, sulfamethoxazole, trimethoprim, and O/129. One strain (0.5%) belonging to the classical biotype was resistant to ampicillin (MIC, 64 $\mu\text{g/ml}$), tetracycline (MIC, 8 $\mu\text{g/ml}$), chloramphenicol (MIC, 32 $\mu\text{g/ml}$), kanamycin (MIC, ≥ 256 $\mu\text{g/ml}$), streptomycin (MIC, ≥ 256 $\mu\text{g/ml}$), sulfamethoxazole (MIC, ≥ 256 $\mu\text{g/ml}$), trimethoprim (MIC, ≥ 256 $\mu\text{g/ml}$), and O/129 (MIC, ≥ 256 $\mu\text{g/ml}$).

Subtypes of O139 and O1 strains showing different susceptibility patterns (with respect to tetracycline, chloramphenicol, furazolidone, streptomycin, sulfamethoxazole, trimethoprim, and O/129) are summarized in Table 2. The major susceptibility patterns of O139 strains (94.8%) and O1 Indian El Tor strains (97%) and major susceptibility pattern A of O1 Bangladeshi El Tor strains (50%) were indistinguishable from each other. Those resistance phenotypes (MICs [in micrograms per milliliter] of 4 to 8 [chloramphenicol], 2 to 8 [furazolidone], 64 to ≥ 256 [streptomycin], 128 to ≥ 256 [sulfamethoxazole], 128 to ≥ 256 [trimethoprim], and ≥ 256 [O/129]) were designated VC MAR1 (for *V. cholerae* multiple-antimicrobial-agent resistance). Major susceptibility pattern B of O1 Bangladeshi El Tor strains (43.3%) added tetracycline resistance (MIC, 8 $\mu\text{g/ml}$) to the VC MAR1 phenotype; this resistance phenotype was designated VC MAR2.

The major susceptibility pattern of O1 Peruvian El Tor strains (98.1%) was distinctly different from the major susceptibility pattern of O1 Indian El Tor strains (97%) (VC MAR1) but was quite similar to minor pattern B of O1 Indian El Tor strains (1.5%). The major susceptibility pattern of O1 Peruvian El Tor strains (98.1%) also resembled the major susceptibility pattern of O1 classical strains (94.4%).

Plasmids of *V. cholerae* strains were isolated and electrophoresed in 0.3 or 0.7% agarose with reference plasmid DNAs of known molecular size (including the 94.5-kb NR1 plasmid [30]) as previously described (15, 34). Six strains of *V. cholerae* O139 (I-2, I-7, I-14, I-15, I-64, and I-72) had plasmids with a molecular size of ca. 200 kb. These 200-kb plasmids encoded resistance to ampicillin, tetracycline, chloramphenicol, kanamycin, gentamicin, sulfamethoxazole, trimethoprim, and O/129 (34a). In contrast, two O139 strains and two O1 Indian El Tor strains with the VC MAR1 phenotype (susceptible to tetracycline, ampicillin, and gentamicin) and six O1 Bangladeshi El Tor strains with the VC MAR2 phenotype tested had no detectable plasmids. Thus far, no correlation has been found between plasmids and the VC MAR phenotypes.

The incidence of tetracycline-resistant O1 El Tor strains in Bangladesh was 1.9% (number of strains tested, 317) in 1990, 7.6% (number of strains tested, 1,377) in 1991, 61.1% (number of strains tested, 1,221) in 1992, and 85.4% (number of strains tested, 669) in 1993; the data were obtained with antibiotic discs. In contrast, in India, tetracycline resistance is not frequently found in O1 El Tor strains (data not shown). Thus, there are apparent differences in tetracycline susceptibility which are related to the geographic origin within the Asian subcontinent. The tetracycline-resistant strains found in this study were all susceptible to minocycline (MIC, 0.06 to 0.25 $\mu\text{g/ml}$).

Cefoperazone resistance was observed for some strains of *V. cholerae* O139 (2.3%) (MIC, 8 to 16 $\mu\text{g/ml}$ [Table 1]) and *V. cholerae* O1 (1.4%) (MIC, 8 to 128 $\mu\text{g/ml}$). *V. cholerae* O139 and O1 also tended to be resistant to nalidixic acid, with resistance rates of 4% (MIC, 4 to 16 $\mu\text{g/ml}$) for O139 (Table 1) and 1.4% (MIC, 4 to 8 $\mu\text{g/ml}$) for O1. Such resistant strains may have been selected by use of the related antimicrobial agents.

Azithromycin was eightfold more active against both O139 and O1 strains than was erythromycin. Since erythromycin has

TABLE 2. Subtypes of *V. cholerae* O139 and O1 showing different MIC patterns

<i>V. cholerae</i> strain	Susceptibility pattern (%)	MIC ($\mu\text{g/ml}$) ^a of:						
		TC	CM	FZ	SM	SMX	TMP	O/129
O139 ^b	Major (94.8)	0.25–0.5	4–8	2–8	64–\geq256	\geq 256	128–\geq256	\geq 256
	Minor A (1.2)	0.25	8	0.12–0.25	128–\geq256	\geq 256	\geq 256	\geq 256
	Minor B (0.6)	0.25	1	4	8	8	0.5	4
El Tor, Bangladesh	Major A (50)	0.25–0.5	8	8	128–\geq256	\geq 256	\geq 256	\geq 256
	Major B (43.3)	8	8	8	128–\geq256	\geq 256	\geq 256	\geq 256
	Minor A (3.3)	0.25	1	0.25	16	\geq 256	0.5	4
	Minor B (3.3)	0.25	1	8	128	\geq 256	0.5	4
El Tor, India	Major (97)	0.25	4–8	4–8	128–\geq256	\geq 256	\geq 256	\geq 256
	Minor A (1.5)	0.25	8	8	8	\geq 256	\geq 256	\geq 256
	Minor B (1.5)	0.25	1	0.25	16	8	0.5	4
El Tor, Peru	Major (98.1)	0.25	1	0.12–0.5	8–16	4–8	0.25–0.5	2–4
	Minor A (1.9)	0.25	1	0.25	8	\geq 256	0.25	2
Classical ^c	Major (94.4)	0.12–0.5	0.5–1	0.06–0.25	4–32	0.25–16	0.12–0.5	0.5–4
	Minor A (1.4)	0.25	1	4	16	8	0.5	4
	Minor B (1.4)	0.25	1	0.25	\geq 256	1	0.25	2
	Minor C (1.4)	0.25	0.5	0.12	8	128	0.5	2

^a Abbreviations for antimicrobial agents: TC, tetracycline; CM, chloramphenicol; FZ, furazolidone, SM, streptomycin; SMX, sulfamethoxazole; TMP, trimethoprim. Higher MICs with each antimicrobial agent are shown in boldface type; higher versus lower MIC ranges are 8 versus 0.12 to 0.5 (tetracycline), 4 to 8 versus 0.5 to 1 (chloramphenicol), 2 to 8 versus 0.06 to 0.5 (furazolidone), 64 to \geq 256 versus 4 to 32 (streptomycin), 128 to \geq 256 versus 0.25 to 16 (sulfamethoxazole), 128 to \geq 256 versus 0.12 to 0.5 (trimethoprim), and \geq 256 versus 0.5 to 4 (O/129).

^b Six multiple-drug-resistant strains (strains I-2, I-7, I-14, I-15, I-64, and I-72 [3.5%]; described in the text) were omitted. The MICs (in micrograms per milliliter) of chloramphenicol, furazolidone, streptomycin, sulfamethoxazole, trimethoprim, and O/129 for the six strains were 32, 4 to 8, \geq 256, \geq 256, \geq 256, and \geq 256, respectively.

^c One multiple-drug-resistant strain (1.4%; described in the text) was omitted. The MICs (in micrograms per milliliter) of chloramphenicol, furazolidone, streptomycin, sulfamethoxazole, trimethoprim, and O/129 for this strain were 32, 0.25, \geq 256, \geq 256, \geq 256, and \geq 256, respectively; the MIC of furazolidone was similar to the MIC of furazolidone for the major type (or minor types B and C).

a good safety record (29) and azithromycin has also been shown to be safe in children (11), azithromycin would be a potential, attractive chemotherapeutic agent of choice in treatment of *V. cholerae* O139 and O1 infections. Newer quinolones were extremely active against both O139 and O1 strains. However, newer quinolones have not usually been recommended in the treatment of cholera in the pediatric age group.

We thank Hisao Kurazono for bacterial strains. This study was supported by a grant-in-aid for scientific research from the Ministry of Education, Science, and Culture of Japan and a grant from the Department of Health and Welfare of Japan.

REFERENCES

- Albert, M. J., A. K. Siddique, M. S. Islam, A. S. G. Faruque, M. Anaruz-zaman, S. M. Faruque, and R. B. Sack. 1993. Large outbreak of clinical cholera due to *Vibrio cholerae* non-O1 in Bangladesh. *Lancet* **341**:704.
- Bauernfeind, A. 1993. Comparative in-vitro activities of the new quinolone, Bay y 3118, and ciprofloxacin, sparfloxacin, tosofloxacin, CI-960 and CI-990. *J. Antimicrob. Chemother.* **31**:505–522.
- Bergan, T., and J. da Fonseca. 1991. Comparative antibacterial activity of the penem ALP201. *Chemotherapy* **37**:413–419.
- Bundesamt für Gesundheitswesen. 1993. *Vibrio cholerae* der Serogruppe O139: ein neuer Choleraerreger mit epidemischem Potential im indischen Subkontinent. *Bull. Bundesamtes Gesundheitswesen* **38**:676–678.
- Chongsa-nguan, M., W. Chaicumpa, P. Moolasart, P. Kandhasingha, T. Shimada, H. Kurazono, and Y. Takeda. 1993. *Vibrio cholerae* O139 Bengal in Bangkok. *Lancet* **342**:430–431.
- Committee on Epidemic Diseases. 1993. Cholera. *Epidemiol. News Bull.* **19**:59.
- Finkelstein, R. A. 1973. Cholera. *Crit. Rev. Microbiol.* **2**:553–623.
- Fisher-Hoch, S. P., A. Khan, I. Haq, M. A. Khan, and E. D. Mintz. 1993. *Vibrio cholerae* O139 in Karachi, Pakistan. *Lancet* **342**:1422–1423.
- Franceschi, G., E. Perrone, M. Alpegiani, A. Bedeschi, C. Battistini, F. Zarini, and C. D. Bruna. 1989. Synthesis and antimicrobial spectrum of FCE 22101 and its orally available ester FCE 22891. *J. Antimicrob. Chemother.* **23**(Suppl. C):1–6.
- Francis, T. I., E. A. Lewis, A. B. O. O. Oyediran, O. A. Okubadejo, D. Montefiore, I. I. Onyewotu, I. Mohammed, E. A. Ayoola, and R. Vincent. 1971. Effect of chemotherapy on the duration of diarrhoea, and on vibrio excretion by cholera patients. *J. Trop. Med. Hyg.* **74**:172–176.
- Hall, R. H., F. M. Khambaty, M. Kothary, and S. P. Keasler. 1993. Non-O1 *Vibrio cholerae*. *Lancet* **342**:430.
- Hopkins, S. 1993. Clinical safety and tolerance to azithromycin in children. *J. Antimicrob. Chemother.* **31**(Suppl. E):111–117.
- Inoue, E., and S. Mitsuhashi. 1994. In vitro antibacterial activity and β -lactamase stability of SY5555, a new oral penem antibiotic. *Antimicrob. Agents Chemother.* **38**:1974–1979.
- Ishiguro, M., H. Iwata, T. Nakatsuka, R. Tanaka, Y. Maeda, T. Nishihara, T. Noguchi, and T. Nishino. 1988. Studies on penem antibiotics. 1. Synthesis and in vitro activity of novel 2-chiral substituted penems. *J. Antibiot. (Tokyo)* **41**:1685–1693.
- Japan Society of Chemotherapy. 1973. Committee report: methods of sensitivity testing for sulfamethoxazole-trimethoprim combination product. *Chemotherapy (Tokyo)* **21**:67–76.
- Japan Society of Chemotherapy. 1981. Committee report. *Chemotherapy (Tokyo)* **29**:76–79.
- Kado, C. I., and S. T. Liu. 1981. Rapid procedure for detection and isolation of large and small plasmids. *J. Bacteriol.* **145**:1365–1373.
- Khan, M. U., A. R. Samadi, M. I. Huq, and W. B. Greenough. 1986. Reappearance of classical *Vibrio cholerae* in Bangladesh, p. 3–12. *In* S. Kuwahara and N. F. Pierce (ed.), *Advances in research on cholera and related diarrheas*, vol. 3. KTK Scientific Publishers, Tokyo.
- Kurazono, T., F. Yamada, M. Yamaguchi, Y. Ohzeki, Y. Okuyama, K. Itoh, and T. Shimada. 1994. The first report of traveler's diarrhea associated with a newly described toxigenic *Vibrio cholerae* O139 strain in Japan. *J. Jpn. Assoc. Infect. Dis.* **68**:8–12. (In Japanese.)
- Lennox, E. S. 1955. Transduction of linked genetic characters of the host by bacteriophage P1. *Virology* **1**:190–206.
- Mitsuhashi, S., and S. Takagi. 1991. In vitro antibacterial activity of FCE22101 and its stability to β -lactamases, p. 13–39. *In* S. Mitsuhashi and G. Franceschi (ed.), *Penem antibiotics*. Japan Scientific Societies Press, Tokyo.
- Nakamura, S., A. Minami, K. Nakata, N. Kurobe, K. Kouno, Y. Sakaguchi, S. Kashimoto, H. Yoshida, T. Kojima, T. Ohue, K. Fujimoto, M. Nakamura,

- M. Hashimoto, and M. Shimizu. 1989. In vitro and in vivo antibacterial activities of AT-4140, a new broad-spectrum quinolone. *Antimicrob. Agents Chemother.* **33**:1167-1173.
21. Nishino, T., Y. Maeda, E. Ohtsu, S. Koizuka, T. Nishihara, H. Adachi, K. Okamoto, and M. Ishiguro. 1989. Studies of penem antibiotics. II. *In vitro* activity of SUN5555, a new oral penem. *J. Antibiot. (Tokyo)* **42**:977-988.
 22. Ono, T., K. Numata, M. Inoue, and S. Mitsuhashi. 1988. Bacteriological evaluation of TE-031 (A-56268), a new macrolide antibiotic: in vitro and in vivo antibacterial activity. *Chemotherapy (Tokyo)* **36**:1-34.
 23. Pierce, N. F., J. G. Banwell, R. C. Mitra, G. J. Caranasos, R. I. Keimowitz, J. Thomas, and A. Mondal. 1968. Controlled comparison of tetracycline and furazolidone in cholera. *Br. Med. J.* **3**:277-280.
 24. Ramamurthy, T., S. Garg, R. Sharma, S. K. Bhattacharya, G. B. Nair, T. Shimada, T. Takeda, T. Karasawa, H. Kurazono, A. Pal, and Y. Takeda. 1993. Emergence of novel strain of *Vibrio cholerae* with epidemic potential in southern and eastern India. *Lancet* **341**:703-704.
 25. Retsema, J., A. Girard, W. Schelkly, M. Manousos, M. Anderson, G. Bright, R. Borovoy, L. Brennan, and R. Mason. 1987. Spectrum and mode of action of azithromycin (CP-62,993), a new 15-membered-ring macrolide with improved potency against gram-negative organisms. *Antimicrob. Agents Chemother.* **31**:1939-1947.
 26. Sahm, D. F., and J. A. Washington II. 1991. Antibacterial susceptibility tests: dilution methods, p. 1105-1116. *In* A. Balows, W. J. Hausler, Jr., K. L. Herrmann, H. D. Isenberg, and H. J. Shadomy (ed.), *Manual of clinical microbiology*, 5th ed. American Society for Microbiology, Washington, D.C.
 27. Sato, K., K. Hoshino, M. Tanaka, I. Hayakawa, and Y. Osada. 1992. Antimicrobial activity of DU-6859, a new potent fluoroquinolone, against clinical isolates. *Antimicrob. Agents Chemother.* **36**:1491-1498.
 28. Tormey, M., L. Mascola, L. Kilman, P. Nagami, E. DeBess, S. Abbott, and G. W. Rutherford III. 1993. Imported cholera associated with a newly described toxigenic *Vibrio cholerae* O139 strain. *Morbid. Mortal. Weekly Rep.* **42**:501-503.
 29. Washington, J. A., and W. R. Wilson. 1985. Erythromycin: a microbial and clinical perspective after 30 years of clinical use. *Mayo Clin. Proc.* **60**:271-278.
 30. Womble, D. D., and R. H. Rownd. 1988. Genetic and physical map of plasmid NR1: comparison with other IncFII antibiotic resistance plasmids. *Microbiol. Rev.* **52**:433-451.
 31. World Health Organization. 1992. Cholera in the Americas. *Weekly Epidemiol. Rec.* **67**:33-40.
 32. World Health Organization. 1992. Cholera in 1991. *Weekly Epidemiol. Rec.* **67**:253-260.
 33. World Health Organization. 1993. Epidemic diarrhoea due to *Vibrio cholerae* non-O1. *Weekly Epidemiol. Rec.* **68**:141-148.
 34. Yamamoto, T., Y. Koyama, M. Matsumoto, E. Sonoda, S. Nakayama, M. Uchimura, W. Paveenkittiporn, K. Tamura, T. Yokota, and P. Echeverria. 1992. Localized, aggregative, and diffuse adherence to HeLa cells, plastic, and human small intestines by *Escherichia coli* isolated from patients with diarrhea. *J. Infect. Dis.* **166**:1295-1310.
 - 34a. Yamamoto, T., G. B. Nair, and Y. Takeda. Unpublished data.