## Canadian National Survey of Prevalence of Antimicrobial Resistance among Clinical Isolates of *Streptococcus pneumoniae*

ANDREW E. SIMOR,<sup>1,2\*</sup> MARIE LOUIE,<sup>1,2</sup> THE CANADIAN BACTERIAL SURVEILLANCE NETWORK,<sup>†</sup> AND DONALD E. LOW<sup>2,3</sup>

> Departments of Microbiology, Sunnybrook Health Science Centre,<sup>1</sup> Mount Sinai Hospital,<sup>3</sup> and University of Toronto,<sup>2</sup> Toronto, Ontario, Canada

Received 20 December 1995/Returned for modification 17 April 1996/Accepted 25 June 1996

The antimicrobial susceptibilities of 1,089 clinical isolates of *Streptococcus pneumoniae* obtained from 39 laboratories across Canada between October 1994 and August 1995 were determined. A total of 91 isolates (8.4%) demonstrated intermediate resistance (MIC, 0.1 to 1.0  $\mu$ g/ml) and 36 (3.3%) had high-level resistance (MIC,  $\geq 2.0 \ \mu$ g/ml) to penicillin. Penicillin-resistant strains were more likely to have been recovered from normally sterile sites (P = 0.005) and to be cross-resistant to several β-lactam and non-β-lactam antimicrobial agents (P < 0.05). These results indicate that there has been a recent significant increase in the prevalence of antibiotic-resistant *S. pneumoniae* in Canada.

In the past few years, the incidence of invasive infections such as bacteremia and meningitis due to Streptococcus pneu*moniae* has been rising (2, 6). These infections continue to be associated with considerable morbidity and mortality. There has also been a recent worldwide increase in the incidence of pneumococcal resistance to several antimicrobial agents, including penicillin and other  $\beta$ -lactams (1, 3, 4, 7, 10, 12, 14). Until very recently, resistant pneumococci were infrequently identified in Canada. Three large Canadian surveys of pneumococcal susceptibility in the 1970s and 1980s found rates of resistance to penicillin of 2.4, 1.3, and 1.5% in Alberta (5), Quebec (13), and Ontario (15), respectively. Furthermore, only intermediate resistance to penicillin (MIC, 0.1 to 1.0 µg/ ml) had been detected; high-level resistance to penicillin (MIC,  $\geq 2.0 \,\mu$ g/ml) was not detected in any of these studies. An increase in penicillin resistance was first noted in southern Ontario in late 1993 (20). Therefore, we set out to determine the prevalence of antimicrobial resistance among clinical isolates of S. pneumoniae in Canada.

A total of 30 hospital-based and 9 privately owned community microbiology laboratories from across Canada participated in the study by submitting consecutive isolates of *S. pneumoniae* recovered from patients between October 1994 and August 1995. Only single isolates from different patients were included in the study. Identification of isolates as *S. pneumoniae* was confirmed by using standard techniques, including Gram stain characteristics, colonial morphology, ethylhydrocuprein susceptibility, bile solubility, and the Pneumoslide Test (Becton Dickinson Microbiology Systems, Cockeysville, Md.). Pneumococcal isolates were serotyped by the National Reference Centre for Streptococcus (Edmonton, Alberta, Canada) on the basis of capsular swelling with type-specific antisera.

All isolates were stored frozen at  $-70^{\circ}$ C. Prior to susceptibility testing, isolates were thawed and subcultured onto blood agar twice. In vitro susceptibility testing was done by a broth microdilution procedure in accordance with National Committee for Clinical Laboratory Standards guidelines (16, 17), by using cation-adjusted Mueller-Hinton broth (Becton Dickinson Microbiology Systems) with 5% lysed horse blood and an inoculum of 10<sup>5</sup> CFU/ml. MICs (defined as the lowest concentration of antibiotic that completely inhibited visible growth) were determined after incubation for 20 to 24 h at 35°C in ambient air. Streptococcus pneumoniae ATCC 6303 (penicillin susceptible), S. pneumoniae ATCC 49619 (penicillin resistant), Staphylococcus aureus ATCC 29213, and Enterococcus faecalis ATCC 29212 were used as control strains. The 17 antimicrobial agents tested (penicillin G, cefuroxime, cefotaxime, ceftriaxone, ceftazidime, cefpirome, cefpodoxime, imipenem, erythromycin, clindamycin, tetracycline, chloramphenicol, ciprofloxacin, ofloxacin, levofloxacin, trimethoprim-sulfamethoxazole,

 TABLE 1. Sources of 1,089 S. pneumoniae isolates collected across

 Canada from October 1994 to August 1995

	•							
Specimen origin (no.)	No. (%) of isolates							
	Penicillin susceptible	Intermediately or highly resistant to penicillin						
Sterile sites (120)	96 (80.0)	$24 (20.0)^a$						
Blood (97)	79 ` ´	18						
Other (23)	17	6						
Nonsterile sites (913)	817 (89.5)	96 $(10.5)^a$						
Upper respiratory tract (539)	492	47						
Lower respiratory tract (365)	316	49						
Other (9)	9	0						
Not known (56)	49	7						
Total	962 (88.3)	127 (11.7)						

 $^{a}P = 0.005$  for sterile-site isolates versus nonsterile-site isolates.

<sup>\*</sup> Corresponding author. Mailing address: Department of Microbiology, B121, Sunnybrook Health Science Centre, 2075 Bayview Ave., North York, Ontario, Canada M4N 3M5. Phone: (416) 480-4549. Fax: (416) 480-6845.

<sup>&</sup>lt;sup>†</sup> Members of the Canadian Bacterial Surveillance Network include L. Abbott, Queen Elizabeth Hospital, Charlottetown, Prince Edward Island; J. Blondeau, St. Paul's Hospital, Saskatoon, Saskatchewan; C. Brown, S. Hanson, and Associates, Edmonton, Alberta; K. Forward, Victoria General Hospital, Halifax, Nova Scotia; D. Hoban, Health Sciences Centre, Winnipeg, Manitoba; P. Kibsey, Royal Jubilee Hospital, Victoria, British Columbia; M. Kuhn, The Moncton Hospital, Moncton, New Brunswick; M. Laverdiere, Hospital Maisonneuve-Rosemont, Montreal, Quebec; T. Louie, Calgary General Hospital, Calgary, Alberta; and R. Szumski, Calgary Medical Laboratories, Calgary, Alberta.

## TABLE 2. Antimicrobial susceptibilities of 962 penicillin-susceptible, 91 intermediately penicillin-resistant, and 36 highly penicillin-resistant S. pneumoniae isolates collected across Canada from October 1994 to August 1995

Antimicrobial agent	Penicillin MIC		No. of isolates for which the MIC ( $\mu$ g/ml) of penicillin was:									
	(µg/ml)	0.03	0.06	0.12	0.25	0.5	1.0	2.0	4.0	8.0	16	32
Penicillin	<0.12 0.12–1.0 ≥2.0	866	96	40	21	8	22	26	10			
Cefuroxime <sup>a</sup>	<0.12 0.12-1.0 $\ge 2.0$		651	297 5	8 30	5 23	1 13	$10 \\ 3$	7 12	2 20	1 1	
Cefpodoxime <sup>b</sup>	<0.12 0.12-1.0 $\ge 2.0$		935 15	21 14	2 25	1 19 2	3 12 5	4 10	2 18	1		
Cefotaxime <sup>a</sup>	<0.12 0.12-1.0 $\ge 2.0$	869 10	70 12	19 25 1	4 17 3	20 4	6 18	$1 \\ 10$				
Ceftriaxone <sup>a</sup>	<0.12 0.12-1.0 $\ge 2.0$	869 4	87 21	5 14	1 27	19 8	5 20	1 8				
Ceftazidime <sup>b</sup>	<0.12 0.12-1.0 $\ge 2.0$	61	40	347 11	470 3	36 26	2 21	5 10 1	1 13 10	7 25		
Cefpirome <sup>b</sup>	<0.12 0.12-1.0 $\ge 2.0$	915 34	41 6	3 31	2 10 7	1 9 20	1 9					
Imipenem <sup>a</sup>	<0.12 0.12-1.0 $\ge 2.0$	958 51	3 12	1 8 12	19 14	1 9	1					
Erythromycin <sup>a</sup>	<0.12 0.12-1.0 $\ge 2.0$			906	36 80 27	4 2 3	3	2 2 3	5 4 2	4 1	2 1	1 1
Clindamycin <sup><i>a</i></sup>	<0.12 0.12-1.0 $\ge 2.0$				954 86 32				3 2	3 1	2 1 1	1 3
Tetracycline <sup><i>a</i></sup>	<0.12 0.12-1.0 $\ge 2.0$							944 72 27		2 1 2	14 5 3	2 13 4
Chloramphenicol <sup>a</sup>	<0.12 0.12–1.0 ≥2.0							902 74 29	55 12	2 2 3	3 3 4	
Ciprofloxacin <sup>b</sup>	<0.12 0.12–1.0 ≥2.0			94	21 4 1	571 22 9	253 55 20	16 12 4	4 1 1	3 2 1		
Ofloxacin	<0.12 0.12-1.0 $\ge 2.0$				7 1	60 2 1	673 19 8	219 66 24	1 2 1	2 1 2		
Levofloxacin <sup>b</sup>	<0.12 0.12-1.0 $\ge 2.0$			38	19 1	626 31 12	272 56 20	5 1 3	1	2 1 1		
Trimethoprim-sulfamethoxazole <sup>a</sup>	<0.12 0.12-1.0 $\ge 2.0$				658 20	187 22 1	39 5 1	48 7 1	12 6 3	12 10 7	6 18 21	3 2
Vancomycin	<0.12 0.12–1.0 >2.0					962 91 36						

 ${}^{a} p \le 0.05$  for antimicrobial resistance of penicillin-susceptible versus penicillin-resistant strains.  ${}^{b}$  No current National Committee for Clinical Laboratory Standards-recommended criteria for intermediate and high-level resistance categories.

and vancomycin) were supplied by the respective manufacturers.

The chi square test or Fisher's exact test was used, as appropriate, to determine the significance of differences between groups.

A total of 1,089 clinical isolates of *S. pneumoniae* were examined. The number of isolates contributed by each of the 39 participating laboratories varied from a low of 11 to a maximum of 124 isolates, with a mean of 30 isolates per laboratory. A total of 443 isolates (41%) were obtained from children under the age of 16 years, whereas 502 isolates (46%) were from those 16 years of age or older. The age of the patient was unknown for 144 (13%) of the isolates.

Reduced penicillin susceptibility was detected in 127 (11.7%) of the isolates: 91 (8.4%) with intermediate resistance and 36 (3.3%) with high-level resistance. Resistance (MIC,  $\geq 2.0 \ \mu g/ml$ ) to cefotaxime or ceftriaxone was detected in 1% of isolates; 2% of isolates had intermediate susceptibility (MIC =  $1.0 \ \mu g/ml$ ). Resistance to these extended-spectrum cephalosporins occurred only in strains with reduced susceptibility to penicillin. Antimicrobial resistance rates varied across the country, increasing from east to west. In three Atlantic provinces (Prince Edward Island, New Brunswick, and Nova Scotia), 13 (7.4%) of 176 isolates were resistant to penicillin. Resistance rates increased to 10.3% (58 of 565) for isolates recovered in central Canada (Quebec, Ontario, and Manitoba), and to 16.1% (56 of 348) for isolates from Western Canada (Saskatchewan, Alberta, and British Columbia) (P =0.001). As shown in Table 1, 20% of 120 sterile-site isolates of S. pneumoniae were resistant to penicillin, compared with 10.5% of 913 nonsterile-site isolates (P = 0.005).

The in vitro activities of the antimicrobial agents tested against the 1,089 study isolates categorized by penicillin susceptibility are summarized in Table 2. Penicillin-susceptible strains of *S. pneumoniae* were generally also susceptible to the other antimicrobial agents tested, whereas penicillin-resistant strains were much more likely to be resistant to other antibiotics. For example, all penicillin-susceptible pneumococcal isolates were also susceptible to the other  $\beta$ -lactam agents tested. However, 100, 28, and 22% of the strains with high-level penicillin resistance were resistant to cefuroxime, cefotaxime, and ceftriaxone, respectively. Resistance to many of the non- $\beta$ -lactam antibiotics was also seen much more frequently in penicillin-resistant isolates (Table 2). Isolates were uniformly susceptible only to vancomycin.

Forty-six randomly selected penicillin-resistant pneumococcal strains were serotyped and found to belong to eight different serotypes: 9V (11 strains), 6B (9 strains), 23F (8 strains), 19A (7 strains), 19F (6 strains), 14 (3 strains), 6A (1 strain), and 21 (1 strain). There was no apparent geographic clustering of any particular serotype.

The results of this investigation confirm that the prevalence of antimicrobial resistance in clinical isolates of *S. pneumoniae* has increased dramatically in Canada in just a few years. Although resistance was detected in all parts of the country, the prevalence of resistance was not uniformly distributed, with a trend for somewhat higher resistance rates in the western prairie provinces. A diversity of serotypes was present, including several which are included in the currently available polyvalent pneumococcal vaccine.

Pneumococcal strains from normally sterile sites (blood, cerebrospinal fluid, pleural fluid, etc.) were almost twice as likely to be resistant to penicillin as were isolates from nonsterile sites (P = 0.005), confirming the importance of drug resistance in invasive *S. pneumoniae* infections (12). Slightly more than one-quarter of the resistant isolates demonstrated high-level resistance (MIC,  $\geq 2.0 \ \mu g/ml$ ) to penicillin. Not surprisingly, since the mechanism of resistance is related to altered penicillin-binding proteins with reduced affinity for  $\beta$ -lactam drugs (9, 11), these strains are also more likely to have reduced susceptibility to other  $\beta$ -lactams, including extended-spectrum cephalosporins. As previously described (1, 12, 14, 18), increased resistance to other classes of antimicrobial agents, such as macrolides, tetracyclines, chloramphenicol, and trimethoprimsulfamethoxazole, was also detected in the current study.

Limited clinical experience and the lack of prospective controlled trials have hindered the development of recommendations for the management of pneumococcal infection due to penicillin-resistant strains. The treatment of otitis media is hampered by the uncertainty of adequate antibiotic penetration into middle ear effusions (18). For bacterial meningitis that may be due to penicillin-resistant S. pneumoniae, recommended empiric treatment is either with ceftriaxone plus vancomycin or with cefotaxime plus vancomycin (8, 19). However, vancomycin therapy may be suboptimal because its penetration into cerebrospinal fluid is variable and treatment failure with vancomycin therapy has been reported (22). Treatment with high doses of cefotaxime has been used successfully in a small number of adult patients with meningitis due to S. pneumoniae with decreased susceptibilities to penicillin and extended-spectrum cephalosporins (21). Further studies are required to determine whether treatment with newer investigational compounds or combinations of antimicrobial agents would be beneficial.

The recent increase in the prevalence of penicillin- and cephalosporin-resistant *S. pneumoniae* as a cause of serious and life-threatening infection in Canada and the United States emphasizes the need for continued surveillance to determine local antimicrobial susceptibility data and to identify changing patterns of resistance. These data are essential for developing appropriate guidelines for empiric treatment of pneumococcal infections. Greater use of pneumococcal vaccine, especially for those at high risk of developing severe pneumococcal infections, should also be promoted while ongoing efforts are directed at the development of a new conjugate vaccine with improved efficacy.

We thank L. Louie and S. Matsumura for technical assistance; Marguerite Lovgren, National Centre for Streptococcus, for serotyping isolates; and L. Cook for secretarial services.

This study was financially supported, in part, by the Canadian Bacterial Diseases Network and by Hoffmann-La Roche Limited and Hoechst Marion Roussel Canada Inc.

## REFERENCES

- Applebaum, P. C. 1992. Antimicrobial resistance in *Streptococcus pneu-moniae*: an overview. Clin. Infect. Dis. 15:77–83.
- Baer, M., R. Vuento, and T. Vesikari. 1995. Increase in bacteraemic pneumococcal infections in children. Lancet 345:661. (Letter.)
- Barry, A. L., M. A. Pfaller, P. C. Fuchs, and R. R. Packer. 1994. In vitro activities of 12 orally administered antimicrobial agents against four species of bacterial respiratory tract pathogens from U.S. medical centers in 1992 and 1993. Antimicrob. Agents Chemother. 38:2419–2425.
- Breiman, R. F., J. C. Butler, F. C. Tenover, J. A. Elliott, and R. R. Facklam. 1994. Emergence of drug-resistant pneumococcal infections in the United States. JAMA 271:1831–1835.
- Dixon, J. M. S., A. E. Lipinski, and M. E. P. Graham. 1977. Detection and prevalence of pneumococci with increased resistance to penicillin. Can. Med. Assoc. J. 117:1159–1161.
- Foster, J. A., and K. L. McGowan. 1994. Rising rate of pneumococcal bacteremia at the Children's Hospital of Philadelphia. Pediatr. Infect. Dis. J. 13:1143–1144. (Letter.)
- Friedland, I. R., and K. P. Klugman. 1992. Antibiotic-resistant pneumococcal disease in South African children. Am. J. Dis. Child. 146:920–923.
- Friedland, I. R., and G. H. McCracken, Jr. 1994. Management of infections caused by antibiotic-resistant *Streptococcus pneumoniae*. N. Engl. J. Med. 331:377–382.

- Hackenbeck, R., M. Tarpay, and A. Tomasz. 1980. Multiple changes of penicillin-binding proteins in penicillin-resistant clinical isolates of *Streptococcus pneumoniae*. Antimicrob. Agents Chemother. 17:364–371.
- Haglund, L. A., G. R. Istre, D. A. Pickett, D. F. Welch, D. P. Fine, and the Pneumococcus Study Group. 1993. Invasive pneumococcal disease in central Oklahoma: emergence of high-level penicillin resistance and multiple antibiotic resistance. J. Infect. Dis. 168:1532–1536.
- Handwerger, S., and A. Tomasz. 1986. Alterations in penicillin-binding proteins of clinical and laboratory isolates of pathogenic *Streptococcus pneumoniae* with low levels of penicillin resistance. J. Infect. Dis. 153:83–89.
- Hofmann, J., M. S. Cetron, M. M. Farley, W. S. Baughman, R. R. Facklam, J. A. Elliott, K. A. Deaver, and R. F. Breiman. 1995. The prevalence of drug-resistant *Streptococcus pneumoniae* in Atlanta. N. Engl. J. Med. 333: 481–486.
- Jetté, L. P., F. Lamothe, and the Pneumococcus Study Group. 1989. Surveillance of invasive *Streptococcus pneumoniae* infection in Quebec, Canada, from 1984 to 1986: serotype distribution, antimicrobial susceptibility, and clinical characteristics. J. Clin. Microbiol. 27:1–5.
- Marton, A., M. Gulyas, R. Munoz, and A. Tomasz. 1991. Extremely high incidence of antibiotic resistance in clinical isolates of *Streptococcus pneumoniae* in Hungary. J. Infect. Dis. 163:542–548.
- Mazzulli, T., A. E. Simor, R. Jaeger, S. Fuller, and D. E. Low. 1990. Comparative in vitro activities of several new fluoroquinolones and β-lactam antimicrobial agents against community isolates of *Streptococcus pneumoniae*. Antimicrob. Agents Chemother. 34:467–469.

- National Committee for Clinical Laboratory Standards. 1994. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically. Approved standard M7-A3. National Committee for Clinical Laboratory Standards, Villanova, Pa.
- National Committee for Clinical Laboratory Standards. 1995. Performance standards for antimicrobial susceptibility testing. M100-S6. National Committee for Clinical Laboratory Standards, Villanova, Pa.
- Nelson, C. T., E. O. Mason, and S. L. Kaplan. 1994. Activity of oral antibiotics in middle ear and sinus infections caused by penicillin-resistant *Streptococcus pneumoniae*: implications for treatment. Pediatr. Infect. Dis. J. 13:585–589.
- París, M. M., O. Ramilo, and G. H. McCracken, Jr. 1995. Management of meningitis caused by penicillin-resistant *Streptococcus pneumoniae*. Antimicrob. Agents Chemother. 39:2171–2175.
- Simor, A. E., L. Louie, J. Goodfellow, and M. Louie. 1995. Emergence of penicillin-resistant *Streptococcus pneumoniae*—southern Ontario, Canada, 1993–1994. Morbid. Mortal. Weekly Rep. 44:207–208.
- Viladrich, P. F., C. Cabellos, R. Pallares, F. Tubau, J. Martínez-Lacasa, J. Liñares, and F. Gudiol. 1996. High doses of cefotaxime in treatment of adult meningitis due to *Streptococcus pneumoniae* with decreased susceptibilities to broad-spectrum cephalosporins. Antimicrob. Agents Chemother. 40:218– 220.
- Viladrich, P. F., F. Gudiol, J. Liñares, R. Pallarés, I. Sabaté, G. Rufi, and J. Ariza. 1991. Evaluation of vancomycin for therapy of adult pneumococcal meningitis. Antimicrob. Agents Chemother. 35:2467–2472.