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## Phospholipase A<sub>2</sub> activity of guinea-pig perfused lungs: stimulation and inhibition by anti-inflammatory steroids

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Guinea-pig isolated lungs release prostaglandin (PG) endoperoxides and thromboxane A<sub>2</sub> (TXA<sub>2</sub>) into the perfusion fluid in response to stimuli including antigen challenge (Piper & Vane, 1969), mechanical trauma (Palmer, Piper & Vane, 1973), bradykinin or arachidonic acid (Vargaftig & Dao Hai, 1972) and rabbit aorta contracting substance - releasing factor (RCS-RF; Piper & Vane, 1969; Nijkamp, Flower, Moncada & Vane, 1976). Nijkamp et al. (1976) demonstrated that in relation to their antiinflammatory potency, corticosteroids inhibited the generation of TXA<sub>2</sub> induced by RCS-RF, but not that due to the precursor arachidonic acid. Thus, inhibition of the release of arachidonic acid could be related to the therapeutic action of these steroids. Flower & Blackwell (1976) demonstrated that arachidonic acid was released from cellular phosphatides in response to similar stimuli, and this led us to speculate that agents which release TXA<sub>2</sub> from lungs do so by "activating" phospholipase A<sub>2</sub>.

For these experiments, the guinea-pig perfused lungs and cascade superfusion apparatus were prepared and TXA<sub>2</sub> generation was measured as previously described (Nijkamp *et al.*, 1976). For assay of phospholipase A<sub>2</sub> activity, a mixture of 18.0 nmoles 2-acyl ([<sup>3</sup>H]-oleoyl) phosphatidylcholine and 1.8 nmoles [<sup>14</sup>C] oleic acid was injected into the pulmonary artery. The perfusate was collected for 7 min and the labelled fatty acids selectively extracted at pH 8.0 with 10 ml *n*-hexane. The solvent was evaporated to dryness and the <sup>3</sup>H/<sup>14</sup>C ratio estimated by conventional liquid scintillation counting techniques.

When injected into the pulmonary artery, histamine

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 $(2-5 \ \mu g)$ , RCS-RF  $(5-10 \ u)$ , bradykinin  $(1-5 \ \mu g)$ and arachidonic acid  $(1-5 \ \mu g)$  caused a release of PG endoperoxides and TXA<sub>2</sub>. Release of TXA<sub>2</sub> was blocked by indomethacin  $(1 \ \mu g/ml)$ . The release induced by histamine or RCS-RF was also blocked by dexamethasone (ID<sub>50</sub> 1.5  $\mu g/ml$ ) and hydrocortisone (ID<sub>50</sub> 33  $\mu g/ml$ ).

There was a small (1-3%) basal hydrolysis of the labelled phosphatide by the perfused lung, which increased gradually with time. This hydrolysis was inhibited by mepacrine  $(20 \ \mu g/ml)$ , procaine  $(40 \ \mu g/ml)$ , betamethasone  $(2 \ \mu g/ml)$ , dexamethasone  $(2 \ \mu g/ml)$  and hydrocortisone  $(50 \ \mu g/ml)$ . The steroids exhibited a time-dependent inhibition, the maximum effect occurring after 30 min infusion. Histamine  $(2 \ \mu g)$ , RCS-RF  $(5 \ u)$  and bradykinin  $(1 \ \mu g)$ stimulated phospholipid hydrolysis by 150–300%. Steroids and mepacrine blocked (60-90%) the stimulation due to histamine and RCS-RF but had only a small effect (10-20%) on the bradykinin stimulation.

In homogenates of guinea-pig lung, phospholipase  $A_2$  activity was inhibited by mepacrine and procaine. Steroids were without effect, indicating that these agents require intact cells to function effectively.

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