Cloning and Analysis of the Restriction-Modification System LlaBI, a Bacteriophage Resistance System from Lactococcus lactis subsp. cremoris W56

NIELS R. NYENGAARD, JENS FALKENBERG-KLOK, AND JYTTE JOSEPHSEN*

Department of Dairy and Food Science, The Royal Veterinary and Agricultural University, DK 1958 Frederiksberg C, Denmark

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The genes coding for the type II restriction-modification (R/M) system *Lla*BI, which recognized the sequence $5'-C \downarrow TRYAG-3'$, have been cloned from a plasmid in *Lactococcus lactis* subsp. *cremoris* W56 and sequenced. The DNA sequence predicts an endonuclease of 299 amino acids (33 kDa) and a methylase of 580 amino acids (65 kDa). A 4.0-kb *Hind*III fragment in pSA3 was able to restrict bacteriophages, showing that the cloned R/M system can function as a phage defense mechanism in *L. lactis*.

In the dairy industry, species of the genus Lactococcus are widely used as starter cultures in the manufacture of cultured milk products. One of the major problems encountered is infection of the lactococcal cultures by lytic bacteriophages, which results in dead or slow cheese vats. Since it is very difficult to avoid contamination with phages, the starter cultures have to be phage resistant. On the basis of their mode of action, four categories of naturally occurring phage resistance mechanisms have been identified in lactococci: adsorption inhibition, penetration blocking (6), restriction-modification (R/M) systems, and abortive infection. Several R/M systems have been isolated from lactococci, but only a few have been cloned and characterized (11). Two m⁵C methylases have been sequenced from the chromosomally encoded type II system ScrFI from Lactococcus lactis subsp. cremoris UC503. This system specifically recognizes 5'-CC \downarrow NGG-3' sequences, where N is any nucleotide (4, 20). A plasmid-encoded type II R/M system, *Lla*DCHI, recognizing $5' - \downarrow$ GATC-3', has recently been cloned and sequenced (13). Two m⁶A methylases and one restriction endonuclease make up the system. Mayo et al. (12) have reported type II endonuclease activity, designated *Lla*I and recognizing the sequence 5'-CC \downarrow WGG-3' (W is A or T), from L. lactis subsp. lactis NCDO 497. The isolation and purification of two type II endonucleases, LlaAI and LlaBI, recognizing 5'- \downarrow GATC-3' and 5'-C \downarrow TRYAG-3', from L. lactis subsp. cremoris W9 and W56, respectively, have been described previously (14). A type IIS R/M system on plasmid pTR2030 from L. lactis subsp. lactis ME2 has also been sequenced (16). It consists of an m^6A methylase and three open reading frames (ORFs) involved in the LlaI restriction activity. Furthermore, plasmid pTR2030 codes for an abortive infection mechanism (11).

L. lactis subsp. *cremoris* W56 has previously been isolated from the Danish mixed cheddar starter TK5 (9). It was shown that *L. lactis* subsp. *cremoris* W56 harbors at least three plasmids, pJW563, pJW565, and pJW566, which encode distinct R/M systems (10). Only *L. lactis* subsp. *cremoris* W56 and the transformant *L. lactis* subsp. *cremoris* MG1614(pJW563), and not transformants harboring plasmid pJW565 or pJW566, ex-

pressed type II activity (15). A restriction endonuclease, designated *LlaBI*, from *L. lactis* subsp. *cremoris* W56 was purified and its recognition sequence was determined (14).

In this study, the genes coding for the LlaBI system from L. lactis subsp. cremoris W56 were cloned and their nucleotide sequences were determined. The bacterial strains and plasmids are listed in Table 1. Lactococcal strains were grown at 30°C in M17 medium (Oxoid) supplemented with 0.5% glucose (GM17) and with 5 mM CaCl₂ when phages were used. Transformations were performed as described by Holo and Nes (8), with the transformants selected on agar medium with 10 µg of chloramphenicol per ml or 2.5 µg of erythromycin per ml. The antibiotics were purchased from Sigma Chemical Co. Escherichia coli was grown at 37°C in LB broth (17) supplemented with chloramphenicol, erythromycin, tetracycline, or ampicillin at 10, 50 to 250, 12.5, or 100 µg/ml, respectively, when required. The isometric-headed phage jj50 (10) and the prolateheaded phage c2, both with double-stranded DNA (11), were propagated and titrated by the method of Terzaghi and Sandine (19). Plasmid DNA was extracted from L. lactis strains by the method of Andresen et al. (2). Subcloning and nucleotide sequencing were performed by standard DNA techniques (17).

A restriction map of plasmid pJW563, which expressed *Lla*BI activity, was constructed (Fig. 1). When direct cloning of the entire R/M system was attempted in *E. coli* XL1Blue or HB101, only plasmids with deletions were found (data not shown). Thus, pJW563 was cloned only in *L. lactis* subsp. *cremoris* MG1614.

Because of difficulties in cloning the entire system in L. lactis subsp. cremoris MG1614, a decision was made to determine the location of the genes for the R/M system on plasmid pJW563. A chloramphenicol resistance cassette was inserted into one of the ClaI sites, resulting in plasmid pJWC1. L. lactis subsp. cremoris MG1614(pJWC1) expressed R/M activity similar to that of the wild-type plasmid. Deletion of the 1.2-kb BclI fragment from pJWC1 resulted in plasmid pJWC2, which showed neither endonuclease nor methylase activity (Fig. 1). When an erythromycin cassette was inserted into the unique BglII site in plasmid pJWC1, the transformant L. lactis subsp. cremoris MG1614(pJWE1) did not express any LlaBI endonuclease activity or phage resistance. However, phages propagated on L. lactis subsp. cremoris MG1614(pJWE1) were not restricted by L. lactis subsp. cremoris MG1614(pJW563), showing that pJWE1 expressed methylase activity. All these results indi-

^{*} Corresponding author. Mailing address: Department of Dairy and Food Science, The Royal Veterinary and Agricultural University, Rolighedsvej 30, DK 1958 Frederiksberg C, Denmark. Phone: 45 35 28 32 32.

Strain or plasmid	Relevant characteristics ^a	Reference or source
Strains		
L. lactis subsp. cremoris MG1614	Plasmid-free derivative of L. lactis subsp. cremoris 712	7, 21
E. coli XL1Blue	Transformation host; mrr ⁺	Stratagene Ltd.
Plasmids		
pJW563	Resident plasmid of W56; R^+/M^+ ; 11.5 kb	10
pSA3	Shuttle vector; Cm ^r Tc ^r Em ^r ; 10.2 kb	3
pBluescriptIISK ⁺	Cloning vector for sequencing; 3.0 kb; Ap ^r	Stratagene Ltd.
pUC7,erm	pUC7 $\Omega(1.1\text{-kb HinPI pIL}253 \text{ erm})$	W. M. de Vos
pJWC1	pJW563 Ω (4.0-kb cam cassette from pVC5 [21]) in ClaI site; R ⁺ /M ⁺ ; Cm ^r	Anne Gravesen
pJWC2	pJWC1 Δ (1.2-kb <i>Bcl</i> I fragment); R ⁻ /M ⁻ ; Cm ^r	This study
pJWE1	pJWC1 $\Omega(1.1\text{-kb } erm \text{ cassette from pUC7}, erm)$ in BglII site; R ⁻ /M ⁺ ; Cm ^r Em ^r	This study
pSNB1	pSA3 $\Omega(4.0\text{-kb HindIII} \text{ fragment from pJW563})$	This study

TABLE	1.	Bacteria	and	plasmids used
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^a Ap^r, ampicillin resistance; Cm^r, chloramphenicol resistance; Em^r, erythromycin resistance; Tc^r, tetracycline resistance.

cated that the endonuclease gene was located near the *Bgl*II site, whereas at least part of the methylase gene was located on the 1.2-kb *Bcl*I fragment.

The entire R/M system was then cloned in shuttle vector pSA3 on a 4.0-kb *Hind*III fragment containing the *Bgl*II and *Bcl*II sites, resulting in plasmid pSNB1. *L. lactis* subsp. *cremoris* MG1614(pSNB1) transformants expressed both *Lla*BI endonuclease and methylase activity (data not shown) and restricted phage jj50 with an efficiency of plaquing of 10^{-4} and phage c2 with an efficiency of plaquing of 10^{-3} . This demonstrates that the genes encode both the endonuclease and the methylase from the *Lla*BI R/M system and that they can be cloned and used to increase phage resistance in *Lactococcus* strains.

When the 4.0-kb *Hin*dIII fragment in pBluescriptIISK⁺ was used to transform *E. coli* XL1Blue, only plasmids with deleted inserts were obtained, indicating that the endonuclease and/or

methylase expression may be lethal to *E. coli* XL1Blue. Additionally, since the *Hind*III₁-*Bg*III fragment was also difficult to clone, it is probably the methylase which caused this apparent lethality in *E. coli* XL1Blue, because of the presence of the Mrr restriction system, which restricts methylated DNA (22).

The nucleotide sequence of 3,476 bp in the 4.0-kb *Hin*dIII fragment containing the *Lla*BI R/M genes was determined and analyzed as shown in Fig. 2. ORF1 was 1,740 bp and predicts a protein of 580 amino acids (65 kDa), while ORF2 was 897 bp and predicts a protein of 299 amino acids (33 kDa). The inactivated *BgI*II site in plasmid pJWE1 (Fig. 1) was located in ORF2, showing that ORF2 codes for the endonuclease R · *Lla*BI. The deleted 1.2-kb *BcI*I fragment (positions 1305 to 2520) in plasmid pJWC2, spanning 856 bp downstream in ORF1, shows that ORF1 codes for the methylase M · *Lla*BI. A secondary structure indicating a putative terminator structure with a 13-bp inverted repeat sequence ($\Delta G = -16.9$ kcal [ca.



FIG. 1. Maps, R/M activities of pJW563 and its derivatives, and the products of the *Lla*BI genes. Plasmids are shown linearized by cleavage at an *Xho*I site; pJW563 is a wild-type plasmid. The positions of the putative ORFs and the direction of transcription are indicated (arrows). The putative sizes (in amino acids) are indicated for the methylase and the restriction endonuclease. Bc, *Bcl*I; Bg, *Bgl*II; C, *Cla*I; E, *Eco*RI; H, *Hind*III; X, *Xho*I.

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FIG. 2. Complete nucleotide sequence of the 3,476 bp of the 4.0-kb *Hind*III fragment containing the *Lla*BI R/M enzyme genes. Nucleotide positions are numbered consecutively starting at the left *Hind*III site (H1, Fig. 1). The restriction endonuclease (nucleotides 2234 to 3131) is encoded by the top strand; the modification enzyme (nucleotides 191 to 1931) is encoded by the bottom strand. Promoter sequences (-10 and -35), Shine-Dalgarno sequences (S.D.) (underlined), stop codons (*), and translation start codons (boldface and gene designations) are indicated. The deduced amino acid sequences for both proteins are also shown. The amino acids are aligned with the first nucleotide of each codon. Motifs I (boldface) and IV (underlined boldface letters) are also indicated.

1698 TTC AAT ATT TTT AAG AGA AAA ATG AAG CTC ATT TTC TTC AAT ATA ATT TCT TAT GTC TAA ATT TGA GTA GCC TAA AAG TTA TAA AAA TTC TCT TTT TAC TTC GAG TAA AAG AAG TTA TAT TAA AGA ATA CAG ATT TAA ACT CAT CGG ATT I Е N К L s F н LENEEIYNRIDLNS 1773 TAG CTC GTT TAT CAA ACT ATT TTT AAG GGA TTC TAT GGG TAT TTT TTT TTC GGT GAA GTA ATT TCT TAT TAT CTC ATC GAG CAA ATA GTT TGA TAA AAA TTC CCT AAG ATA CCC ATA AAA AAA AAA CCC CTT CAT TAA AGA ATA ATA GAG L E N I L S N K L S E 1848 Т Ρ і к к Е Т F Y N E GCT TAA TAT TTC TTT GCT TGA ATA TTT ATC GAG TAT TTT TTT TAT AAA CTC TAT ATT TGT TTG TTT ATC TAT AAC CGA ATT ATA AAG AAA CGA ACT TAT AAA TAG CTC ATA AAA AAA ATA TTT GAG ATA TAA ACA AAC AAA TAG ATA TTG S L I E K S S Y K D L I K K I F E I N T Q K D I V 1000 SLI 1923 M LlaBI -10 -35 OREX S.D. м Е L М 2013 V L N E R L K E L N I S Q N K F A K Q S H I R P I GTT CTT AAC GAA CGG CTA AAA GAA CTA AAT ATA TCA CAA AAT AAG TTT GCG AAG CAA TCA CAT ATT AGG CCG ATA CAA GAA TTG CTT GCC GAT TTT CTT GAT TTA TAT AGT GTT TTA TTC AAA CGC TTC GTT AGT GTA TAA TCC GGC TAT 2088 *** 0 CAA TAA ATGATATCTGCAATAACAGTACTAAAAGAATAGAAGTTTCAACTATCAACAAAATACTAATTCAATTAAATAAGATAGGTATTCGTAAATA -35 R LlaBI 2270 D L E L A I T D Q I V D G S K V N K K G K L F L N GAT TTA GAA CTA GCT ATT ACT GAT CAA ATA GTT GAC GGT TCT AAA GTA AAT AAA GAA GGG AAA TTA TTT TTA AAT CTA AAT CTT GAT CGA TAA TGA CTA GTT TAT CAA CTG CCA AGA TTT CAT TTA TTT TCT CCC TTT AAT AAA AAT TTA 2345 G A E A K Q S L I R S S K L I N Y V H E F V K H E GGA GCA GAA GCA AAA CAA TCT TTA ATT AGA TCT AGT AAA CTT ATT ATT TAT GTT CAC GAG TTT GTA AAA CAT GAA CCT CGT CTT CGT TTT GTT AGA AAT TAA TCT AGA TCA TTT GAA TAA TTA ATA CAA GTG CTC AAA CAT TTT GTA CTT 2420 L I R N S V E E S L I F P P L G Q T N P E I K L T CTA ATA AGA AAT AGT GTT GAA GAA TCT CTG ATA TTC CCC CCA TTA GGT CAG ACA AAC CCT GAA ATA AAA CTT ACT GAT TAT TCT TTA TCA CAA CTT CTT AGA GAC TAT AAG GGG GGT AAT CCA GTC TGT TTG GGA CTT TAT TTT GAA TGA 2495 G M G M F K Q K D Q D V C V K P Q G V L P E R T L I G GGT ATG TTT AAA CAA AAG GAT CAA GAT GTT TGT GTA AAG CCT CAG GGA GTT TTA CCC GAA AGA ACT TTA ATT GGA CCA TAC AAA TTT GTT TTC CTA GTT CTA CAA ACA CAT TTC GGA GTC CCT CAA AAT GGG CTT TCT TGA AAT TAA CCT 2570 W G P M I N S G L Y C D Y G R A Y A E R V L S I N TGG GGA CCT ATG ATA AAT TCG GGA TTA TAC TGT GAT TAT GGT CGC GCT TAT GCA GAA AGA GTA TTA TCT ATC AAT ACC CCT GGA TAC TAT TTA AGC CCT AAT ATG ACA CTA ATA CCA GCG CGA ATA CGT CTT TCT CAT AAT AGA TAG TTA 2645 V R S Q L S S L D K N S D T L F E R M F A E A L N GTA AGA AGT CAA TTA AGT AGT CTA GAT AAA AAT TCT GAT ACG TTA TTT GAG CGG ATG TTT GCA GAA GCA TTA AAT CAT TCT TCA GTT AAT TCA TCA GAT CTA TTT TTA AGA CTA TGC AAT AAA CTC GCC TAC AAA CGT CTT CGT AAT TTA 2720 L H v G v K Ι М Е v P Ε TTA CAC GAG TTG TAT CCA AAA ATA GTT ATG GGA GAA GTA TAT GTT ATT CCA GTT TAT GAA TAC GAC GAC CAA GCA AAT GTG CTC AAC ATA GGT TTT TAT CAA TAC CCT CTT CAT ATA CAA TAA GGT CAA ATA CTT ATG CTG CTG GTT CGT 2795 M I N N Q V K F K S R R T N L E K Y I N F F Y Y L ATG ATA AAT AAT CAA GTT AAG TTC AAG TCA AGA AGA AGA ACA AAT TTA GAA AAA TAC ATT AAT TTT TTC TAT TAT TTA TAC TAT TTA TTA GTT CAA TTC AAG TTC AGT TCT TCT TGT TTA AAT CTT TTT ATG TAA TTA AAA AAG ATA ATA AAT 2870 G S G R D E Q D L E E D K Q K Y E R C A L V I I D F agt ggc aga gat cag gat ctt gaa gaa gaa gaa caa aag tac gaa agg tac gaa agg tac gaa at ttt TCA CCG TCT CTA CTT GTC CTA GAA CTT CTT CTG TTT GTT TTC ATG CTT TCC ACG CGT AAC CAA TAT TAT CTA AAA 2945 V Y K N T A RG ΕL Κ R G А AGA GGA GAT CAA GCC AAA GTC TAT AAA AAT ACT GCA GAG TTA AAA GCT AGG GGC TTA GTC AGA AAT GAT TTT GAG TCT CCT CTA GTT CGG TTT CAG ATA TTT TTA TGA CGT CTC AAT TTT CGA TCC CCG AAT CAG TCT TTA CTA AAA CTC 3020 VE T. А E т D к F Т Е D Τ. τ. 3095 S V A K F E N Q T R P L *** TCT GTT GCG AAG TTT GAA AAT CAA ACG CGC CCT CTC TGAACTCCAAATATCTTAGGCTGGTATTCCCATTAATACCTTGATTTCAGT AGA CAA CGC TTC AAA CTT TTA GTT TGC GCG GGA GAG ACTTGAGGTTTATAGAATCCGACCATAAGGGTAATTATGGAACTAAAGTCA v 3182 TCTGTGGCTTTTCGGCTTCTCCAAGGTAAAGAAGCCAAGAAAAATATATAAGGAGCTTACCAGACGTAGGGGAATTAGCACCTTCTCCGACATGCCTC 3281 ACTTTGATAAAATTTATTCCGTCGTTTAATAGGTCGATGGTCTTGTTCTATTAAATTGTTAAGATACTTCACAGTTCGGTGCTCTGTCTTAGTATATAA TGAAACTATTTTAAATAAGGCAGCAAATTATCCAGCTACCAGAACAAGATAATTTAACAATTCTATGAAGTGTCAAGCCACGAGACAGAATCATATATT 3380

ACCCACACTCTGTAACTTTCTAAAGCGGAGCCAAGAGAAGGTGCTTTATCGTGCAATTGGTGCGACGATCAAAATATTATTGGGAATACCTGCTTA 3 ' TGGGTGTGAGACAATGAAAGATTTCGCCTCGGTTCTCTTCCACGAAATAGCACGTTAACTACGCCTGCTAGTTTTATAATAACCCTTATGGACGAAAT 5 '

FIG. 2-Continued.

-70.7 kJ]) was found downstream of the methylase gene with a 2-bp overlap. The results show that the *Lla*BI R/M system consists of two divergently transcribed genes encoding a methylase, M · *Lla*AI, and an endonuclease, R · *Lla*AI.

No primary sequence similarities between the restriction endonuclease R \cdot *Lla*BI and the corresponding methylase M \cdot *Lla*BI or other type II restriction endonucleases were found. As shown in Fig. 2, the deduced amino acid sequence of M \cdot *Lla*BI has all the characteristics of subclass γ of the m⁶A methylases: a motif I, a short variable region (14 amino acids), and a motif IV with asparagine in the first position (NPPY) (23).

During the cloning of the *Lla*BI system, it was found that subclones of pJW563 containing fragments of the *Lla*BI methylase were not resistant to *PstI* restriction, although plasmid pJW563 was resistant to digestion by the *PstI* restriction endonuclease (data not shown). The *PstI* endonuclease recognizes 5'-CTGCA \downarrow G-3' and cuts as indicated by the arrow. *Lla*BI can recognize the same sequence, 5'-C \downarrow TGCAG-3' (and 5'-C \downarrow TATAG-3'), but cuts the recognition sequence at a different position (*Lla*BI generates 5' overhangs while *PstI* yields 3' overhangs). This indicates that the adenine in the *PstI* recognition sequence has been methylated by the *Lla*BI methylase. This reinforces the assumption that M \cdot *Lla*BI is an m⁶A methylase.

Preceding the *LlaBIR* gene, and in the same direction, several putative small ORFs were found. ORFX, of 30 codons (Fig. 2), was the largest and may code for an inhibitory peptide preventing endonuclease subunit association, as found for the *Pvu*II R/M system (1).

The high C+G content in the recognition sequences of *Scr*FI (5'-CC \downarrow NGG-3') and *Lla*I (5'-CC \downarrow WGG-3'), in contrast to those of *Lla*AI or *Lla*DCHI (5'- \downarrow GATC-3') and *Lla*BI (5'-C \downarrow TRYAG-3'), may have a practical implication in the use of these systems as phage defense mechanisms, since lactococcal phage DNA has <40% C+G (11).

The average G+C content of the *Lla*BI genes is 27.8% (31.5% for R \cdot *Lla*BI and 25.7% for M \cdot *Lla*BI), which is much lower than the 34 to 43% G+C content normally found in lactococci as measured by determination of the melting temperature (18). This may indicate that the *Lla*BI R/M system originates from a genus other than *Lactococcus* or that it is an inherent feature of particular R/M systems. Genes coding for the phage resistance mechanism abortive infection in *L. lactis* also have been found to have a low G+C content (26 to 29%) (5).

Nucleotide sequence accession number. The nucleotide sequence (3,476 bp) of pSNAI has been deposited in the EMBL Nucleotide Sequence Database under accession no. X97263.

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