Short reports

Klinikum der Christian-Albrechts-Universität zu Kiel, Frauenklinik Onkologisches Labor, Michaelisstrasse 16, D-24105 Kiel, Germany J Weimer M Kiechle N Arnold

Gemeinschaftspraxis Dr med A Ovens-Raeder Dr med C Waldenmaier, Theodolindenstrasse 97, 81545 München, Germany U Wiedemann A Ovens-Raeder

Charité, Campus Virchow-Klinikum Institut für Humangenetik Molekulare Zytogenetik, Augustenburger Platz 1, 1353 Berlin, Germany H Neitzel H Tönnies

Klinikum Rosenheim, Pettenkoferstrasse 10, 83022 Rosenheim, Germany E Ruhenstroth

Correspondence to: Dr Weimer, Joerg.Weimer@t-online.de

Revised version received 18 October 1999 Accepted for publication 2 December 1999

Delineation of a complex karyotypic rearrangement by microdissection and CGH in a family affected with split foot

Jörg Weimer, Marion Kiechle, Ute Wiedemann, Holger Tönnies, Heidemarie Neitzel, Eberhard Ruhenstroth, Angela Ovens-Raeder, Norbert Arnold

Abstract

We report on a male patient and members of his family with additional material in chromosome 3. This derivative chromosome 3 was transmitted from his mother who had a complex rearrangement between chromosomes 2, 3, and 7. It was possible to delineate her chromosomal rearrangement by microdissection and reverse painting and to exclude these aberrations from being responsible for neonatal deaths and several abortions in this family. Two members of this family suffer from ectrodactyly or split hand/foot malformations (SHFM) of the feet which



Figure 1 Bilateral split feet of the patient's sister. The mother of the patient also had similar malformations of her feet, which were amputated and replaced by prostheses in infancy.



Figure 2 Pedigree of the patient's family. The mother (1) and sister (2) exhibit the same chromosomal aberration and both have bilateral deformities of their feet. The patient (3) exhibits only widely spaced first and second toes, intestinal malformations, and the maternally transmitted derivative chromosome 3. NND=neonatal death.

possibly correlates with the derivative chromosome 7 containing a breakpoint in the *SHFM1* critical region involving several homeobox genes. (*f Med Genet* 2000;37:442–445)

Keywords: microdissection; CGH; SHFM; ectrodactyly

Split hand/foot malformation (SHFM) is a heterogeneous limb developmental disorder, characterised by missing digits, fusion of the remaining digits, and a deep median cleft in the hands and feet.¹² The hereditary form of SHFM is often correlated with a chromosomal breakage in the *SHFM1* critical region 7q21.3-q22.1.¹² The *DSS1* gene,³ as well as the distal-less homeobox genes DLX5 and DLX6,¹ are located in this region. DSS1 is possibly responsible for SHFM phenotypes.³ Marinoni *et al*⁴ reported the coexistence of split foot and developmental anomalies with a deletion of three microsatellite markers in 7q21.2-q22.1. We delineate a chromosomal rearrangement in a family with two members suffering from bilateral split feet but no other developmental anomalies, and one member with very mild deformities of the feet and other developmental anomalies. Also, several cases of neonatal death and abortions have occurred in this family.

Case report

The 5 month old patient has minor malformations like widely spaced first and second toes, significant nail hyperplasia, disproportionate size of fingers to hand, a relatively long thorax, epicanthus medialis, intestinal malformations, and possibly a spina bifida occulta. So far he has no head control. Like his mother, his sister also has split feet (fig 1), whereas he himself only has widely spaced first and second toes. A hereditary connection between these dysmorphic symptoms and chromosomal aberrations seems very likely.

The patient's maternal grandfather has two childless sisters, one of whom had three abortions and the other three neonatal deaths (fig 2).



Figure 3 Presumed mechanism of translocation in G banded ideograms.²⁰ The fluorescence images are M-FISH pictures of the derivative chromosomes 2, 3, and 7 compared with their G banded chromosomes.

CYTOGENETIC ANALYSIS

Initially, GTG banding of this newborn boy with mild dysmorphic features showed a 46,XY,der ins(3)(p?) karyotype with unspecified breakpoints (see G banding of the derivative chromosome 3 in fig 3). Analysis of the



Figure 4 CGH results with DNA of the patient. The ideogram shows a gain limited to 2q33-2q34 and 7q22-7q31. The fluorescence signals are given on the left.

maternal karyotype and the karyotype of the patient's sister shows an identical complex translocation pattern including chromosomes 2, 3, and 7. Fluorescence in situ hybridisation with commercial human whole chromosome painting probes (WCP) of chromosomes 2, 3, and 7 indicates an insertion of additional chromosomal material from chromosomes 2 and 7 into the short arm of chromosome 3 of the patient (see M-FISH pictures in fig 3). The chromosome 2 material was distally inserted next to the proximally inserted chromosome 7 material in the derivative chromosome 3. In his mother's karyotype the same composition of signals of her derivative chromosome 3 were found. The transmitted derivative chromosome 3 of the patient is the same as in his mother's and sister's karyotype. Furthermore, translocation products of chromosomes 2 and 7 were detected in the mother's and sister's karyotype.

Comparative genomic hybridisation (CGH)^{6 7} analysis of the patient's DNA was performed to define the contents of the additional inserted chromosomal material of the derivative chromosome 3 (fig 4), and thus the inserted regions could almost be limited to 2q33~2q34 and 7q22~7q31. Chromosome microdissection⁸ followed by reverse painting^{10 11} was applied to characterise the exact breakpoints of the balanced translocation in the patient's mother. We generated whole chromosome painting probes of the derivative chromosomes 2, 3, and 7 by microdissection. The multicolour fluorescence in situ hybridisation (M-FISH)¹² results of the reverse painting (fig 5) identified the translocated areas. To detect the point of insertion in chromosome 3, a further experiment was necessary. The best strategy to identify insertion points is to dissect

444



Figure 5 Reverse painting of the maternal derivative chromosomes der(2), der(3), and der(7) generated by microdissection on normal metaphases.



Figure 6 Reverse painting of a distal part of the maternal aberrant arm of the derivative chromosome 3 ($3pter \rightarrow 3p21.3::2q32 \rightarrow 2q34::7q22 \rightarrow 7q22$) isolated by microdissection and hybridised on normal metaphases.

proximally or distally to the inserted region. We used a partial chromosome painting probe of the distal p arm of the derivative chromosome 3, der(3)(3pter \rightarrow ?::2q32 \rightarrow 2q34::7q22), obtained by microdissection,^{13 14} to determine the breakpoint of the insertion into the derivative chromosome 3. The results of this microdissection and reverse painting (fig 6) showed a very distinct signal on chromosome 3 distal to the insertion breakpoint at the telomeric end of der(3p) as well as on $2q32 \rightarrow 34$, but almost no signal on chromosome 7. Therefore, the insertion breakpoint of the derivative chromosome 3 could be determined as 3p21.3. The results of microdissection and reverse painting pinpoint the inserted areas in chromosome 3 as 2q32->2q34::7q22->7q31.3 (figs 5 and 6). All

Table 1 Known genes and FISH signal distribution of cosmid signals located in the SHFM1 critical region of the mother

SHFM1 critical region	Gene	Cosmid probes	Distance to DSS1	Distribution of signals		
7q21.3 7q22.1	PDK4 DNCI1 SLC25A13 DSS1 DLX6, DLX5	cos426/61d4 cos437/237d7 cos405/24g11 cos274/64b4 cos256/191f10 cos DLX	400 kb 100 kb	der (7) + + +	chromosome 7 + + + + + + + +	der (3) + +

PDK4 = pyruvate dehydrogenase kinase isoenzyme 4. *DNCI1* = dynein cytoplasmic intermediate polypeptide 1. *SLC25A13* = adult onset type II citrullinaemia (CTLN2). *DSS1* = deleted in split hand/split foot 1 region. *DLX5* and *DLX6* = distal less homeobox gene 5 and 6. Additional information on the cosmid probes can be found at: http://www.genet.sicklids.on.ca/chromosome7/

reverse painting results showed the following complex karyotype: 46,XX,rev ish t(2;3;7;) (2pter \rightarrow 2q32::7q31.3 \rightarrow 7qter; 3pter \rightarrow 3p21.3:: 2q32 \rightarrow 2q34::7q22 \rightarrow 7q31.3::3p21.3 \rightarrow 3qter; 7pter \rightarrow 7q22::2q34 \rightarrow 2qter). All identified breakpoints are shown in fig 3.

Further investigations were performed to confirm the breakpoint location of the derivative chromosome 7 in the *SHFM1* critical region. Using six cosmids located in the *SHFM1* critical region, the breakpoint could be narrowed down to 7q22.1 (table 1). The breakpoint is distal to cos405/24g11 and proximal to cos274/64b4 in the *SHFM1* critical region.

Discussion

We suggest two possibilities for the development of this complex aberration. The first event is a reciprocal translocation between chromosomes 2 and 7 with the breakpoints at 2q34 and 7q22 (fig 3). One product of this translocation event is identical to one derivative chromosome in the karyotype of the patient's mother and sister (derivative chromosome 7). The other derivative product is the starting point for a second translocation with a chromosome 3. The chromosomal region $(2q32 \rightarrow 2q34::7q22 \rightarrow 7q31.3)$ of this intermediate product is inserted into 3p21.3 (fig 6). The derivative chromosomes 2 and 3 in the patient's mother are derived from the second translocation event. Another event with breakpoints in 2q32 and 7q31 in the first translocation is a possibility. If so, the second translocation event would have happened between the derivative chromosome 7 as a presumed intermediate product and a chromosome 3 with an inverted insertion. Regarding the G banding of the derivative chromosome 3, we prefer the mechanism in fig 3, since the G banding of the derivative chromosome 3 is in accordance with the ideogram of the rearrangement model.

Split hand/foot malformation (SHFM) is an autosomal dominant developmental malformation. Variations of SHFM where patients have deformities of the feet without hand deformities have been described.4 15 Ectrodactyly has been described in several patients, often associated with cytogenetically visible rearrangements involving the chromosomal region (7)(q21.2-22.1).^{4 15 16} We could confirm one breakpoint of the complex aberration in the (7)(q21.3-22.1) SHFM1 region,^{15 16} next proximally to the DSS13 and the distal-less homeobox genes like DLX5 and DLX6.3 Neither DSS1 nor the DLX genes are directly affected by the breakpoint on the derivative chromosome 7 of the mother and daughter. If DSS1 is not affected indirectly by the close location of the breakpoint (between around 100 kb for cos274/64b4 and 400 kb for cos405/24g11 proximal to DSS15), a further developmental gene for split foot could exist. A close balanced reunion junction near DSS1 could influence the transcription of DSS1 in the developmental process and cause malformations of the feet.

The malformations in our patient are possibly the result of trisomy of genes in the inserted regions like DSS1, DLX6, and DLX5. The derivative chromosome 7 showing a breakpoint in 7q22 could have caused the foot malformations of the patient's mother and sister if the supposed gene for ectrodactyly, SHFM1, is influenced.

From this hypothesis, further SHFM phenotypes should be present in this family (fig 2) if both translocation events of the complex rearrangement between chromosomes 2, 3, and 7 occurred in more than one meiotic cell division. The derivative chromosome 7 would be one of the first results of the first translocation event. Then the second translocation event could have happened at gametogenesis of the next generation. However, no foot malformations have been reported in the patient's grandparents and their sisters (fig 2). Therefore, it is very likely that both meiotic translocation events happened at gametogenesis of one maternal grandparent simultaneously. However, it should be noted that ectrodactyly sometimes skips a generation.¹⁷⁻¹⁹ The patient's grandfather's sisters with three abortions and three neonatal deaths definitely did not harbour the translocation.

This work was supported by Deutsche Forschungs-Gemeinschaft (Ar262/1-1). We thank Dr Stephen W Scherer from the Department of Genetics, The Hospital for Sick Children, Toronto for suggestions and cosmid probes.

- 1 Scherer SW, Poorkaj P, Massa H, Soder S, Allen T, Nunes M, Geshuri D, Wong E, Belloni E, Little S. Physical map-ping of the split hand/split foot locus on chromosome 7 and implication in syndromic ectrodactyly. *Hum Mol Genet* 1994;**3**:1345-54.
- 2 Scherer SW, Poorkaj P, Allen T, Kim J, Geshuri D, Nunes M, Soder S, Stephens K, Pagon RA, Patton MA. Fine mapping of the autosomal dominant split hand/foot locus on chromosome 7, band q21.3-q22.1. Am J Hum Genet 1994:55.12-20
- 3 Crackower MA, Scherer SW, Rommes JM, Hui CC, Poorkaj P. Soder S, Cobber JD, Hudgins L, Evans JP, Tsui LC. Characterization of the split hand/split foot malformation locus SHFM1 at 7q21.3-q22.1 and analysis of a candidate gene for its expression during limb development. *Hum Mol* Genet 1996:5:571-9.
- 4 Marinoni JC, Stevenson RE, Evans JP, Geshuri D, Phelan MC, Schwartz CE. Split foot and developmental retarda-

tion associated with a deletion of three microsatellite markers in 7q21.2-q22.1. *Clin Genet* 1995;47:90-5.

- 5 Scherer S. Personal communication, Department of Genetics, The Hospital for Sick Children, Toronto. Additional information on the cosmid probes can be found at: http://www.genet.sickkids.on.ca/chromosome7/
- 6 Kallioniemi A, Kallionemi OP, Sudar D, Rutovitz D, Gray JW, Waldmann FM, Pinkel D. Comparative genomic hybridization for molecular genetic analysis of solid tumors. *Science* 1992;**258**:818-21.
- Du Manoir S, Speicher MR, Joos S, Schröck E, Popp S, Dohmer H, Kovacs G, Robert-Nicoud M, Lichter P, Cremer T. Detection of complete and partial chromosome gains and losses by comparative genomic in situ hybridization. Hum Genet 1993;90:590-610.
- Senger G, Lüdecke HJ, Horsthemke B, Claussen U. Micro-8 dissection of banded human chromosomes. Hum Genet 1990;84:507-11.
- Weimer J, Kiechle M, Senger G, Wiedemann U, Ovens-Raeder A, Schuierer S, Kautza M, Siebert R, Arnold N. An easy and reliable procedure of microdissection technique for the analysis of chromosomal breakpoints and marker chromosomes. Chromosome Res 1999;7:355-62
- Carter NP, Ferguson-Smith MA, Perryman MT, Telenius H, Pelmear AH, Leversha MA, Glancy MT, Wood SL, 10 Cook K, Dyson HM, Ferguson-Smith ME, Wilatt LR. Reverse chromosome painting: a method for rapid analysis of aberrant chromosomes in clinical cytogenetics. J Med Genet 1992;29:299-307.
- 11 Thangavelu M, Pergament E, Epinosa R III, Bohlander SK. Characterization of marker chromosomes by microdissection and fluorescence in situ hybridization. Prenat Diagn 1994;14:583-8.
- 12 Speicher MR, Ballar SG, Ward DC. Karyotyping human chromosomes by combinatorial multi-fluor FISH. Nat Genet 1996;12:368-75.
- 13 Senger G, Chudoba I, Friedrich U, Tommerup N, Claussen U. Brondum-Nielsen K. Prenatal diagnosis of a half-cryptic translocation using chromosome-microdissection. Prenat Diagn 1997:17:369-74
- 14 Müller-Navia J, Nebel A, Schleiermacher E. Complete and precise characterization of marker chromosomes by amplification of microdissection in prenatal diagnosis. Hum Genet 1995;96:661-7
- Ignatius J, Knuutila S, Scherer SW, Trask B, Kere J. Split hand/split foot malformation, deafness, and mental retar-15 dation with a complex cytogenetic rearrangement involving 7q21.3. *J Med Genet* 1996;**33**:507-10.
- Palmer SE, Scherer SW, Kukolich M, Wijsmann EM, Tsui LC, Stephens K, Evans JP. Evidence for locus heterogen-eity in human autosomal dominant split hand/split foot malformations. Am J Hum Genet 1994;55:21-6.
- 17 MacKenzie HJ, Penrose LS. Two pedigrees of ectrodactyly. Ann Eugen 1951;16:88-96.
- 18 Graham JB, Badgley CE. Split-hand with unusual complica-
- tions. Am J Hum Genet 1955;7:44-50.
 19 Neugebauer H. Spalthand und -fuss mit familiärer Besonderheit. Z Orthop 1962;95:500-6.
 20 ISCN (1995). Table 1. In: Mittelman F, ed. An international
- system for human cytogenetic nomenclature. Basel: S Karger, 1995