- 22 Hirst M, Grewal P, Flannery A, Slatter R, Maher E, Barton D, Fryns JP, Davies K. Two new cases of FMR1 deletion associated with mental impairment. Am J Hum Genet 1995;56:67-74.
- 23 De Graaff E, De Vries BB, Willemsen R, van Hemel JO, Mohkamsing S, Oostra BA, van den Ouweland AM. The fragile X phenotype in a mosaic male with a deletion show-ing expression of the FMR1 protein in 28% of the cells. Am
- *J Med Genet* 1996;**6**4:302-8. 24 Schmucker B, Ballhausen WG, Pfeiffer RA, Mosaicism of a microdeletion of 486 bp involving the CGG repeat of the FMR1 gene due to misalignment of GTT tandem repeats
- FMRI gene due to misalignment of GT1 tandem repeats at chi-like elements flanking both breakpoints and a full mutation. *Hum Genet* 1996;98:409-14.
 25 Sutherland GR. Heritable fragile sites on human chromosomes. II. Distribution, phenotypic effects, and cytogenetics. *Am J Hum Genet* 1979;31:136-48.
 26 Mila M, Castellvi-Bel S, Gine R, Vazquez C, Badenas C, Sanchez A, Estivill X. A female compound heterozygote (and subtraction of the subtraction) of the the CCC ENDIA.
- (pre- and full mutation) for the CGG FMR1 expansion. Hum Genet 1996;98:419-21.
- 27 Brown V, Small K, Lakkis L, Feng Y, Gunter C, Wilkinson KD, Warren ST. Purified recombinant Fmrp exhibits selective RNA binding as an intrinsic property of the frag-ile X mental retardation protein. f Biol Chem 1998;273: 15521-7.
- 28 Hagedorn CH, Spivak-Kroizman T, Friedland DE, Goss DJ, Xie Y, Expression of functional eIF4e human: purifica-tion, detailed characterization, and its use in isolating eIF-4e binding proteins. *Protein Express Purif* 1997;9:53-60.
- 29 Devys D, Lutz Y, Rouyer N, Bellocq JP, Mandel JL. The FMR1 protein is cytoplasmic, most abundant in neurons and appears normal in carriers of a fragile X premutation. *Nat Genet* 1993;4:335-40.

- 30 Willemsen R, Mohkamsing S, de Vries B, Devys D, van den Ouweland A, Mandel JL, Galjaard H, Oostra B. Rapid antibody test for fragile X syndrome. Lancet 1995;345: 1147-8.
- Willemsen R, Smits A, Mohkamsing S, van Beerendonk H, de Haan A, de Vries B, van den Ouweland A, Sistermans E, Galjaard H, Oostra BA. Rapid antibody test for diagnosing fragile X syndrome: a validation of the technique. *Hum* Genet 1997;**99**:308-11.
- Snow K, Doud LK, Hagerman R, Pergolizzi RG, Erster SH, 32 Thibodeau SN. Analysis of a CGG sequence at the FMR-1 locus in fragile X families and in the general population. Am J Hum Genet 1993;53:1217-28.
- 33 Hiremath LS, Webb NR, Rhoads RE. Immunological detection of the messenger RNA cap-binding protein. *β* Biol Chem 1985;260:7843-9.
- Duncan R, Hershey JWB. Identification and quantification of levels of protein synthesis initiation factors in crude HeLa cell lysates by two-dimensional polyacrylamide gel electrophoresis. *J Biol Chem* 1983;**258**:7228-35.
- 35 Duncan R, Milburn SC, Hershey JWB. Regulated phosphorylation and low abundance of HeLa cell initiation factor eIF-4F suggests a role in translational control. *J Biol Chem* 1987;**262**:380-8.
- Wei CL, MacMillan SE, Hershey JWB. Protein synthesis initiation factor eIF-1A is a moderately abundant RNA-binding protein. *J Biol Chem* 1995;270:5764-71.
- binding protein. J Biol Chem 1995;210:3104-11.
 Erster SH, Brown WT, Goonewardena P, Dobkin CS, Jenkins EC, Pergolizzi RG. Polymerase chain reaction analysis of fragile X mutations. *Hum Genet* 1992;90:55-61.
 Williams LC, Hegde MR, Nagappan R, Bullock J, Faull RLM, Winship J, Snow K, Love DR. Null alleles at the Huntington disease locure implications for disenseries and 38
- Huntington disease locus: implications for diagnostics, and CAG repeat instability. *Genet Testing* 2000;4:55-60. Kozak M. Recognition of AUG and alternative initiator codons is augmented by G in position +4 but is not generally affected by the nucleotides in positions +5 and +6. *EMBO J* 1997;16:2482-92. 39

Non-invasive evaluation of arterial involvement in patients affected with Fabry disease

Pierre Boutouyrie, Stéphane Laurent, Brigitte Laloux, Olivier Lidove, Jean-Pierre Grunfeld, Dominique P Germain

EDITOR—Fabry disease (FD) (OMIM 301500) is an X linked recessive disease resulting from deficiency of the lysosomal hydrolase α -galactosidase A.¹ The enzymatic defect leads to the widespread deposition of uncleaved neutral glycosphingolipids in the plasma and lysosomes, especially in vascular endothelial and smooth muscle cells. Initial clinical signs include skin lesions (angiokeratoma), excruciating acral pain, and benign corneal opacities. Progressive glycosphingolipid deposition in the microvasculature of hemizygous males subsequently leads to failure of target organs and to ischaemic complications involving the kidneys, heart, and brain.^{2 3} Much interest is currently shown in emerging therapies for FD and recent studies have reported that genetic engineering has removed many of the obstacles to the clinical use of enzyme replacement and that infusions of purified a-galactosidase A are safe and biochemically active.45 However, clinical and laboratory indicators of benefit are lacking, given the slow course of the disease. This emphasises the need for non-invasive surrogate endpoints to delineate target organ damage and to monitor the efficacy of enzyme replacement therapies.

Methods and results

In the present study, we determined intimamedia thickness (IMT) at the site of the radial artery, a distal, muscular, medium sized artery, in a cohort of 21 hemizygous male FD patients, with a mean age of 32 years (SD 13, range 13-56 years), compared with 21 age and sex matched normal controls. All patients were diagnosed with FD by the presence of both clinical signs and a markedly decreased α -galactosidase A activity in leucocytes (<4 nmol/h/mg protein, normal values 25-55 nmol/ h/mg protein). No patient had end stage renal disease. Measurements of the radial artery parameters were obtained with a high precision echotracking device (NIUS 02, SMH, Bienne, Switzerland) as previously described.67 Briefly, the radiofrequency signal was visualised and the peaks corresponding to the blood-intima and media-adventitia interface were electronically tagged and followed over several cardiac cycles. Internal diameter and wall thickness were then measured with a precision of about 10 µm. Four to six measurements were averaged.⁶⁷ Radial artery IMT was measured 2 cm upstream from the wrist.

Compared to controls, FD patients had considerably higher IMT values at the site of the radial artery (fig 1). IMT was twice as high in

J Med Genet 2001;38:629-631

Department of Pharmacology and **INSERM EMIU 0107,** Hôpital Européen Georges Pompidou, 75015 Paris, France P Boutouyrie S Laurent **B** Laloux

Department of Nephrology, Hôpital Necker, Paris, France O Lidove I-P Grunfeld

Department of Genetics, Hôpital **Européen Georges** Pompidou, 20 rue Leblanc, 75015 Paris, France D P Germain

Correspondence to: Dr Germain, dominique.germain@ hop.egp.ap-hop-paris.fr



Figure 1 Correlation between radial artery intima-media thickness and age in patients with Fabry disease (circles) and in control subjects (triangles). Correlations are significant (p<0.001) in both populations and slopes differ significantly (59 (SD 14) v 25 (SD 4) µm per 10 years, p<0.001).

FD patients than in controls, even after adjustment for body surface area, age, and mean blood pressure (p<0.001). Radial artery IMT

A Control subject

increased significantly with age in each group. However the slope was 2.3-fold higher in FD patients than in controls (p<0.001) (fig 1).

Discussion

In the present study, we describe evidence of a major, accelerated hypertrophy of the wall of a medium sized artery in a cohort of patients with FD. The magnitude of the difference in radial artery IMT was very large, with virtually no overlap between FD patients and controls. With age, the radial artery wall thickening was 2.3-fold faster in FD patients than in controls. The high definition echotracking system used in the present study has been previously validated in large subsets of patients with various diseases, and its accuracy and reproducibility are well accepted.⁶⁷

The most commonly proposed explanation for the pathogenesis of cardiovascular lesions in FD patients is the slow deposition of uncleaved neutral glycosphingolipids within the arterial and cardiac tissues. However, the hypothesis of



Figure 2 Bidimensional scans and radiofrequency signals (RF) of the right radial artery from a control (A) and a patient with Fabry disease (B). Lumen and posterior wall contours have been emphasised. Intima-media thickness (IMT) was measured from the distance between the RF peaks corresponding to the blood-intima and media-adventitia interfaces. Note the irregularity and the prominent thickening of the arterial wall in the Fabry patient.

a sole lysosomal accumulation of sphingolipids is somewhat simplistic since in the most advanced reported cases of left ventricle hypertrophy in FD patients, the amount of uncleaved glycosphingolipids found in the cardiac tissue did not exceed 1.6% of tissue weight (10-20 mg/g wet weight).8 Other mechanisms are thus probably involved. First, although accumulation of globotriaosylceramide is the main mechanism in FD, the metabolism of other glycosphingolipids may also be disregulated.9 Among them, lactosylceramide, which mimics the biological function of cytokines, growth factors, and stress signalling molecules¹⁰¹¹ and accumulates in vascular tissues of FD patients,^{8 9} could act as a second messenger and potentiate the hypertrophy of the arterial wall. Second, the smaller internal diameter of the radial artery in FD patients may be the result not only of wall hypertrophy encroaching the lumen (fig 2), but also endothelial dysfunction. Deposition of glycosphingolipids occurs predominantly in the lysosomes of endothelial and smooth muscle cells, with consequent cellular dysfunction.3 An altered endothelium dependent relaxation of arterial smooth muscle could occur at the site of the radial artery or downstream, in arterioles, influencing the tonic flow dependent vasodilatation. The mechanism of flow dilatation is known to occur physiologically at the site of the radial and brachial arteries,¹² and has been related to changes in basal and stimulated nitric oxide (NO) release.¹² Finally, both in the media and intima, smooth muscle cells with glycosphingolipid inclusions secrete important quantities of extracellular matrix, notably elastic fibres.8 Proliferation of smooth muscle cells and extracellular matrix deposition may thus contribute to the hypertrophy of the radial artery observed in FD patients.

In conclusion, this study presents the first non-invasive demonstration of a major increase in arterial wall thickness at the site of the radial artery in a cohort of patients with confirmed FD. The assessment of the involvement of the large arteries, through non-invasive procedures, could prove useful in monitoring new therapies for FD in providing an intermediate phenotype or a surrogate marker. However, the prognostic significance of the radial artery wall hypertrophy and its ability to regress with

emerging treatments, such us enzyme replacement^{4 5 13 14} or gene therapy,¹⁵ remains to be determined during follow up studies.

This study received financial support from the Institut National de la Santé et de la Recherche Médicale (INSERM) and from Vaincre les Maladies Lysosomales (VML)

- 1 Brady RO, Gal AE, Bradley RM, Martensson R, Warshaw AL, Laster L. Enzymatic defect in Fabry's disea ceramide-trihexosidase deficiency. $N Engl \not = M$ Med 1967;276:1163-7. 2 Desnick RJ, Ioannou YA, Eng CM. α-galactosidase A
- deficiency: Fabry disease. In: Scriver CR, Beaudet AL, Sly WS, Valle D, Kinzler KE, Vogelstein B, eds. *The metabolic* and molecular bases of inherited diseases. 8th ed. New York: McGraw-Hill, 2001:3733-74. Germain DP. Fabry disease, Clinical and genetic aspects.
- Therapeutic perspectives. Rev Med Intern 2000;21:1086-103
- K. Schiffmann R, Murray GJ, Treco D, Daniel P, Sellos-Moura M, Myers M, Quirk JM, Zirzow GC, Borowski M, Loveday K, Anderson T, Gillespie F, Oliver KL, Jeffries NO, Doo E, Liang TJ, Kreps C, Gunter K, Frei K, Crutchfield K, Selden RF, Brady RO. Infusion of alpha-galactosidase A reduces tissue globotriaosylceramide storage in patients with Fabry disease. Proc Natl Acad Sci USA 2000;97:365-70
- 70.
 5 Eng CM, Cochat P, Wilcox WR, Germain DP, Lee P, Waldek S, Caplan L, Heymans H, Braakman T, Fitzpatrick MA, Huertas P, O'Callaghan MW, Richards S, Tandon PK, Desnick RJ. Enzyme replacement therapy in Fabry disease: results of a placebo-controlled phase 3 trial. Am J Hum Genet 2000;67:A134.
- Boutouyrie P, Bussy C, Lacolley P, Girerd X, Laloux B, Laurent S. Association between local pulse pressure, mean blood pressure and arterial remodeling. *Circulation* 1999; 100:1387-93.
- Girerd X, Mourad JJ, Acar C, Heudes D, Chiche S, Bruneval P, Mignot JP, Billaud E, Safar M, Laurent S. Noninvasive measurement of medium-sized artery intimamedia thickness in humans: in vitro validation. J Vasc Res 1994;**31**:114-20.
- 8 Elleder M, Bradova V, Smid F, Budesinsky M, Harzer K Kustermann-Kuhn B, Ledvinova J, Belohlavek, Kral V, Dorazilova V. Cardiocyte storage and hypertrophy as a sole manifestation of Fabry's disease. Report on a case simular ing hypertrophic non-obstructive cardiomyopathy. Virchows Arch A Pathol Anat Histopathol 1990;417:449-55.
 9 Desnick RJ, Blieden LC, Sharp HL, Hofshire PJ, Moller JH.
- Cardiac and valvular anomalies in Fabry disease. Clin morphologic and biochemical studies. Circulation 1976;54: 818-25
- 10 Chatterjee S. Sphingolipids in atherosclerosis and vascular biology. Arterioscler Thromb Vasc Biol 1998;18:1523-33. 11 Kolter T, Sandhoff K. Recent advances in the biochemistry
- I kolci I, Samino I, Recent advances an Interformation of sphingolipidoses. Brain Pathol 1998;8:79-100.
 Joannides R, Richard V, Haefeli WE, Linder L, Luscher TF, Thuillez C. Role of basal and stimulated release of nitric Animez C. Role of basa and sumfaced release of mittee oxide in the regulation or radial artery caliber in humans. *Hypertension* 1995;26:327-31.
 Ioannou Y, Zeidner K, Friedman B, Desnick R. Fabry
- disease: enzyme replacement therapy in a-galactosidase A deficient mice. Am J Hum Genet 2000;68:14-25.
 Eng CM, Guffon N, Wilcox WR, Germain DP, Lee P, Waldek S, Caplan L, Linthorst GE, Desnick RJ. A multicenter,
- randomized, double-blind, placebo-controlled study of the safety and efficacy of recombinant human α -galactosidase A replacement therapy in Fabry disease. N Engl J Med (in press).
- 15 Ziegler RJ, Yew NS, Li C, Cherry M, Berthelette P, Romanczuk H, Ioannou YA, Zeidner KM, Desnick RJ, Cheng SH. Correction of enzymatic and lysosomal storage defects in Fabry mice by adenovirus-mediated gene transfer. Hum Gene Ther 1999;10:1667-82.