

PAPER

Age associated axonal features in HNPP with 17p11.2 deletion in Japan

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Objective: To clarify age related changes in the clinicopathological features of hereditary neuropathy with liability to pressure palsy (HNPP) in Japanese patients with deletion of 17p11.2, particularly concerning axonal abnormalities.

Methods: Forty eight proband patients from 48 HNPP families were assessed as to clinical, electrophysiological, and histopathological features, including age associated changes beyond those in controls.

Results: Motor conduction studies showed age associated deterioration of compound muscle action potentials in nerves vulnerable to repetitive compression (median, ulnar, and peroneal nerves), but not in others such as the tibial nerve. Sensory conduction studies revealed more profound reduction of action potentials than motor studies with little age related change. Large myelinated fibre loss was seen in the sural nerve irrespective of age at examination.

Conclusions: Irreversible axonal damage may occur at entrapment sites in motor nerves in HNPP patients, progressing with aging. Sensory nerves may show more profound axonal abnormality, but without age association. The electrophysiological features of HNPP are presumed to be a mixture of abnormalities occurring from early in life and acquired features caused by repetitive insults at entrapment sites. Unlike Charcot-Marie-Tooth disease type 1A, age associated axonal damage may not occur unless the nerves are subjected to compression.

Hereditary neuropathy with liability to pressure palsy (HNPP) is an autosomal dominant disorder characterised by recurrent transient nerve palsies associated with compression at the typical anatomic sites of potential nerve entrapment.^{1,2} Tomacula, which represent focal thickening of the myelin sheath, characteristically are seen in both sensory and motor nerves in HNPP.^{3–6} This disorder usually is associated with a 1.5 Mb deletion of locus 17p11.2, which contains the gene for peripheral myelin protein 22 (PMP22).^{7–9} HNPP therefore appears to represent a reciprocal product of Charcot-Marie-Tooth disease type 1A (CMT1A), which is associated with duplication of PMP22.¹⁰ PMP22 is an important factor for regulation of Schwann cell proliferation and apoptosis.¹¹ As the Schwann cell plays an important role in maintenance of the axon, axonal loss associated with demyelination has been reported to occur in patients with CMT1A.^{12–15} Age associated reduction of compound muscle action potential (CMAP) amplitude resulting from large-axon loss has been reported in CMT1A¹⁵ and is closely related to clinical manifestations and functional impairment.^{14,15}

In Western countries, the clinical and electrophysiological features of HNPP have been described on a large scale.^{16–20} Characteristic electrophysiological findings are multifocal slowing of conduction at sites of entrapment, prolonged distal latency (DL), mild slowing of motor nerve conduction velocity (MCV), and diffuse abnormality of sensory nerve conduction velocity (SCV).^{16–18–20} However, there have been no similar large scale investigations of the clinical and electrophysiological features of HNPP in Asian subjects. Furthermore, it has not been clarified whether electrophysiological and histopathological abnormalities, particularly axonal features, worsen with aging in HNPP as happens in CMT1A.

The present investigation was carried out in Japan and we studied HNPP including its electrophysiological and histopathological features, especially in relation to aging.

METHODS

Patients and DNA diagnosis

An HNPP survey was conducted by the study group for hereditary neuropathy in Japan under the auspices of the Ministry of Health, Labor, and Welfare.^{15,21} A total of 48 proband patients from 48 HNPP families, whose 17p11.2 deletion was confirmed, were investigated. The mean age (SD) of the patients at examination was 41.8 (18.5) years (table 1). All subjects underwent clinical examination by at least one neurologist. Patients with chronic alcoholism or vitamin deficiency were not included. Four patients manifested mild glucose intolerance. To confirm the diagnosis of HNPP, DNA analyses for the presence of a chromosome 17p11.2–12 deletion, which includes a 1.5 Mb region containing the PMP22 gene between CMT1A-REP repeats, were performed in all patients. For most patients these analyses were performed at the Department of Neurology at Nagoya University Graduate School of Medicine as described previously,²² while DNA was analysed at other institutions for the rest. The characteristic deletion in HNPP was detected by Southern analysis, probing with PMP22 cDNA, and CMT1A-REP fragments as described previously.^{22–24} Hybridisation with PMP22 cDNA and pNEA102, pHK1.0P, and pHK5.2 probes, which map within the CMT1A-REP, was carried out

Abbreviations: CMAP, compound muscle action potential; CMT1A, Charcot-Marie-Tooth disease type 1A; DL, distal latency; HNPP, hereditary neuropathy with liability to pressure palsy; MCV, motor nerve conduction velocity; PMP22, peripheral myelin protein 22; SCV, sensory nerve conduction velocity; SNAP, sensory nerve action potential

to determine the gene dose of the 1.5 Mb region containing PMP22. Deletion of one copy of the PMP22 gene, compared to the presence of two copies in normal controls, was genetically identified as HNPP. Informed consent was obtained in all patients, and the study as a whole was approved by the Ethics Committee of Nagoya University Graduate School of Medicine.

Electrophysiological study

Motor and sensory conduction was measured in the median, ulnar, tibial, peroneal, and sural nerves, using a standard method with surface electrodes for stimulation and recording.^{25, 26} Motor conduction was investigated in the median, ulnar, tibial, and peroneal nerves, recording from the abductor pollicis brevis, abductor digiti minimi, abductor hallucis brevis, and extensor digitorum brevis muscles, respectively. The following nerve segments were used for calculating MCV: wrist to elbow for the median nerve, wrist to distally at the elbow for the ulnar nerve, ankle to popliteal fossa for the tibial nerve, and ankle to distally at the fibular head for the peroneal nerve. Sensory conduction was investigated in the median, ulnar, and sural nerves, using antidromic recording from ring electrodes at the second and fifth digit for the median and ulnar nerves respectively, and bar electrodes at the ankle for the sural nerve. SCV was calculated for the distal segment. Amplitudes of CMAP and sensory nerve action potential (SNAP) were measured from the baseline to the first negative peak. Waveforms also were analysed to assess temporal dispersion. For motor nerves, we measured duration from the onset to the first crossing of the baseline in the CMAP.²⁷ For sensory nerves, duration from the onset of the SNAP to the first negative peak rather than to the first crossing of the baseline was measured to avoid artefacts from overlapping muscle action potentials.²⁵ This was necessary because some motor axons have thresholds similar to those of large myelinated sensory axons, resulting in superimposition on the SNAP that modifies the waveform, especially when abnormal nerves are examined.^{28, 29} Because of a delay at the neuromuscular junction, the initial phase of the waveform of SNAP is less likely to be affected by muscle action potentials than the later phase.²⁹

Control values were obtained in 171 normal volunteers (51.0 (SD 16.3) years of age; male:female, 89:82) for the median nerve, 170 (51.2 (SD 16.4) years of age; male:female,

88:82) for the ulnar nerve, 161 (51.8 (SD 16.6) years of age; male:female, 85:76) for the tibial nerve, 171 (54.2 (SD 16.7) years of age; male:female, 92:79) for the peroneal nerve, and 163 (52.2 (SD 16.7) years of age; male:female, 85:78) for the sural nerve.

Histopathological study

Sural nerve biopsy was performed in 14 patients as described previously.^{30, 31} Informed consent was obtained beforehand. Specimens were divided into two portions. The first portion was fixed in 2.5% glutaraldehyde in 0.125 M cacodylate buffer (pH 7.4) and embedded in epoxy resin for morphometric study. The density of myelinated fibres was assessed in toluidine blue stained semithin sections using a computer assisted image analyser (Luzex FS; Nikon, Tokyo, Japan) to calculate the densities of small and large myelinated fibres as described previously.³²⁻³⁴ A fraction of the glutaraldehyde fixed sample was processed for a teased fibre study, in which at least 100 single fibres were isolated; their pathologic condition was assessed microscopically according to criteria described previously.^{32, 35} The second portion of the specimen was fixed in 10% formalin solution and embedded in paraffin. Sections were cut by routine methods and stained with haematoxylin and eosin as well as by the Klüver-Barrera and Masson trichrome methods. Control values were obtained from 13 autopsy cases in which patients died of non-neurologic diseases (48.5 (SD 23.5) years of age; male:female, 7:6). Specimens were processed in the same manner as for HNPP patients.

Statistical analysis

Quantitative data are presented as the mean (SD) and were compared with control values using the Mann-Whitney U test. To determine the relationship of electrophysiological and histopathological indices and age at examination, Pearson's correlation coefficient analysis was carried out. To determine whether worsening of these indices in HNPP patients with aging was significantly greater than in normal controls, regression slopes of patient and control groups were compared. Values of *p* less than 0.05 were considered to indicate significance.

RESULTS

Clinical features

The age at first awareness of neuropathic symptoms in the 48 probands was 33.1 (SD 19.3) years (table 1). The male:female ratio was 38:10. An obvious family history of recurrent transient nerve palsies was present for 24 patients (50%). Only one patient (2%) reported athletic impairment during childhood. Deformity in the distal part of the lower limbs such as hammer toe or pes cavus was present in two patients (4%). Atrophy was noted in the leg in six patients (13%). The pattern of neuropathic symptoms was multiple mononeuropathy associated with recurrent transient nerve palsies in 41 patients (85%), while the other seven (15%) manifested mainly a symmetric polyneuropathy pattern. A history of transient nerve palsy was noted in the median, ulnar, radial, and peroneal nerves in 11 (23%), 18 (38%), seven (15%), and 29 (60%) patients, respectively. Signs of brachial plexus palsy were reported in 10 (21%). With respect to the activities of daily living, all patients were non-disabled or only mildly disabled, except for two (4%) who became unable to walk.

Electrophysiological features

Motor conduction studies showed variable degrees of abnormality in individual nerves (table 2). For the median nerve, MCV was significantly slowed compared to normal controls ($p < 0.0001$). This slowing of MCV was present regardless of age at examination, and there was no

Table 1 Characteristics of 48 Japanese HNPP probands with deletion of 17p11.2-12

Clinical features	n (%)
Age at onset, years	33.1 (SD 19.3)
Age at examination, years	41.8 (SD 18.5)
Men/women	38/10
Family history	24 (50%)
Athletic impairment during childhood	1 (2%)
Pes cavus or hammer toe	2 (4%)
Atrophy in the legs	6 (13%)
Pattern of neuropathy	
Multiple mononeuropathy	41 (85%)
Symmetric polyneuropathy	7 (15%)
History of transient nerve palsy	
Median nerve	11 (23%)
Ulnar nerve	18 (38%)
Radial nerve	7 (15%)
Peroneal nerve	29 (60%)
Brachial plexus	10 (21%)
Activity of daily living	
Able to walk	46 (96%)
Unable to walk	2 (4%)
Bedridden	0

Age at onset, age at first awareness of neuropathic symptoms; Family history, obvious family history of recurrent transient nerve palsies.

significant difference in regression slopes in the correlation between MCV and age at examination (regression slope -0.073 for HNPP ν -0.064 for controls). DL was very prolonged (179% of controls) and prolongation tended to worsen as age at examination increased ($r = 0.47$). The CMAP was reduced to various degrees in most patients and showed further reduction with advancing age ($r = -0.67$; fig 1). Worsening of both DL and CMAP with age was significantly more prominent than in controls, as evident from comparison of regression slopes ($p < 0.0001$ and < 0.01 , respectively).

For the ulnar nerve, mild to moderate slowing of MCV and prolongation of DL were noted regardless of age at examination, while CMAP decreased with advancing age ($r = -0.65$). CMAP diminution with aging was significantly worse in patients than in controls (regression slope -0.109 for HNPP ν -0.021 for controls; $p < 0.0001$). For the tibial nerve, slowing of MCV and prolongation of DL also were mild to moderate in most patients of all ages. Reduction of CMAP was also present in all ages examined but, in contrast to other nerves, the relationship of reduction to aging was indistinguishable from that in controls (regression slope -0.062 for HNPP ν -0.069 for controls). For the peroneal nerve, the age associated decrement in CMAP was significantly greater than in controls ($p < 0.05$). Slowing of MCV and prolongation of DL were present in patients of all ages, but no significant worsening with aging was seen in comparison with controls.

As for sensory conduction studies, slowing of conduction velocity was present as in motor nerves. SCV of the median nerve tended to slow with increasing age at examination

($r = -0.41$). This age associated worsening was significantly greater than in controls ($p < 0.05$), while SCV of the ulnar and sural nerves did not show a correlation with age. Reduction of SNAP was conspicuous in the median (24% of control amplitude), ulnar (28%), and sural (42%) nerves. Age associated reduction of SNAP was seen in the median ($r = -0.50$), ulnar ($r = -0.45$), and sural ($r = -0.37$) nerves, but the rate of change was not worse than in controls.

Duration of CMAP and SNAP was prolonged in all nerves examined compared to normal controls, suggesting the presence of temporal dispersion.²⁷ Compared to controls, significant age associated worsening was seen only in the SNAP of the median nerve ($p < 0.0001$).

Histopathological features

Average total myelinated fibre density in patients' sural nerves was mildly, but not significantly, reduced compared to normal controls (7738 (SD 1253) ν 8561 (SD 1289) fibers/mm²; table 3). The density of large myelinated fibres was significantly reduced from that in controls (2458 (SD 730) ν 3258 (SD 736) fibers/mm²; $p < 0.01$) but that of small myelinated fibres was not (5280 (SD 1025) ν 5302 (SD 655) fibers/mm²). Axonal sprouting was not conspicuous in any case. Although the density of large myelinated fibres decreased as age at examination increased ($r = -0.70$), the rate of reduction was indistinguishable from that in controls (regression slope -27.1 for HNPP ν -26.0 for controls) because large myelinated fibres were reduced even at younger ages. Teased fibre preparations revealed frequent tomacular change (41.5% (SD 15.8%)). The frequency of segmental

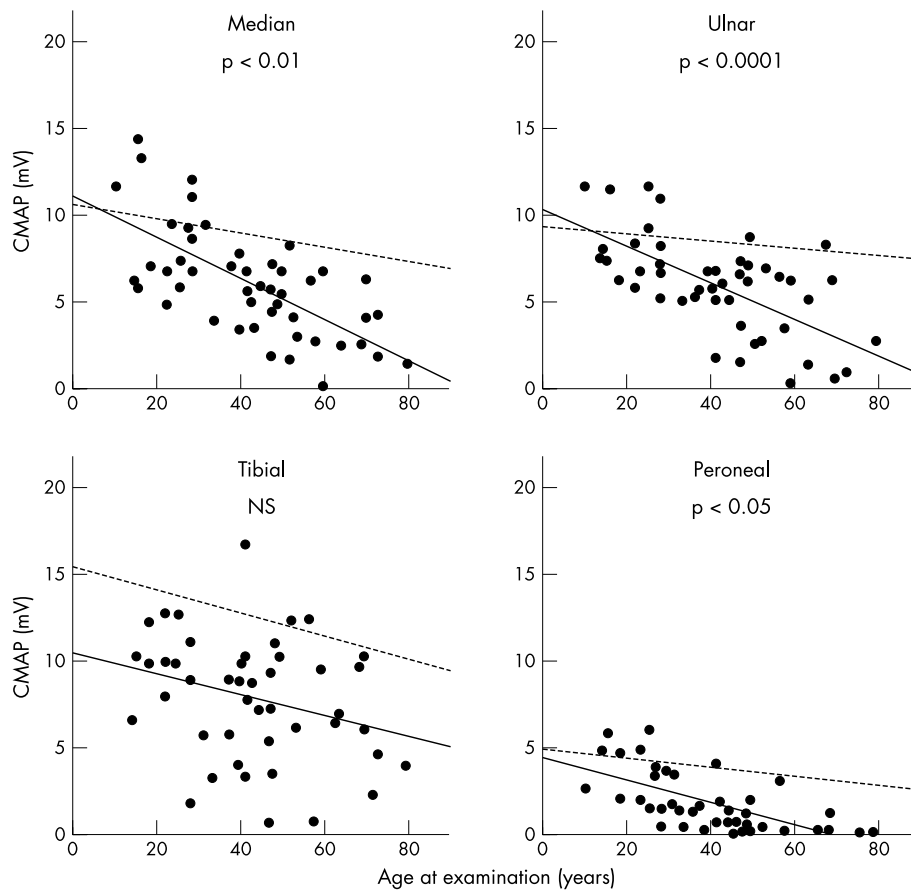


Figure 1 Correlation between CMAP and age at examination in HNPP patients and normal controls. Filled circles represent indices in HNPP patients, bold lines represent regression lines for HNPP patients, and broken lines represent regression lines for normal controls. Comparing regression slopes of normal controls and HNPP patients, CMAP of the median, ulnar, and peroneal nerves, but not the tibial nerve, in HNPP patients were significantly more reduced with increasing age at examination.

Table 2 Nerve conduction studies

	HNPP							Controls		
	Nerve conduction measures				Correlation to aging			Correlation to aging		
	n	Mean (SD)	% of controls	p Values for controls*	r †	Regression slope	p Values for controls‡	Mean (SD)	r †	Regression slope
Motor conduction										
Median nerve										
MCV (m/s)	47	46.0 (5.3)	80	<0.0001	-0.25	-0.073	NS	57.6 (3.8)	-0.27	-0.064
DL (ms)	47	6.1 (1.8)	179	<0.0001	0.47	0.046	<0.0001	3.4 (0.4)	0.19	0.005
CMAP (mV)	48	6.3 (3.2)	77	<0.0001	-0.67	-0.122	<0.01	8.2 (2.9)	-0.24	-0.042
Duration (ms)	32	5.4 (0.8)	115	<0.0001	0.13	0.006	NS	4.7 (0.9)	-0.07	-0.004
Ulnar nerve										
MCV (m/s)	47	46.9 (8.3)	81	<0.0001	0.04	0.018	NS	58.0 (4.6)	-0.22	-0.062
DL (ms)	47	3.8 (0.8)	146	<0.0001	0.17	0.009	NS	2.6 (0.3)	0.06	0.001
CMAP (mV)	48	6.0 (3.0)	81	<0.0001	-0.65	-0.109	<0.0001	7.4 (1.8)	-0.20	-0.021
Duration (ms)	28	5.9 (1.2)	116	<0.0001	-0.22	-0.016	NS	5.1 (0.7)	-0.01	-0.001
Tibial nerve										
MCV (m/s)	45	39.6 (4.5)	86	<0.0001	-0.02	-0.006	NS	46.0 (3.8)	-0.34	-0.079
DL (ms)	45	5.5 (1.3)	138	<0.0001	0.15	0.011	NS	4.0 (0.6)	0.11	0.004
CMAPs (mV)	45	7.9 (3.7)	67	<0.0001	-0.29	-0.062	NS	11.8 (3.5)	-0.33	-0.069
Duration (ms)	25	5.7 (1.3)	114	<0.01	-0.18	-0.012	NS	5.0 (0.7)	-0.17	-0.008
Peroneal nerve										
MCV (m/s)	38	35.7 (5.7)	76	<0.0001	-0.11	-0.042	NS	47.4 (4.5)	-0.38	-0.101
DL (ms)	38	7.7 (2.3)	167	<0.0001	-0.002	-0.00004	NS	4.6 (1.1)	0.04	0.002
CMAP (mV)	41	1.9 (1.8)	56	<0.0001	-0.65	-0.067	<0.05	3.4 (2.0)	-0.22	-0.027
Duration (ms)	16	6.4 (0.9)	131	<0.0001	-0.09	-0.006	NS	4.9 (0.9)	-0.17	-0.009
Sensory conduction										
Median nerve										
SCV (m/s)	42	38.6 (10.1)	69	<0.0001	-0.41	-0.235	<0.05	56.3 (5.3)	-0.26	-0.085
SNAP (µV)	48	6.8 (6.2)	24	<0.0001	-0.50	-0.178	NS	28.0 (11.5)	-0.45	-0.327
Duration (ms)	26	0.9 (0.4)	150	<0.0001	0.56	0.011	<0.0001	0.6 (0.1)	-0.11	-0.001
Ulnar nerve										
SCV (m/s)	41	36.8 (8.4)	68	<0.0001	-0.13	-0.069	NS	54.5 (5.5)	-0.28	-0.093
SNAP (µV)	48	6.6 (6.4)	28	<0.0001	-0.45	-0.170	NS	23.8 (10.3)	-0.37	-0.240
Duration (ms)	26	0.9 (0.2)	150	<0.0001	0.08	0.001	NS	0.6 (0.1)	-0.05	-0.00004
Sural nerve										
SCV (m/s)	43	36.4 (6.9)	74	<0.0001	-0.13	-0.052	NS	49.2 (4.8)	-0.12	-0.035
SNAP (µV)	48	7.1 (5.9)	42	<0.0001	-0.37	-0.124	NS	16.8 (7.8)	-0.38	-0.177
Duration (ms)	21	0.9 (0.3)	129	<0.05	0.23	0.004	NS	0.7 (0.1)	0.21	0.002

*Mann-Whitney U test; †Pearson's correlation coefficient; ‡regression slopes of HNPP and controls were compared. Control values were obtained in 171 normal volunteers for the median nerve, 170 for the ulnar nerve, 161 for the tibial nerve, 171 for the peroneal nerve, and 163 for the sural nerve. CMAP, compound muscle action potential; DL, distal latency; Duration, duration from the onset to the first crossing of the baseline in the CMAP and duration from the onset of the SNAP to the first negative peak; MCV, motor nerve conduction velocity; NS, not significant; SCV, sensory nerve conduction velocity; SNAP, sensory nerve action potential.

de/re-myelination also was significantly high (25.6% (SD 13.9%), $p < 0.001$). Axonal degeneration was slightly increased (3.6% (SD 3.8%)) and was seen even in young patients in contrast to controls.

DISCUSSION

This study demonstrated clinical, electrophysiological, and histopathological features of Japanese HNPP patients with the 17p11.2 deletion. Although recurrent transient nerve

palsies are the characteristic feature of this disease, a minority of patients showed a symmetric polyneuropathy pattern, as previously reported.^{16-18 36} Electrophysiological features of slowing of conduction velocities and varying degrees of abnormality among individual nerves, agreed well with previous reports of Western populations.¹⁶⁻²⁰ Slowing of MCV in our series seemed more marked than in previous reports.^{16-18 20} The fact that we only examined probands of HNPP families and did not include affected siblings could

Table 3 Histopathological study of the sural nerve

	HNPP (n = 14)			Controls (n = 13)				
	Mean (SD)	p Values for controls*	Correlation to aging			Mean (SD)	Correlation to aging	
			r †	Regression slope	p Values for controls‡		r †	Regression slope
Myelinated fibre density (no./mm²)								
Total	7738 (1253)	NS	-0.45	-29.6	NS	8561 (1289)	-0.73	-39.9
Large	2458 (730)	<0.01	-0.70	-27.1	NS	3258 (736)	-0.83	-26.0
Small	5280 (1025)	NS	-0.05	-2.5	NS	5302 (655)	-0.50	-13.9
Teased fibre study (%)								
Tomacular change	41.5 (15.8)	-	-0.21	-0.18	-	-	-	-
Segmental de/re-myelination	25.6 (13.9)	<0.001	0.39	0.30	NS	6.9 (6.5)	0.82	0.22
Axonal degeneration	3.6 (3.8)	NS	-0.35	-0.07	<0.05	1.6 (1.8)	0.81	0.06

*Mann-Whitney U test; †Pearson's correlation coefficient; ‡regression slopes of HNPP and controls were compared. NS, not significant.

account for the difference, or greater slowing might be characteristic of Japanese patients. In the peroneal nerve, it seems that the amplitude of CMAP is lower and the distribution of DL is wider than in Western populations even in normal controls.²⁶ Japanese people usually sit on the floor at home, rather than on chairs, and sometimes sit with their legs folded underneath them. This traditional Japanese sitting position may induce peroneal nerve injury.

A striking finding in our study was a reduction in CMAP with increasing age at examination. This feature was observed in the median, ulnar, and peroneal nerves but not in the tibial nerve. The median nerve passes through the carpal tunnel, predisposing it to entrapment injury, while the ulnar and peroneal nerves are vulnerable to repetitive compression injury at the cubital tunnel and fibular head, respectively, as suggested by the high frequency of episodic palsy of these nerves compared with the tibial nerve. Repetitive movement and nerve stretching at these sites also may contribute to injury. Thus, individual nerve-specific CMAP reduction with increasing age probably resulted from the cumulative effects of repetitive damage; conduction slowing caused by demyelination would be prominent at entrapment sites, as previously reported.^{16–18, 20} In the present study, demyelination also showed progression over time as demonstrated by age associated prolongation of DL and SCV in the median nerve for conduction through the entrapment site. However, in the ulnar and peroneal nerves, where electrophysiological indices were recorded distally from sites vulnerable to compression, no age associated worsening of MCV, SCV, or DL was observed, suggesting that myelin abnormality distal to the entrapment site does not worsen with advancing age. Thus, CMAP reduction in the median, ulnar, and peroneal nerves would reflect secondary axonal involvement complicating demyelination at the entrapment site. This age associated axonal involvement in a primarily demyelinating condition is similar to that observed in CMT1A with PMP22 duplication.^{12, 14, 15} However, unlike CMT1A, axonal damage may not occur unless the nerves are subjected to compression. PMP22 duplication in Schwann cells results in disturbance of axonal cytoskeletal organisation, resulting in distal axonal degeneration and fibre loss.¹³ However, the effect of PMP22 deletion on the axonal cytoskeleton is less severe.¹³ PMP22 deletion in itself may not cause progressive axonal involvement associated with aging, though compression induced demyelination may elicit secondary axonal loss because of deficient Schwann cell signalling to the axonal cytoskeleton.³⁷

SNAP of the median, ulnar, and sural nerves showed marked reduction even in nerves relatively free from compression and tended to decrease with increasing age at examination. Unlike findings for CMAP, however, rates of reduction with aging did not differ significantly from those in normal controls. Sensory axons may be less susceptible than motor nerves to changes caused by entrapment.

Reduction in CMAP and SNAP may be at least partly attributed to dispersion with phase cancellation as a result of demyelinating change, as suggested by significant prolongation of waveform duration.^{27, 38} Sural nerve biopsy specimens showed a reduction in large myelinated fibre density irrespective of age, which may indicate a developmental abnormality of axons or a loss of axons relatively early in life. This axonal loss also may contribute to reduction in amplitudes. At any rate, reduction in myelinated fibres of sensory nerves in HNPP patients did not appear to be associated with acquired damage at the entrapment sites. Thus, the electrophysiological features of HNPP are a mixture of abnormalities occurring from an early stage in life and acquired features caused by repetitive insults at entrapment sites. One therapeutic strategy in HNPP patients may be

directed toward prevention of axonal damage associated with entrapment.

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