ELECTRONIC REVIEW

Biomarkers of exposure to polycyclic aromatic hydrocarbons from environmental air pollution

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Metabolites of pyrene and DNA adducts have been used as biomarkers of high level exposure to polycyclic aromatic hydrocarbons (PAHs). A systematic review was performed to evaluate whether these biomarkers are also valid markers of low level environmental exposure to PAHs. Thirty five studies were identified with more than 10 subjects that evaluated environmental air pollution to PAHs in relation to metabolites of PAHs, mainly hydroxypyrene (1-OHP), PAH-DNA adducts, or protein adducts. PAH metabolites and, to a less extent, PAH-DNA adducts correlated well at the group level with exposure to B(a)P even at low levels of air pollution. The use of these biomarkers should be more widely implemented in combination with more traditional techniques for the assessment of general population exposure to PAHs from ambient air pollution.

> **P**olycyclic aromatic hydrocarbons (PAHs) are a group of organic chemicals characterised by chemical stability, low volatility, and low solubility in water. PAHs are formed during the incomplete combustion of organic substances, are widespread in the environment, and typically occur in mixtures. Health damage associated with PAH exposure has been evaluated repeatedly by different health and environmental protection agencies, such as the International Agency for Research on Cancer (IARC), the Environmental Protection Agency (EPA), the National Toxicology Program (NTP), and the Agency for Toxic Substances and Disease Registry (ATSDR). Seven compounds, including benzo(a)anthracene, benzo(b)fluoranthene, benzo(j)fluoranthene, benzo(k)fluoranthene, benzo(a)pyrene, dibenz(a,h)anthracene, and indeno(1,2,3-c,d)pyrene, are considered by several agencies as human carcinogens.

> The main sources of human exposure to PAHs are occupation, passive and active smoking, food and water, and air pollution.¹ The total intake of carcinogenic PAHs in the general population has been estimated to be 3 μ g/day.² Levels of occupational exposure of benzo(a)pyrene (B(a)P), which is one of the main PAH compounds, vary widely in different industrial activities and job titles, ranging from 0.1 to 48 000 ng/m³.³⁻⁵ In smokers, B(a)P levels range from 0.5 to 7.8 μ g/100 cigarettes when exposure is from mainstream smoke and from 2.5 to 19.9 μ g/100 cigarettes when it comes from

side-stream smoke. Levels from passive smoking are lower, ranging from 0.0028 to 0.76 μ g/ m³ of B(a)P.6 Beside occupational exposure, dietary intake seems the most important source of PAHs in non-smokers; in the USA, dietary intake of total PAHs has been estimated to be almost 2 µg/kg food,⁶ with high levels of B(a)P found especially in charcoaled meat (8 μ g/kg), while in a variety of other food products B(a)P levels from 0.09 to 30 μ g/kg^{6 7} have been reported. Levels in water may range from 0 to 13 μ g/l B(a)P. There is a high variation in atmospheric PAHs levels across geographical areas with B(a)P concentrations ranging from 0.01 to 100 ng/m³ B(a)P).⁸ Airborne PAHs are usually analysed by gas chromatography/mass spectrometry9 10 or high performance liquid chromatography,11-14 mostly from particles collected in a filter after extraction with organic solvents.

Pollution of air by PAHs is mainly due to the incomplete combustion of wood or fuel used for residential heating and industrial or motor vehicle exhaust.¹¹ The level of exposure to PAHs through this source is low compared to other sources such as diet, occupation, or tobacco smoke. The half-life of airborne PAHs is of the order of days but can be longer when they are bound to small particles.² In the cities where residential heating is the main source of PAHs, air pollutant PAH levels during winter are frequently higher than in summer.^{15–17} Nevertheless, some studies that found very low levels of B(a)P (range 0.03–3.3 ng/m³) did not show a seasonal variation.¹¹

Inhaled PAHs are absorbed mainly through the bronchial epithelium. Absorption can be influenced by the size of the particles, with a slower clearance of those adsorbed on particles compared to PAHs in pure crystals.18 19 After absorption, PAHs are distributed to tissues where they are biotransformed by phase I metabolic enzymes to chemically reactive intermediates, that may bind covalently to DNA (DNA adducts), give rise to mutation and, eventually, tumour initiation. Furthermore, PAH metabolites are mostly conjugated with glucuronic acid by phase II enzymes and excreted as hydroxylated metabolites or in a small proportion as sulphate or even unconjugated. PAHs are excreted mainly through the faeces; only about 10% are excreted in the urine.

Abbreviations: B(a)P, benzo(a)pyrene; PAH, polycyclic aromatic hydrocarbon

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Main messages

- Existing biomarkers for environmental exposure to PAHs correlate well at group level, with exposure to B(a)P even at low levels of air pollution.
- This association is stronger for PAH metabolites, particularly hydroxypyrene, but is also observed for PAH-DNA adducts.
- The use of these biomarkers when assessing environmental exposure at the individual level is limited.

The difficulty of finding an index substance arises from the fact that the composition of PAH mixtures depends on the combustion source. The most common compound used as a reference substance for carcinogenic PAHs is benzo(a)pyrene (B(a)P). B(a)P was the first PAH recognised to display carcinogenic activity; more importantly, it has a high carcinogenic potency and is present in all environmental media.

The most commonly used biomarkers of PAH exposure are metabolites of PAHs, particularly 1-hydroxypyrene, and PAH-DNA or protein adducts. 1-Hydroxypyrene (1-OHP) is the principal product of pyrene metabolism, representing 90% of its metabolites.²⁰ Following inhalation, the half-life of 1-OHP is on average about 18-20 hours.²¹⁻²³ Pyrene is the only known precursor of 1-OHP;²⁴ it forms a consistent proportion of higher molecular weight PAHs in the environment.²⁵ The two main analytical methods employed to measure 1-OHP are high performance liquid chromatography combined with fluorescence detection (HPLC/FD), and gas chromatography with mass spectrometry (GC/MS). For HPLC/FD, the detection limit is 0.05 μ g/l, with a recovery rate of 85%,²⁶ and for GC/MS it is around 0.1 µg/l with a higher recovery rate that reaches more than 93%.27 PAH adducts have been mainly employed as a measure of PAHs linked to DNA in target tissues (biological effective dose), although protein adducts have also occasionally been determined. The half-life of DNA adducts in lymphocytes is in the order of months, while for protein adducts it depends on the type of protein where the chemical is bound. In the case of albumin adducts, the halflife is around 20 days, while for haemoglobin adducts it is around 120 days. ELISA is used to measure specific DNA adducts, and ³²P-postlabelling is used for the determination of bulky DNA adducts, also called aromatic adducts. Protein adducts are measured mostly in blood proteins using CG/MS, HPLC, or ELISA.

Studies examining PAH exposure through smoking or in the occupational environment have shown that at high levels 1-OHP and DNA adducts correlate well with B(a)P concentrations,²⁸ whereas there exists less evidence on the correlation of PAH or B(a)P concentrations⁵ with protein adducts. In this review we evaluate whether these biomarkers are valid markers of low level environmental exposure to PAHs.

METHODS

Selection of studies

We reviewed all the studies which measured 1-OHP or adducts as biomarkers of exposure to PAHs in populations not occupationally exposed to these agents. We did not include studies on occupation, active and passive smoking, or diet since there already exist several recent reviews⁵ ²⁸ ²⁹ and also because the level of exposure is too high to be comparable with the levels of PAHs in air. A search in the electronic database PubMed for all papers published from 1980 to 2002 on PAHs biomarkers was conducted. The keywords used were "(benzopyrene OR pah OR pahs OR

Policy implications

The use of biomarkers of exposure to environmental levels of PAHs should be more widely implemented in combination with more traditional techniques for the assessment of general population exposure to PAHs from ambient air pollution.

pyrene) AND (biological monitoring OR biomonitoring OR biological markers OR biomarker OR biomarkers)". In addition, we checked our personal archives and examined the references of the papers identified.

In the first search, we found 723 articles that included both animal and human studies. We reviewed the abstracts and primarily selected 122 papers, after excluding animal studies, reviews, and human studies with less than 10 subjects. We further excluded studies on occupationally exposed subjects and two studies on environmental exposure to PAHs, because DNA adduct levels were presented in units non-comparable to those of other studies.^{17 30} Furthermore, five studies on 1-OHP were not included: two measured B(a)P metabolites instead of 1-OHP,^{31 32} two reported 1-OHP levels in units not comparable to the rest of the studies,^{33 34} and the fifth study focused on intra- and inter-individual variability of 1-OHP.35 Finally, we included 36 studies of DNA and/or protein adducts or metabolites of PAHs that referred to exposure to PAHs through environmental air pollution.^{3 7–9 11–13 15–17 27} In the selected studies, biomarker levels were presented for non-smokers or were adjusted for smoking, except for two studies that did not indicate if results were adjusted,12 38 and another study49 that did not adjust for smoking habits. Among the 19 studies analysing 1-OHP, 12 included nonsmoking individuals. Eight studies on DNA adducts recruited only non-smokers, and five also had determinations on protein adducts.

Statistical analysis

We estimated between study differences and correlated mean biomarker levels with mean levels of B(a)P or the sum of eight carcinogenic PAHs in air (including benzo(a)anthracene, benzo(b)fluoranthene, benzo(k)fluoranthene, chrysene, benzo(a)pyrene, dibenzo(ah)anthracene, indeno(1,2,3-cd)pyrene, and benzo(ghi)perylene). The Pearson coefficient was estimated after log transformation of the mean values of benzo(a)pyrene in air and biomarker levels. Within study differences were also evaluated by comparing the mean biomarker levels in each study between exposed and unexposed (or less exposed) subjects. We did not perform a formal meta-analysis because studies were heterogeneous and did not allow a joint statistical evaluation. Heterogeneity between studies was tested through the Kruskal-Wallis equality of populations rank test that evaluates the hypothesis that several samples originate from similar populations. The p values of the *t* test were p = 0.028 for 1-OHP and p = 0.007 for DNA adducts, indicating that the levels of biomarkers in the populations evaluated were different. The application of statistical techniques to adjust for interlaboratory variation in the adducts analysis, such as the calculation of the "frequency ratio" that models the within study differences in adducts,^{29 62} was not feasible due to the small number of studies and the heterogeneity in design between studies.

RESULTS 1-Hydroxypyrene

We identified 19 studies evaluating 1-OHP in relation to air pollution. Fourteen had environmental measurements of the

Table 1 Studies with B(a)P measurements in air and 1-OHP

		Type of PAHs		PAH analysis	Smoking	1-OHP	
Author, year	Country	sampled	PAH monitoring	technique	status	analysis	Season of sampling
Zhao, 1992	China	B(a)P	Personal	HPLC/FD	NS	HPLC/FD	Winter +summer
Kanoh, 1993	Japan	B(a)P	Stationary	n.a.	NS	HPLC/FD	Winter + summer
Hemminki, 1994	Sweden	_	-	_	NS	HPLC/FD	Winter
Ovrebo, 1995	Poland	B(a)P	Personal	GC/MS	All	HPLC/FD	Winter + summer
Gundel, 1996	Germany	B(a)P	Stationary	n.a.	NS	HPLC/FD	Winter + summer
Cerna, 1997	Czech	B(a)P	Stationary	n.a.	NS	HPLC/FD	Winter
	Republic		,				
Gilbert and Viau, 1997	Canada	B(a)P	Stationary	n.a.	NS	HPLC/FD	Summer
Hara, 1997	Japan	B(a)P	Personal	HPLC/FD	NS	HPLC/FD	Summer
Vyskocyl, 1997	Czech Rep	B(a)P	Stationary	HPLC/FD	NS	HPLC/FD	Winter + summer
Poirier, 1998	Germany,	B(a)P	Stationary	GC/MS	All	HPLC/SFS	Summer
	Kuwait		,				
Merlo, 1998	Italy	B(a)P	Personal	HPLC/FD	NS	HPLC/FD	Summer
Motykiewicz, 1998	Poland	B(a)P	Stationary	GC	NS	HPLC/FD	Summer
Chuang, 1999	USA	B(a)P	Stationary	GC/MS	NS	GC/MS	Winter + summer
Pastorelli, 1999	Italy	Pyrene	Personal	HPLC/FD	All	HPLC/FD	Winter + summer
Siwinska, 1999	Poland	B(a)P	Stationary	HPLC/FD	NS	HPLC/FD	Winter
Saint-Amour, 2000	Canada	B(a)P	Stationary	GC/MS	NS	GC/MS	Winter
Bouchard, 2001	Canada	10 PAHs	Stationary	GC/MS	NS	GC/MS	Summer
Staessen, 2001	Belgium	-	_ ′	_	All	n.a.	Summer
Ruchiwarat, 2002	Thailand	Carcinogenic PAHs*	Personal	HPLC/FD	NS	HPLC/FD	Winter

*Carcinogenic PAHs: include benzo(a)anthracene, benzo(b)fluoranthene, benzo(k)fluoranthene, chrysene, benzo(a)pyrene, dibenzo(ah)anthracene, indeno(1,2,3-cd)pyrene, benzo(ghi)perylene; B(a)P: benzo(a)pyrene.

HPLC, high performance liquid chromatography; FD, fluorescence detection; GC/MS, gas chromatography/mass spectrometry; SFS, synchronous fluorescence spectrometry.

NS, non-smokers; All, smokers and non-smokers.

n.a., information not available

B(a)P in air and urinary 1-OHP (table 1). Of the remaining five studies, two had no environmental data of any PAH,^{39 60} one measured the sum of 10 PAHs as a measure of exposure,²⁷ another presented the sum of eight carcinogenic PAHs,⁶¹ and one measured only pyrene in air⁵³. 1-OHP levels were not corrected for creatinine in two studies^{39 53} and these were not further considered. Of the 14 studies with air B(a)P and 1-OHP levels (expressed as µmol/mol creatinine), five presented biomarker levels of 1-OHP as geometric mean, two as the median, and the remaining eight studies presented mean levels. Geometric means were converted to arithmetic means when both the geometric mean and geometric standard deviation were available. We used the following formula⁶³ that is based on maximum likelihood estimation, under the assumption of a log-normal distribution of environmental BaP64 and of 1-OHP concentrations:65 $AM = GM^{*}exp((0.5^{*}(\ln(GSD))2^{*}((N-1)/N))), \text{ where } AM =$ arithmetic mean, GM = geometric mean, GSD = geometric standard deviation, and N = number of subjects. Personal monitoring of B(a)P was done in five studies. In five other studies information on methods of determination of B(a)P was lacking.27 44 46 47 58 Levels of B(a)P exposure in the selected populations were in the range 0.10-64.5 ng/m³ and the levels of 1-OHP ranged from 0.01 to 3.39 µmol/mol creatinine.

Figure 1 shows within study differences in mean, median, or geometric mean 1-OHP levels of subjects exposed and unexposed to environmental PAHs. The figure includes all studies except those by Chuang and colleagues⁵¹ and Gundel and colleagues,⁴⁴ because in these two studies the reference groups were smokers. In 15 of 21 pairs of data exposed subjects had higher levels of 1-OHP than non-exposed subjects, and in 10 of the 21 comparisons the difference were statistically significant. However, none of the studies found a statistically significant correlation between environmental exposure to PAHs or B(a)P and urinary concentration of 1-OHP at the individual level.

When evaluating between study differences, a statistically significant correlation was found between the log transformed mean values of B(a)P and log transformed mean values of 1-OHP (Pearson correlation coefficient, r = 0.76; p < 0.001), on the basis of 17 pairs of data of B(a)P and 1-OHP. The Pearson correlation coefficient was higher when evaluating mean values of B(a)P and median values of 1-OHP (r = 0.83, p = 0.019, 7 pairs of data).

A higher correlation was found when the analysis was restricted to the four studies with personal sampling of B(a)P and presenting mean values for 1-OHP (r = 0.83; p = 0.040), compared to studies doing stationary monitoring (r = 0.70, p = 0.017) (fig 2). In this correlation we considered independently every pair of data of B(a)P and 1-OHP, and some studies contributed therefore with more than one estimate.

DNA adducts

From the 17 articles on DNA adducts included, seven studies measured B(a)P in air,^{9 11 13 16 43 52 56 59} five studies reported measurements of other pollutants, either particulate matter or sums of different PAHs,^{12 41 45 49 61} and one study reported both B(a)P and sums of PAHs.¹³ The remaining four studies^{38 39 42 57} determined adducts but the exposure was defined using proxy variables, usually whether subjects lived in an urban or rural environment (table 2). Sampling of PAHs was done through personal monitoring in six studies, with only one study sampling B(a)P.¹¹ The method used to determine adduct levels was mostly ³²P-postlabelling, applying either nuclease P1 or butanol enrichment procedures. Most studies were performed during winter, with some studies having measurements both in winter and summer. One study⁵² found very high adduct levels compared to other studies in both exposed and unexposed subjects (table 2), and was not considered further.

In the 25 pairs of exposed-unexposed subjects measured in which adducts have been measured by ³²P-postlabelling, 15 had higher DNA adduct levels in exposed compared to unexposed, whereas in 10 of the pairs unexposed subjects had higher levels (fig 3). In the three studies determining adducts using immunoassay, the exposed had higher levels than the unexposed in three of four pairs of data (fig 3). The mean DNA adduct level measured by immunoassay was



Figure 1 Comparison of levels of 1-OHP between exposed (dark bars) and unexposed (light bars) subjects environmentally exposed to PAHs. * Statistically significant differences between exposed and unexposed subjects in each pair of data. w, winter; s, summer.

14.33 adducts/ 10^8 nucleotides, and by ³²P-postlabelling 4.44 adducts/ 10^8 nucleotides. The mean airborne B(a)P levels were also higher in those studies using immunoassay (20.94 ng/m³ B(a)P) compared to levels in studies using ³²P-postlabelling (8.67 ng/m³ B(a)P).

After log transformation of data for B(a)P levels in air and DNA adduct levels, the correlation between studies using stationary measurement of mean B(a)P in air and reporting mean DNA adducts was significant with a Pearson correlation coefficient of 0.60 (p = 0.038) on the basis of 12 pairs of

log transformed BaP and adducts data from 3 studies (fig 4A). This correlation includes data points that are not independent since in some studies the same subjects were evaluated using both ELISA and ³²P-postlabelling. The correlation coefficient was higher for analysis using ELISA (Pearson correlation coefficient r = 0.99, p = 0.019, 3 pairs of data) compared to ³²P-postlabelling (r = 0.46, p = 0.209, 9 pairs of data) (figs 4B and 4C). One additional study⁴³ reported median adduct levels and is not included in the graph or the correlations. Fewer studies reported results on personal B(a)P



Figure 2 Correlation between environmental B(a)P levels and levels of 1-OHP in urine. (A) Pearson coefficient of natural log transformed values (p value). Personal monitoring and mean 1-OHP. r=0.831 (p=0.04). (B) Stationary monitoring and mean 1-OHP. r=0.70 (p=0.017).

Author, year	Country	Type of air PAHs sampled	PAH monitoring	PAH analysis technique	Smoking status	DNA adduct analysis	Season of sampling
Perera, 1992	Poland	B(a)P	Stationary	GC/MS	All	³² P-postlabelling; ELISA	Winter + summer
Grzybowska,1993	Poland	n.a.	- `	_	All	³² P-postlabelling (nuclease P1)	Winter + summer
Hemminki, 1994	Sweden	n.a.	—	—	NS	³² P-postlabelling (nuclease P1)	Winter
Binkova, 1995	Czech Republic	Carcinogenic PAHs	Personal	HPLC	NS	³² P-postlabelling (Nuclease P1 & Butanol extraction)	Winter
Farmer, 1996	Europe	Particulate matter	Stationary	n.a.	NS	³² P-postlabelling with HPLC	n.a.
Binkova, 1996	Czech Republic	Carcinogenic PAHs*; B(a)P	Personal	HPLC	NS	³² P-postlabelling (butanol extraction)	Winter
Nielsen, 1996	Denmark	B(a)P	Stationary	n.a.	NS	³² P-postlabelling (nuclease P1 and butanol extraction)	Winter
Yang, 1996	Italy	n.a.	_	_	n.a.	³² P-postlabelling (nuclease P1)	Winter
Lewtas, 1997	Czech Republic	Carcinogenic PAHs*	Personal	HPLC	All	³² P-postlabelling (butanol extraction)	n.a.
Whyatt, 1998	Poland	PM10	Stationary	n.a.	All	ELISA	Winter
Poirier, 1998	Germany, Kuwait	B(a)P	Stationary	GC	n.a.	³² P-postlabelling (nuclease P1); Delfia	Winter + summer
Peluso, 1998	Italy	B(a)P	Personal	HPLC	NS	³² P-postlabelling (nuclease P1)	Winter + summer
Autrup, 1999	Denmark	B(a)P	Stationary	n.a.	NS	³² P-postlabelling (butanol extraction)	Winter
Marafie, 2000	Kuwait, UK	B(a)P	Stationary	n.a.	NS	³² P-postlabelling (nuclease P1 and butanol extraction)	Winter + summer
Palli, 2000	Italy	n.a.	_	_	All	³² P-postlabelling (nuclease P1)	Winter + summer
Georgiadis, 2001	Greece	Carcinogenic PAHs*	Personal	n.a.	NS	³² P-postlabelling (nuclease P1)	Winter + summer
Ruchiwarat, 2002	Thailand	Carcinogenic PAHs*	Personal	HPLC	NS	³² P-postlabelling (butanol extraction)	Winter

*Carcinogenic PAHs: include benzo(a)anthracene, benzo(b)fluoranthene, benzo(k)fluoranthene, chrysene, benzo(a)pyrene, dibenzo(ah)anthracene, indeno(1,2,3-cd)pyrene, benzo(ghi)perylene.

B(a)P, benzo(a)pyrene; HPLC, high performance liquid chromatography; GC/MS, gas chromatography/mass spectrometry.

NS, non-smokers; All, smokers and non-smokers.

n.a., information not available.



Mean DNA adduct levels

Median DNA adduct levels

Figure 3 Comparison of the levels of DNA adducts, analysed by ³²P-postlabelling and immunoassay in exposed (dark bars) and unexposed (light bars) subjects to environmental PAHs. * Statistically significant differences between exposed and unexposed subjects in each pair of data. w, winter; s, summer; n.a., information not available

monitoring and DNA adducts, using median adduct levels (r = 0.53, p = 0.220, 7 pairs of data).

Four studies¹² ¹³ ⁵⁹ ⁶¹ and a pilot study⁴¹ measured the eight carcinogenic PAHs in air and determined adducts through ³²P-postlabelling. One of these studies⁴¹ reported median adduct levels rather than means, and was not considered further. Mean PAHs levels by personal monitoring were 14.9 ng/m³ (range 1.62–72.9 ng/m³); mean DNA adducts was 2.54 adducts/10⁸ nucleotides (range 0.92–5.59 adducts/10⁸ nucleotides). The Spearman correlation coefficient between carcinogenic PAHs and DNA adducts was 0.617 (p = 0.077).

Protein adducts

Nine studies determined albumin adducts using different techniques,^{13 15 55 39 40 43 45 52 61} one assessed BPDE-haemoglobin adducts,⁵³ and another analysed both biomarkers.⁷ Albumin adduct levels in exposed subjects were higher than in unexposed subjects in five of these studies (fig 5), while in the remaining studies this relation was inverse. The mean albumin adduct levels in exposed was 2.75 fmol B(a)P/µg while in the unexposed subjects it was 3.04 fmol B(a)P/µg. Airborne B(a)P was only measured in two studies,^{15 52} while another study measured particulate matter,⁴⁵ and another measured the sum of eight carcinogenic PAHs.⁶¹

DISCUSSION

Studies on air pollution and health⁶⁶ have focused on short term effects and have applied predominantly measures of exposure to ambient air pollutants. The use of biomarkers has focused on highly exposed populations, such as populations living in heavily exposed environments67 or occupational groups.^{68 69} Occupational studies and studies on active and passive smoking have reported positive correlations between B(a)P and some biomarkers, such as 1-OHP and PAH-DNA adducts,28 but in exposure situations where B(a)P concentrations were 10 to 100-fold higher than those normally found in ambient air concentrations. The use of biomarkers for evaluating low level environmental exposures has been limited and their use has not been generally accepted for the study of such exposures.⁷⁰ In this review we showed that both PAH metabolites (1-OHP) and PAH-DNA adducts can be usefully applied to assess environmental exposure to PAHs at a group level. The correlation between environmental levels and 1-OHP was high in particular when personal monitoring of B(a)P was done, indicating that this biomarker can distinguish between fairly small exposure gradients. For DNA adducts the correlation, although statistically significant, was not as high and results were based on few studies. There was considerable heterogeneity between studies concerning methods used to measure exposure; for example, type of compounds measured and use of stationary or personal sampling, the methods used for biomarker determination, and the availability of information on confounders. These differences have probably resulted in an underestimation of the correlation between atmospheric PAHs and DNA adducts.

All the analyses presented are based on the correlation of mean levels of the environmental exposure and the biomarker levels. Individual levels would only be available if the raw data from each study were recovered and analysed. The overall variance in the populations studied is, therefore, underestimated in our analysis and this should affect the tests for statistical significance. It is, however, unlikely that comparison of mean values of both the exposure and the outcome has affected the main results since the slope of the correlation should not depend on the individual values within each study. What is more likely is that the slope we presented is an underestimate due to errors in the measurement of variables. The application of statistical techniques to adjust for inter-laboratory variation in the adducts analysis, such as the calculation of the "frequency ratio" that models the within study differences in adducts,^{29 62} was not feasible due to the small number of studies and the heterogeneity in design between studies.



Figure 4 Correlation between stationary measured mean B(a)P levels in air and mean DNA adducts. (A) Scatterplot of natural log transformed values of mean DNA adduct levels and stationary monitoring of B(a)P (overall). Pearson coefficient: r=0.60 (p=0.038). (B) Adducts measured by ELISA. Pearson coefficient: r=0.99 (p=0.019). (C) Adducts measured by ³²P-postlabelling. Pearson coefficient: r=0.46 (p=0.209).

PAHs in the air are present at concentrations from tens of ng/m^3 in non-polluted or low-polluted areas to hundreds of ng/m^3 in polluted areas. Most studies have used B(a)P as a reference substance for carcinogenic PAHs because this compound has a carcinogenic potency 10–100 times greater than many other PAHs. Nonetheless, the presence in the environment of other PAHs, some of which carcinogenic, does not allow an accurate estimation of the risk linked to a PAH mixture on the basis of B(a)P concentration alone. The major contributors to air PAHs in the urban and suburban atmosphere are mobile sources from diesel and gasoline engines. Emissions from these sources contain mainly benzo(g,h,i)perylene, pyrene, fluoranthene, and phenanthrene,^{71 72} so that measuring only B(a)P as an index substance may result in exposure underestimation.

High correlation coefficients were found between personal B(a)P measurements in air and 1-OHP levels, while the correlation was lower for PAH-DNA adducts. These results were less clear when comparing levels among exposed and unexposed subjects in a dichotomous way, and in a few studies higher levels of 1-OHP and PAH-DNA adducts were found among unexposed subjects. This is probably due to the smaller differences in level of exposure to PAHs within studies, compared to differences between studies. The correlation between studies indicates that differences as low as 5 ng/m³ of B(a)P measured through personal monitoring can be identified using these biomarkers. In several studies differences between exposed and non-exposed subjects were smaller than 5 ng/m³ and in one study those defined as unexposed had higher levels of B(a)P than those defined as exposed. The less clear pattern identified when comparing within study differences may also be due to the lack of adequate control of confounders in many studies. Smoking status of subjects was not specified in some studies, while dietary intake was not considered by the great majority of the authors, although diet has been shown to be the major contributor to the total intake of PAHs² among non-smokers. Differences in the season of blood drawing may have been another factor affecting results. Air pollution originates in many cities mainly from residential heating and vehicle exhausts, and in general B(a)P levels in winter are higher

than those measured during summer.¹⁶ This should be reflected in the adduct levels and could underestimate the correlation between PAH exposure and the level of biomarker in those cases where the levels of atmospheric pollutants were only an annual mean.¹⁶ ⁴⁴ ⁴⁷ ⁷³ These results point out the extreme importance of evaluating adequately both the biomarker and all the various potential environmental exposures to PAHs, since levels of these biomarkers reflect the exposure from many different sources.¹¹ ⁴³ ⁵⁵ ⁷⁴ Dickey and colleagues⁷⁵ estimated that 2.7 measures of PAH-DNA adducts should be determined to obtain the individual correct level in order to minimise intra-individual variability. Similarly, several measurements of 1-OHP are required for a reliable assessment of environmental exposure to PAHs.³⁵

The analytical techniques used to measure adducts is another factor affecting results. Standardisation of ³²Ppostlabelling has been achieved by means of an interlaboratory trial in 1999.⁷⁶ This technique requires precision and several variations of the assay can be performed, which have been shown to produce very different results. These assay variations can explain some of the differences in DNA adduct levels measured by ³²P-postlabelling. Higher adduct levels resulted from ELISA compared with ³²P-postlabelling, although it is not clear if this was due to higher sensitivity of the method. However, absolute comparisons between the two assays should only be made after calibration in a laboratory.

Protein adducts should, in principle, reflect the formation of DNA adducts.⁵² Results on protein adducts are, however, not consistent, although the number of studies analysed was quite small, compared to that of studies measuring DNA adducts. In addition, the techniques to measure protein adducts are still highly variable between laboratories, which complicates interpretation of results.

Biomarkers to assess exposure to PAHs at high levels are well studied, but more work is needed to validate these biomarkers when exposure occurs at low, environmental levels. At the group level, DNA adducts and particularly 1-hydroxypyrene seem to reflect exposure well, even at low levels of air pollution. By contrast, protein adducts do not appear yet as valid markers to assess environmental exposure





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REFERENCES

- 1 Jongeneelen FJ. Methods for routine biological monitoring of carcinogenic PAH-mixtures. Sci Total Environ 1997;199:141-9.
- 2 Mumtaz MM, George JD, Gold KW, et al. ATSDR evaluation of health effects of chemicals. IV. Polycyclic aromatic hydrocarbons (PAHs): understanding a complex problem. *Toxicol Ind Health* 1996;12:742–971.
- 3 Ovrebo S, Fjeldstad PE, Grzybowska E, et al. Biological monitoring of polycyclic aromatic hydrocarbon exposure in a highly polluted area of Poland Environ Health Perspect 1995;103:838-43.
- Zhao ZH, Quan WY, Tian DH. Urinary 1-hydroxypyrene as an indicator of human exposure to ambient polycyclic aromatic hydrocarbons in a coal-burning environment. *Sci Total Environ* 1990;92:145–54.
 Angerer J, Mannschreck C, Gundel J. Biological monitoring and biochemical and the second secon
- effect monitoring of exposure to polycyclic aromatic hydrocarbons. Int Arch Occup Environ Health 1997;**70**:365–77.
- 6 IARC. Polynuclear aromatic compounds. Part 1. Chemical, environmental and experimental data. IARC Monogr Eval Carcinog Risk Chem Hum 1983;32:1-453.
- 7 Scherer G, Frank S, Riedel K, et al. Biomonitoring of exposure to polycyclic aromatic hydrocarbons of nonoccupationally exposed persons. Cancer Epidemiol Biomarkers Prev 2000;**9**:373–80.
- Vyskocil A, Fiala Z, Fialova D, et al. Environmental exposure to polycyclic aromatic hydrocarbons in Czech Republic. Hum Exp Toxicol 1997:16:589-95.
- Poirier MC, Weston A, Schoket B, *et al.* Biomonitoring of United States Army soldiers serving in Kuwait in 1991. *Cancer Epidemiol Biomarkers Prev* 9 1998;**7**:545-51
- 10 Mumford JL, Lee X, Lewtas J, et al. DNA adducts as biomarkers for assessing exposure to polycyclic aromatic hydrocarbons in tissues from Xuan Wei women with high exposure to coal combustion emissions and high lung cancer
- mortality. Environ Health Perspect 1993;99:83–7.
 Peluso M, Merlo F, Munnia A, et al. ³²P-postlabeling detection of aromatic adducts in the white blood cell DNA of nonsmoking police officers. Cancer Epidemiol Biomarkers Prev 1998;7:3-11.
- 12 Lewtas J, Walsh D, Williams R, et al. Air pollution exposure-DNA adduct dosimetry in humans and rodents: evidence for non-linearity at high doses. *Mutat Res* 1997;378:51–63.
- Binkova B, Levisa J, Miskova I, et al. Biomarker studies in northern Bohemia. Environ Health Perspect 1996;104(suppl 3):591–7.
 Kang DH, Rothman N, Poirier MC, et al. Interindividual differences in the
- concentration of 1-hydroxypyrene-glucuronide in urine and polycyclic aromatic hydrocarbon-DNA adducts in peripheral white blood cells after charbroiled beef consumption. Carcinogenesis 1995;16:1079-85.
- Kure EH, Andreassen A, Ovrebo S, et al. Benzo(a)pyrene-albumin adducts in 15 humans exposed to polycyclic aromatic hydrocarbons in an industrial area of Poland. Occup Environ Med 1997;**54**:662–6.
- Perera FP, Hemminki K, Gryzbowska E, et al. Molecular and genetic damage 16 in humans from environmental pollution in Poland [erratum in Nature 1993;361:564]. Nature 1992;360:256-8.
 17 Motykiewicz G, Michalska J, Pendzich J, et al. A molecular epidemiology
- study in women from Upper Silesia, Poland. Toxicol Lett 1998;96-97:195-202
- Creasia DA, Poggenburg JK Jr. Nettesheim P. Elution of benzo[alpha]pyrene from carbon particles in the respiratory tract of mice. J Toxicol Environ Health 1976;**1**:967–73
- Sun JD, Wolff RK, Kanapilly GM. Deposition, retention, and biological fate of 19 inhaled benzo(a)pyrene adsorbed onto ultrafine particles and as a pure aerosol. *Toxicol Appl Pharmacol* 1982;**65**:231–44.
- 20 Brzeznicki S, Jakubowski M, Czerski B. Elimination of 1-hydroxypyrene after human volunteer exposure to polycyclic aromatic hydrocarbons. Int Arch Occup Environ Health 1997;**70**:257–60.
- Jongeneelen FJ. Bos RP. Excretion of pyrene and hydroxypyrene in urine. 21 Cancer Lett 1990;51:175-9.
- 22 Buckley TJ, Lioy PJ. An examination of the time course from human dietary exposure to polycyclic aromatic hydrocarbons to urinary elimination of 1-
- hydroxypyrene. Br J Ind Med 1992;**49**:113–24. **Van Schooten FJ**, Jongeneelen FJ, Hillebrand MJ, *et al.* Polycyclic aromatic hydrocarbon-DNA adducts in white blood cell DNA and 1-hydroxypyrene in

the urine from aluminum workers: relation with job category and synergistic effect of smoking. Cancer Epidemiol Biomarkers Prev 1995;4:69-7

- Jacob J, Grimmer G, Raab G, et al. The metabolism of pyrene by rat liver 24 microsomes and the influence of various mono-oxygenase inducers Xenobiotica 1982;12:45-53.
- 25 Butler JD, Crossley P. An appraisal of relative airborne sub-urban concentrations of polycyclic aromatic hydrocarbons monitored indoors and outdoors. *Sci Total Environ* 1979;11:53–8.
- 26 Jongeneelen FJ, Anzion RB, Henderson PT. Determination of hydroxylated metabolites of polycecia aromatic hydrocarbons in urine. *J Chromatog*. 1987;**413**:227–32.
- Bouchard M, Pinsonneault L, Tremblay C, et al. Biological monitoring of environmental exposure to polycyclic aromatic hydrocarbons in subjects living in the vicinity of a creosote impregnation plant. Int Arch Occup Environ Health 2001;74:505-13.
- Dor F, Dab W, Empereur-Bissonnet P, et al. Validity of biomarkers in environmental health studies: the case of PAHs and benzene. Crit Rev Toxicol 1999:29:129-68.
- Peluso M, Ceppi M, Munnia A, et al. Analysis of 13 ³²P-DNA postlabeling studies on occupational cohorts exposed to air pollution. Am J Epidemiol 2001;153:546-58.
- 30 Motykiewicz G, Malusecka E, Grzybowska E, et al. Immunohistochemical quantitation of polycyclic aromatic hydrocarbon-DNA adducts in human lymphocytes. *Cancer Res* 1995;**55**:1417–22. **Buckley TJ**, Waldman JM, Dhara R, *et al*. An assessment of a urinary
- 31 biomarker for total human environmental exposure to benzo{a}pyrene. Int Arch Occup Environ Health 1995;**67**:257–66.
- 32 Ptashekas J, Ciuniene E, Barkiene M, et al. Environmental and health in Vilnius and Siauliai residents. J Environ Pathol Toxicol Oncol 1996;15:135-41
- Perico A, Gottardi M, Boddi V, et al. Assessment of exposure to polycyclic aromatic hydrocarbons in police in Florence, Italy, through personal air sampling and biological monitoring of the urinary metabolite 1-hydroxypyrene. Arch Environ Health, 2001;56:506-12.
 Sithisarankul P, Vineis P, Kang D, et al. Association of 1-hydroxypyren-glucuronide in human urine with cigarette smoking and broiled or roasted meat consumption. Biomarkers 1997;2:217-21.
 Sithisarankul F, Kang L, et al. Exploration of 1-hydroxypyren-glucuronide in human urine with cigarette smoking and broiled or roasted meat consumption. Biomarkers 1997;2:217-21.
- Siwinska E, Mielzynska D, Smolik E, *et al.* Evaluation of intra- and interindividual variation of urinary 1- hydroxypyrene, a biomarker of exposure to polycyclic aromatic hydrocarbons. Sci Total Environ 1998:**217**:175–83.
- **Zhao ZH**, Quan WY, Tian DH. The relationship between polynuclear aromatic hydrocarbons in ambient air and 1-hydroxypyrene in human urine. *J Environ* 36 Sci Health 1992;A27:1949-66
- Kanoh T, Fukuda M, Onozuka H, et al. Urinary 1-hydroxypyrene as a marker 37 of exposure to polycyclic aromatic hydrocarbons in environment. Environ Res 1993;62:230-41
- Grzybowska E, Hemminki K, Chorazy M. Seasonal variations in levels of 38
- DNA adducts and X-spots in human populations living in different parts of Poland. Environ Health Perspect 1993;99:77–81.
 Hemminki K, Zhang LF, Kruger J, et al. Exposure of bus and taxi drivers to urban air pollutants as measured by DNA and protein adducts. Toxicol Lett 1994;72:171–4.
- Crawford FG, Mayer J, Santella RM, et al. Biomarkers of environmental tobacco smoke in preschool children and their mothers. J Natl Cancer Inst 1994:86:1398-402.
- Binkova B, Lewtas J, Miskova I, et al. DNA adducts and personal air monitoring of carcinogenic polycyclic aromatic hydrocarbons in an environmentally exposed population. *Carcinogenesis* 1995;**16**:1037–46. **Yang K**, Airoldi L, Pastorelli R, *et al.* Aromatic DNA adducts in lymphocytes of
- 42 humans working at high and low traffic density areas. *Chem Biol Interact* 1996;**101**:127–36.
- 43 Nielsen PS, Okkels H, Sigsgaard T, et al. Exposure to urban and rural air pollution: DNA and protein adducts and effect of glutathione-S-transferase enotype on adduct levels. Int Arch Occup Environ Health 1996;**68**:170-6.
- 4 Gundel J, Mannschreck C, Buttner K, et al. Urinary levels of 1-hydroxypyrene, 1-,2-,3-, and 4-hydroxyphenanthrene in females living in an industrial area of Germany. Arch Environ Contam Toxicol 1996;31:585–90.
- 45 Farmer PB, Sepai O, Lawrence R, et al. Biomonitoring human exposure to environmental carcinogenic chemicals. Mutagenesis 1996;11:363-81
- 46 Cerna M, Pastorkova A, Myers SR, et al. The use of a urine mutagenicity assay in the monitoring of environmental exposure to genotoxins. *Mutat Res* 1997;**391**:99–110.
- Gilbert NL, Viau C. Biological monitoring of environmental exposure to PAHs 17 in the vicinity of a Soderberg aluminium reduction plant. Occup Environ Med 1997:54:619-21.
- 48 Hara K, Hanaoka T, Yamano Y, et al. Urinary 1-hydroxypyrene levels of garbage collectors with low-level exposure to polycyclic aromatic ydrocarbons. Sci Total Environ 1997;199:159-64.
- Whyatt RM, Santella RM, Jedrychowski W, et al. Relationship between ambient air pollution and DNA damage in Polish mothers and newborns. Environ Health Perspect 1998;106(suppl 3):821-6.
- .50 Merlo F, Andreassen A, Weston A, et al. Urinary excretion of 1hydroxypyrene as a marker for exposure to urban air levels of polycyclic aromatic hydrocarbons. *Cancer Epidemiol Biomarkers Prev* 1998;7:147–55.
- Chuang JC, Callahan PJ, Lyu CW, et al. Polycyclic aromatic hydrocarbon exposures of children in low-income families. J Expo Anal Environ Epidemiol 1999;9:85-98.
- Autrup H, Daneshvar B, Dragsted LO, et al. Biomarkers for exposure to 52 ambient air pollution-comparison of carcinogen-DNA adduct levels with

other exposure markers and markers for oxidative stress. Environ Health Perspect 1999;107:233-8.

- 53 Pastorelli R, Guanci M, Restano J, et al. Seasonal effect on airborne pyrene, urinary 1-hydroxypyrene, and benzo(a)pyrene diol epoxide-hemoglobin adducts in the general population. Cancer Epidemiol Biomarkers Prev 1999.8.561-5
- 54 Siwinska E, Mielzynska D, Bubak A, et al. The effect of coal stoves and environmental tobacco smoke on the level of urinary 1-hydroxypyrene. Mutat Res 1999;445:147-53.
- Tang D, Warburton D, Tannenbaum SR, et al. Molecular and genetic damage 55 from environmental tobacco smoke in young children. Cancer Epidemiol Biomarkers Prev 1999;8:427-31.
- Biomarkers Prev 1999;8:427–31.
 Marafie EM, Marafie I, Emery SJ, et al. Biomonitoring the human population exposed to pollution from the oil fires in Kuwait: analysis of placental tissue using ³²P-postabeling. Environ Mol Mutagen 2000;36:274–82.
 Palli D, Vineis P, Russo A, et al. Diet, metabolic polymorphisms and DNA adducts: the EPIC-Italy cross-sectional study. Int J Cancer 2000;87:444–51.
 Saint-Amour M, Tremblay C, Jacques L, et al. [Biological monitoring of
- exposure to polycyclic aromatic hydrocarbons among people living nearby an aluminium smelter in the province of Quebec]. Rev Epidemiol Sante Publique 2000:48:439-48.
- 2000, 40.40, 40. 9 Georgiadis P, Topinka J, Stoikidou M, et al. Biomarkers of genotoxicity of air pollution (the AULIS project): bulky DNA adducts in subjects with moderate to low exposures to airborne polycyclic aromatic hydrocarbons and their relationship to environmental tobacco smoke and other parameters. Carcinogenesis 2001;22:1447-57
- Staessen JA, Nawrot T, Hond ED, et al. Renal function, cytogenetic 60 measurements, and sexual development in adolescents in relation to environmental pollutants: a feasibility study of biomarkers. Lancet 2001:357:1660-9
- Ruchirawa M, Mahidol C, Tangjarukij C, et al. Exposure to genotoxins present in ambient air in Bangkok, Thailand—particle associated polycyclic aromatic hydrocarbons and biomarkers. *Sci Total Environ* 2002;287:121–32. 61
- 62 Mastrangelo G, Fadda E, Marzia V. Polycyclic aromatic hydrocarbons and cancer in man. Environ Health Perspect 1996;104:1166-70.
- 63 Boleij J, Buringh B, Heederick D, Kromhout H. Occupational hygiene of chemical and biological agents. Amsterdam: Elsevier Science B.V., 1995.

- 64 Ott WR. A physical explanation of the lognormality of pollutant concentrations. J Air Waste Manage Assoc 1990;40:1378–83.
- **Roggi C**, Minoia C, Sciarra GF, *et al*. Urinary 1-hydroxypyrene as a marker of exposure to pyrene: an epidemiological survey on a general population group. *Sci Total Environ* 1997;**199**:247–54. 65
- 66 Ballester DF, Tenias JM, Perez-Hoyos S. [The effects of air pollution on health: an introduction]. Rev Esp Salud Publica 1999;73:109-21
- 67 Kim H, Cho SH, Kang JW, et al. Urinary 1-hydroxypyrene and 2-naphthol concentrations in male Koreans. Int Arch Occup Environ Health 2001;74:59-62.
- 68 Clavel J, Mandereau L, Limasset JC, et al. Occupational exposure to polycyclic aromatic hydrocarbons and the risk of bladder cancer: a French case-control study. Int J Epidemiol 1994;23:1145-53.
- Van Rooij JG, Van Lieshout EM, Bodelier-Bade MM, et al. Effect of the reduction of skin contamination on the internal dose of creosote workers exposed to polycyclic aromatic hydrocarbons. Scand J Work Environ Health 1993;19:200-7.
- 70 Jongeneelen FJ, Bos RP, Anzion RB, et al. Biological monitoring of polycyclic aromatic hydrocarbons. Metabolites in urine. Scand J Work Environ Health 1986.12.137-43
- 71 Santadonato J, Howard P, Basu D. Health and ecological assessment of polynuclear aromatic hydrocarbons. J Environ Pathol Toxicol 1981;5:1–364.
- 72 Rogge WF, Hildemann LM, Mazurek MA, et al. Sources of fine organic aerosol. 2. Noncatalyst and catalyst-equipped automobiles and heavy-dust diesel trucks. Environ Sci Technol 1993;27:636-51.
- Whyatt RM, Bell DA, Jedrychowski W, et al. Polycyclic aromatic hydrocarbon-DNA adducts in human placenta and modulation by CYP1A1 73 induction and genotype. Carcinogenesis 1998;19:1389-92
- 74 Pearce N, de Sanjose S, Boffetta P, et al. Limitations of biomarkers of exposure in cancer epidemiology. *Epidemiology* 1995;6:190–4. 75 **Dickey C**, Santella RM, Hattis D, *et al.* Variability in PAH-DNA adduct
- measurements in peripheral mononuclear cells: implications for quantitative cancer risk assessment. Risk Anal 1997;17:649-56
- 76 Phillips DH, Castegnaro M. Standardization and validation of DNA adduct postabelling methods: report of interlaboratory trials and production of recommended protocols. *Mutagenesis* 1999;**14**:301–15.