Supplementary Data

Substrate Recognition, Protein Dynamics, and Novel Iron-Sulfur Cluster in

Pseudomonas aeruginosa Adenosine Phosphosulfate Reductase

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Materials and Methods

Materials

Nonradioactive APS was purchased from Biolog Life Sciences Institute, $\geq 95\%$ (Bremen, Germany). [³⁵S]SO₄²⁻ (specific activity 1491 Ci/mmol) was obtained from MP Biochemicals (Irvine, California, United States). Molecular biology grade DTT was from Invitrogen (Carlsbad, California, United States). *E. coli* Trx protein was purchased from EMD Biosciences (San Diego, California, United States). DPCC-treated trypsin was purchased from Sigma. Depending upon availability, PEI-Cellulose TLC plates (20 cm x 20 cm) were purchased from J.T. Baker (Phillipsburg, New Jersey, United States) or EMD Biosciences. ³⁵S-labeled APS and PAPS were prepared by incubating [³⁵S]Na₂SO₄, ATP, ATP sulfurylase (Sigma), inorganic pyrophosphatase (Sigma) and recombinant APS kinase together as previously described¹. All other chemicals were purchased from J. T. Baker and were of the highest purity available ($\geq 95\%$).

Protein Expression and Purification

The gene encoding the *M. tuberculosis* APS reductase was amplified from H37Rv *M. tuberculosis* genomic DNA and cloned in a protein expression vector as previously described². The gene encoding *Mycobaterium smegmatis* APS reductase was amplified from *M. Smegmatis* genomic DNA and the gene encoding *P. aeruginosa* APS reductase was amplified from *P. aeruginosa* genomic DNA ATCC 47085D (ATCC, Manassas, Virginia, United States) as previously described¹. Briefly, APS reductase genes were amplified via PCR and cloned into the pET24b vector (Novagen) using the 5' Nde I and 3' Xho I restriction enzyme sites. The expression plasmid encoding

Rhizobium meliltoi APS reductase was generated as previously described³.

Proteins were expressed by transforming a reductase-containing plasmid into BL21(DE3) cells (Novagen) grown on LB-agarose containing 50 µg/ml kanamycin. An isolated colony was grown in 5 ml of LB broth containing 50 µg/ml kanamycin. The culture was grown at 37 °C overnight. This culture was used to inoculate 1 L of LB broth containing 50 µg/ml kanamycin. The culture was grown with shaking (250 rpm) at 37 °C to an OD of 0.6, and isopropyl- β -D-thiogalactopyranoside (IPTG) was added to a final concentration of 0.4 mM and the cells harvested after 4 h. Subsequently, 1 L of cells were collected by centrifugation and resuspended in 30 ml of lysis buffer (20 mM sodium phosphate, pH 7.4, 0.5 M sodium chloride (NaCl), 10 mM imidazole, 1mM methionine) together with an EDTA-free protease inhibitor tablet (Roche, Indianapolis, Indiana, United States). After sonication, DNase and RNase (Sigma) were added to the lysate at 10 µg/ml and 5 µg/ml, respectively, and stirred for 10 min on ice. The cell lysate was cleared by centrifugation and the supernatant was applied to a 5-ml HiTrap Chelating column (Amersham, Piscataway, New Jersey, United States). The column was washed with ten column volumes in 20 mM phosphate, pH 7.4, 0.5 M NaCl and 50 mM imidazole, and was eluted with 20 mM phosphate, pH 7.4, 0.5 M NaCl and 250 mM imidazole. Fractions containing the desired protein were pooled and concentrated using Amicon 10,000-Da molecular weight cut-off centrifugal filters (Millipore, Billerica, Massachusetts, United States) before injection onto a 16/60 Superdex 200 prep grade gel filtration column. The standard gel filtration buffer was 50 mM Tris-HCl, pH 8.0, 10% Glycerol, 5 mM DTT with ionic strength adjusted to 150 mM with NaCl. Fractions

containing APS reductase were pooled, aliquoted into single use portions, snap-frozen in liquid nitrogen and stored at -80 °C. Protein concentrations were determined precisely by quantitative amino acid analysis (AAA Service Laboratory, Boring, Oregon, United States).

Mass Spectrometric Analysis of Dissolved Crystals and Enzyme-Substrate Complexes

Solutions were infused at a rate of 2 µl/min into an Apollo electrospray source (Bruker, Billerica, MA), operated in the positive mode. The syringe and spray chamber were wrapped with ice bags to maintain low temperature, in order to prevent the protein from precipitating. All ions were collected using gated trapping and detected using chirp excitation. Broad band data were acquired using an average of 16-64 time domain transients containing 32 K or 1 M data points. The original time domain free induction decay (FID) spectra were zero filled, Gaussian-multiplied and Fourier transformed. All data were acquired and processed using Bruker Xmass version 6.0.0 software. The parameters of the electrospray ionization (ESI) source, ion optics, and cell were tuned for the best signal-to-noise ratio and were maintained for systematic experiments.

Crystals from 10 drops were harvested and centrifuged, yielding a brown pellet, which was washed three times in reservoir solution. The pellet dissolved readily in 10 μ l of 2 mM β -octyl glucoside in H₂O, and the resulting solution was discernibly light greenbrown in color. The solution was incubated overnight with Biobeads to remove detergent and frozen. Subsequently, this sample was dialyzed against 50 mM ammonium acetate using Amicon 10,000-Da molecular weight cut-off centrifugal filters

to remove residual salt and detergent, and the protein concentration was determined (20 μ M). This solution was then diluted with 80:20 acetonitrile:water containing 1% formic acid for mass spectrometry analysis. The derived molecular weights correspond to the full length polypeptide plus SO₃⁻ (31359.8 Da; theoretical 31360.1 Da) and the same minus the three N-terminal amino acids (31018.2 Da; theoretical 31018.6 Da) (50% of the protein used for crystallization lacked these residues) (Supplementary Figure 4(a)). The data indicate that the enzyme in the crystals is quantitatively sulfated.

For enzyme-substrate incubation experiments with *M. tuberculosis* APS reductase (Supplementary Figures 4(b), (c)), appropriate volumes of enzyme (after buffer exchange to ammonium acetate) and APS were mixed in ammonium acetate buffer and the mixtures were chilled on ice for at least 15 min before being introduced into the mass spectrometer.

Supplementary References

- Carroll, K. S., Gao, H., Chen, H., Stout, C. D., Leary, J. A. & Bertozzi, C. R. (2005). A conserved mechanism for sulfonucleotide reduction. *PLoS Biology* 3, e250.
- 2. Williams, S. J., Senaratne, R. H., Mougous, J. D., Riley, L. W. & Bertozzi, C. R.

(2002). 5'-Adenosinephosphosulfate lies at a metabolic branch point in mycobacteria. *J. Biol. Chem.* **277**, 32606-32615.

- 3. Schwedock, J. & Long, S. R. (1990). ATP sulphurylase activity of the nodP and nodQ gene products of *Rhizobium meliloti*. *Nature* **348**, 644-647.
- Thompson, J. D., Gibson, T. J., Plewniak, F., Jeanmougin, F. & Higgins, D. G. (1997). The CLUSTAL_X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Res.* 25, 4876-4882.

Supplementary Figure Legends

Supplementary Figure 1. Routes of sulfate assimilation. Inorganic sulfate is adenylated by ATP sulfurylase (a) to form APS. Higher plants and the majority of sulfate reducing bacteria use APS as their source of sulfite $(c_1 \rightarrow d \rightarrow e)$. In some organisms, APS kinase (b) phosphorylates APS at the 3'-hydroxyl to form PAPS for use as a sulfate donor for sulfotransferases or as a source of sulfite. The lower pathway of sulfate reduction $(c_2 \rightarrow d \rightarrow e)$ is utilized by γ -proteobacteria such as *E. coli* and some fungi. Depending on the organism, APS or PAPS is reduced to sulfite by APS reductase (c₁) and PAPS reductase (c₂), respectively. Sulfite is reduced to sulfide by sulfite reductase (d) and incorporated into cysteine by O-acetylserine-(thiol) lyase (e). Important

metabolites such as methionine and coenzyme A are, in turn, synthesized from cysteine.

Supplementary Figure 2. Structure based sequence alignment of 38 APS reductases from prokaryotes and plants, and 34 PAPS reductases from prokaryotes and eukaryotes. The Clustal X (v1.81) Multiple Sequence Alignment program⁴ was used first to define profiles for each group, then to align all APS reductase sequences (species names in square brackets) and all PAPS reductase sequences separately, and then to align all 72 sequences as one group. The figure is color coded by residue property. The bar graph indicates the degree of conservation per position and is included in Figure 3 under an abbreviated sequence alignment for four species.

Supplementary Figure 3. Electron density for APS in each of four subunits of *P. aeruginosa* APS reductase. The unbiased σ_A -weighted |Fo|-|Fc| map, based on the final model at 2.70 Å resolution, is contoured at 2.5 σ and 5.0 σ .

Supplementary Figure 4. (a) ESI FT-ICR mass spectrum of dissolved crystals of *P. aeruginosa* APS reductase demonstrating quantitative sulfonation of enzyme in the crystals. The two molecular weights correspond to the full-length enzyme plus SO_3^- , and the same minus the three N-terminal amino acids (~50% of the protein used for crystallization lacked these N-terminal residues). These molecular weights represent the apo-enzymes with the noncovalently bound [4Fe-4S] cluster dissociated from the protein during preparation of the sample for mass analysis (see Methods). (b) ESI FT-

ICR mass spectrum of 10 μ M *M. tuberculosis* APS reductase with 20 μ M APS in ammonium acetate showing that in the presence of excess substrate that the product AMP can be displaced, and that the sulfonated enzyme (E-SO₃⁻) can also bind APS, as observed in the crystals. Previously, 15 μ M enzyme was incubated with 10 μ M APS; under those conditions, only the sulfonated enzyme with AMP bound was observed (Figure 7(a) of ref. 7). (c) ESI mass spectrum of the same mixture dissolved in 80:20 acetonitrile:water containing 1% formic acid, illustrating the release of the noncovalently bound nucleotide and the iron-sulfur cluster. The inset shows the deconvoluted mass of the thiosulfonate intermediate in the apo-form.

Supplementary Figure 5. Partial trypsin proteolysis of *M. tuberculosis* (a) and *P. aeruginosa* (b) APS reductase showing protection of the C-terminal tail and the Arg-loop upon formation of the thiosulfonate intermediate at equimolar concentration. The time course of the trypsin digestion is shown in the presence (+APS) and absence (-APS) for each enzyme. In (a) *M. tuberculosis* APS reductase (50 μ M active site concentration) was incubated with or without 50 μ M APS for 10 min at RT, and trypsin was added at a final concentration of 10 μ g/ml and incubated at 4 °C. In (b) *P. aeruginosa* APS reductase (40 μ M active site concentration) was incubated at 4 °C. All samples were analyzed by SDS-PAGE using a 4-12% gradient Criterion gel. Trypsin digest fragments were purified by reverse phase HPLC and analyzed by electrospray mass spectrometry. In the presence of APS, the starred fragments, HR/G – End* for *M. tuberculosis* and ER/N – SK/A* for *P. aeruginosa*,

represent the mass of the peptide fragment plus an additional 80 Da for the covalent sulfite adduct. Full length *M. tuberculosis* APS reductase without N-terminal Met, is 28,356.87 Da; full length *P. aeruginosa* APS reductase is 31,279.6 Da.

Supplementary Figure 6. Electron density for the [4Fe-4S] cluster and its Cys ligands in Subunit B of *P. aeruginosa* APS reductase. The σ_A -weighted 2|Fo|-|Fc| map, based on the final model at 2.70 Å resolution, is contoured at 1.0 σ and 5.0 σ . The C α -C β -S γ -Fe torsion angle for Cys140 is indicated; this angle is *cis* (+10°) so that the C α and Fe atoms are eclipsed and only 3.5 Å apart.

Supplementary Table 1 Effect of reductants on APS reductase activity ^a			
Reductant ^b	E°′, mV	Activity (pmol/min) ^c	
Thioredoxin	-260	40	
GSH	-230	≤ 0.1	
Reduced lipoic acid	-290	≤ 0.1	
DTT	-330	≤ 0.1	
Dithionite	-527	≤ 0.1	

(a) Rate of APS reduction measured with various reductants. Each value reflects the average of at least two independent experiments, and the standard deviation was less than 15% of the value of the mean.

(b) 10 μM thioredoxin or 10 mM chemical reductant was used in each assay (see Methods).

(c) Due to the slow nature of the reactions measured with chemical reductants, reported rates are considered upper limits.



Supplementary Fig. 1

Page 1 of APSr and PAPSr sequences

is]	MSGETTRLTEPOLRELAARGAAELDGATATDMLRWTDETFGDIGGAGGGVSGHRG	WT 57
sa]	MLPFATIPATERNSAAQHQDPSPMSQPFDLPALASSLADKS-PQDILKAAPEHFGI	DE 56
ae]	FEHFG	DE 33
ns	MSPTPDVVELATTYANKS-AQDILKLAFAEFG	DD 33
daj	MSOPFDVAALAATVANKS -PODILKLA	
111		08 31
nel		ON 33
111	VVRAGLEGORSSENVFORSCSVPOSDKHRGCLNAAIATPEVVOEDINVEDLALGLVDAS PLETMDRA LAMPOO	SD 111
nsl	LRKRWAVNYMEPGKRNDSVAAASAVADLAAVVEEKARALEKEVEVADYEKTAVELONASPLEIMDRALAEFG	NE 114
ys]	AARGQRARAVRSLRAAEPAROPVAASAAAAAPAAPVADEAAALAAVDYEALARELEGASPLEIMDRALAMFG	SE 112
um]	LSSTVLNSRRRSAVKPLYAEPKRNDSIVPSAATIVAPEVGESVEAEDFEKLAKELQNASPLEVMDKALEKFG	DD 115
us]	ISPSVNVSRRRLAVKPINAEPKRNESIVPSAATTVAPEVEEKVDVEDYEKLADELQNASPLEIMDKSLAKFG	ND 114
ea]	VSSVSLSLSGKRSSVKPLNVQSIAKESFGSMVASEVTEKQDVVEVEDFEELAKSLETASPLEIMDKALETFG	ND 116
nal	RTNVSAASLSLSGKRSSVKALNVQSITKESIVASEVTEKLDVEVEDPEELAKRLENASPLEIMDKALEKFG	ND 111
orj	PMKPLCAVDPGKKSQSPVAPLAAVPSSTSDAVTLVQAAETSIVDVVDPKLASELEKASPLEIMDKA	
21	MTAATTNSARN - FRIFTAGRSCRIDELSENSWGARIRIAADARDARD VAT - FRE	82
sel	IIWNRRLLGEEPDSFMTEWGIOKGRRRNEMDRSDFNLDLDEINRAFAGED-PRKLLAHVVEK	PA 78
um]	MRR	YP 35
um]	MVQVENVEN	IE 32
is]	MMAIARPLAAIAGLTAAAAARWHDLOSRLAIIARQY	P- 36
ei]	AA	P- 30
hal	MSGLSDIAVVDGGAPASAL-RPTLWTMPEYTGSLADLDEKERELSARLAGIAARF	2- 55
	MSALQEAAV	- 54
til		20 49
esl	DATKESLERLC - EVKGLI	20
is]	MLTYDNWEEPTITFPEDDPYKG-ALSVLKWAYGH	DQ 37
um]	MSPAPGA	39
ns]	VTVNALKNAGPTQDPEVSPEGPRTTAPLPEEVAHRNRELVDKHADELYHADAATILEWAAEHVPGR	65
14]	MFKRDTFGTK-	41
us]	MTTTDPRDVIRWALAGYP	D- 25
18]	MTALEDRSPASRRTPPLLTFTPETDPLDVVRWALKA	0- 40
ns]		D- 50
isl	VSYISDGREMTSLPECKSVAASAMTOPAAATGTPALPPAAALDALRDAR - PADVIAAALRT	RD 74
ii]	MNK	23
ium	MSQLDLNALNELPKVDRVMALAETNAQLEKLS-AEERVAWALENLP	GE 47
oli	MSKLDLNALNELPKVDRILALAETNAELEKLD-AEGRVAWALDNLP	GE 47
ora	MAEFNLEALNALPKDEQVAALAAVNGQLEQLS-AQERVSWALENLP	GD 47
515	MSQ	SE 47
115	MSA	Q 47
cus		GE 52
cus	MLDSVA	GA 52
eri	MP	GN 47
rae	MPNRTVPTLEELLTLNKVQQTLRLTEVNQHLESLT-AQERVVWGLENLQ	GN 50
dum	MPNYALADLLEKTKIEQILQLAEINAELEALS-AQERVRWALANLG	SE 47
nia	MNKLLNVINHCWTVTEFNSLDINEQKLVLVKINRYLESLN-TVDRFKWAIEY	KQ 55
lda	MRR	SQ 48
720		0 43
ris	MTA	00 43
osa	MTVLPALPPLDDLETLNVHLETLS-AENRVCWALER	DH 39
ina	MSK	PQ 43
tis	MSEFKNILQLDKQSQTSMLADANGILKNMS-AEQRVQWALENLP	ET 45
sis	MSSSELKALLTAPKSVQQAELERINRFLAGLT-AQERVLWGLAYLP	GN 47
ina	MPNEVVTTAPTGAAVDHSVTPAQLQALLLAPKAEQTAALENINLFLAGLT-AQQRVAWGLAYLPA	GD 65
ans		
315		CO 52
nii	MPOSNIHLINSHVDNHOVSEHYNKOTHOAYKPEISIKEVNOSLENAN-ACOVYEWTRETFG	EG 63
nus	MTDGSIDTVSTATSTAAGQSPSVQVASTLEMPLEDARIHLANLE-AQKRLEWALKO	QG 55
eum	MLQQSLIRLNKNGQYNYQDYRYLPELDLGVVNQELSQAD-PHTLVQWGLKIFK	EG 54
tus	MPA	SG 34
lis	MTA	r G 42
mbe	MSSIDT	5- 47
lae		43
asa	SOTSTEASSEKI.DCKFDDNASVDSGFASANSSTANLDI.TSLCKFDDNASVDSGFASANS	N- 77
tus	TSDAEAAVDSRDLOTESGYVSGDSSDVYTPEIVFTKPHLOFLNROLOFLE-POEILRWG	H- 73

[Mycobacterium tuberculos [Pseudomonas aerugino [Pseudomonas syring [Pseudomonas fluoresce [Pseudomonas puti-[Azotobacter vineland [Methylococcus_capsulat [Microbulbifer_degrada [Ceratopteris_richard [Polygonatum_pubesce [Zea ma [Lycopersicon esculent [Catharanthus_rose [Brassica_junc [Arabidopsis thalia [Lemna min [Chlamydomonas_reinhardt [Nocardioides sp. JS614 [Desulfitobacterium hafnien [Pyrobaculum aerophil] [Clostridium thermocell: [Bordetella parapertuss [Burkholderia mall [Ralstonia eutrop [Ralstonia_solanacear [Dechloromonas aromati [Sinorhizobium melilo [Wolinella succinogen [Bacillus subtil [Corynebacterium glutamic [Corynebacterium efficie [Nocardioides sp. JS6 [Chloroflexus aurantiac [Deinococcus_geothermal [Deinococcus radiodura [Nitrobacter_winogradsky [Nitrobacter hamburgens [Cytophaga hutchinson Salmonella_typhimur: Escherichia c Erwinia carotov Yersinia_pseudotuberculo Zymomonas mobi Photorhabdus luminesc Vibrio_parahaemolyti Vibrio vulnifi Vibrio fisch Vibrio_chole Photobacterium profun Candidatus Blochman Candidatus Blochmannia flor Buchnera_aphidic Xanthomonas ory Xanthomonas campest Xylella fastidi Thiocapsa roseopersic Pseudoalteromonas_haloplank Shewanella oneiden Shewanella frigidimar Shewanella denitrific Shewanella amazonen Colwellia psychrerythra Crocosphaera watso Prochlorococcus mari Trichodesmium_erythra Synechococcus elonga Anabaena variabi Schizosaccharomyces po Saccharomyces cerevis Thermosynechococcus elonga Neurospora_cra Aspergillus fumiga

CLUSTAL X (1.81) MULTIPLE SEQUENCE ALIGNMENT

Page 2 of APSr and PAPSr sequences

[Mycobacterium_tuberculosis]	TCNYVVASNMADAVLVDLAAKVRPG	VPVIFLDTGYHFVETIGTRDAIESVYDVRVLNVTPEHTVAEQDELL	128
[Pseudomonas_aeruginosa]	-LWISFS-GAEDVVLVDMAWKLNRN	VKVFSLDTGRLHPETYRFIDQVREHYGIAIDVLSPDPRLLEPLVKEKG	127
[Pseudomonas_syringae]	-LWISFS-GAEDVVLVDMAWKLNKN	VKVFSLDTGRLHPETYRFIEQVRDHYKIDIEIISPDQRALEPFVKEKG	104
[Pseudomonas fluorescens]	-LWISFS-GAEDVVLVDMAWKLNKN	VKVFSLDTGRLHPETYRFIDQVREHYKIDIELVSPDYTKLEPFVKEKG	104
[Pseudomonas putida]	-LWISFS-GAEDVVLVDMAWKLNKQ	VKVFSLDTGRLHPETYRFIDQVREQYNLPIEILSPDRAKLDPFVKEKG	104
[Azotobacter vinelandii]	-LWISFS-GAEDVVLLDMAWKLNKN	VKVFSLDTGRLHPETYRFIEQVREHYGLAIEVLSPDARLLEPLVREKG	104
[Methylococcus capsulatus]	- IAVSFS-GAEDVVLVEMAARLRPG	IEVFTLDTGRLHPETYRFIEEVRERFAVRLDVLSPDAAELEAMVREKG	102
[Microbulbifer degradane]	TATER CAPDUVI VDMASKTKPC		103
[Arcroburbirer_degradans]	TATADA CARDUNITEUN VIMO	DEDUCCI DECRIVERED DE DEVERATO TRUDVERED SARVORIVARAO	103
[Ceracopteris_richardii]	-INIAFS-GARDVALIEIA-KLIGK	PERVESEDIGRENPETERFEDEVERHIGIRIEIMEPDASEVEAUVRSKG	104
[Polygonatum_pubescens]	-IAIAFS-GAEDVALIEYA-KLTGR	PFRVFSLDTGRLNPETYRIFDEVERHINIHIEIMFPDAVEVQALVRTKG	185
[Zea_mays]	-IAIAFS-GAEDVALIEYA-KLTGR	PFRVFSLDTGRLNPETYQLFDKVEKHYGIRIEYMFPDASEVQELVRTKG	183
[Lycopersicon_esculentum]	-IAIAFS-GAEDVALIEYA-HLTGR	PYRVFSLDTGRLNPETYQLFDTVEKHYGIRIEYMFPDSVEVQALVRTKG	186
[Catharanthus roseus]	- IAIAFS - GAEDVALIEYA - HLTGR	PFRVFSLDTGRLNPETYKFFDTVEKQYGIHIEYMFPDAVEVQALVRSKG	185
[Brassica juncea]	- IAIAFS - GAEDVALIEYA - HLTGR	PFRVFSLDTGRLNPETYRLFDTVEKHYGIRIEYMFPDAVEVQALVRNKG	187
[Arabidopsis thaliana]	- IATAFS-GABDVALIEVA-HLTGR	PYRVFSLDTGRLNPETYRLFDTVEKHYGIRIEYMFPDAVEVOALVRNKG	182
[Lemna minor]	- TATAFS-GARDVALTEVA-RLTGR	PERVESI, DTGRI, NPETVREEDEVEKRYNTRTEVMEPDAVEVOAL VRSKG	187
[Chlamydomonas reinhardtii]	VATAFS CARDUALTEVA HITCP		141
[Negardioidag ap 76614 2]	PC WERE WADAWI AUT UCDVADE		140
[Nocardioides_sp05014_2]	FC-VIDSMADAVLANDVSKVAPG		143
esuilicobacterium narniensej	RIADASSISIBDQVLTQMLLKIDAR	ARIFFIDIGRNFQQTIDLMEETMSRIEFHIEVIAPENSELEPFLAEIG	191
[Pyrobaculum_aerophilum]	NIALAFSGQAEDVVVLDIMHKVAPD	KIRVFMLDTGRLPEEIVELVDKVREHVGVEIEIVYPDTKEIEEFVKRVG	109
[Clostridium_thermocellum]	KVALASSLSIEDQVLTDILLKINP	KVRVFFLDTGRHFQQTYDLMEETMHRYGFHYEVYAPESKELEQAVSKYG	105
[Bordetella_parapertussis]	DAALASSLAABDMVLTHAIYAGG	LDLEVFTLDTGRLHTETLGVLDAVATRYGRAITVYRPDAAAVQAHVDAHG	109
[Burkholderia mallei]	KVKFASSLAAEDMLLAHAILSKG	VAIGIFSLNTGRLHAETLDMIERVRERYGYEIEQFHPRQDAVDAYVAEHG	103
[Ralstonia eutropha]	RARFATSLAAEDMVLTDAILRGTPAV	-RAGIRVFTLNTGRLHAETLAVLDEVKVHYGYTVEOFTPDTEAVENYLKKHG	132
[Ralstonia solanacearum]	TAKFASSLAABDMVVTDAILRSPEAV	-RAHLPIFTLOTGRLHAETLAMLERIREHYGYAIEOFAPDARAVOAYVRDHG	131
[Dechloromonag aromatica]	SAAFANSI CARDMULTDI. TUKAK	- I.PTETESL DTGRI.PLETYDT, TAAVDKHYGI, KI.KI.VEPOAREVESVURNHG	108
[Ginerhizehium melileti]	PAUPTTOL OT POULTA A TOCMP	TOTEUNTI ATCOL ENERGY TO TE TROVERY DEDAVINOVO	121
[Sinornizobium merricorr]	A TRADO POURT DI LI VUYDO	OPPUPUT PROVI PROVI SPUPUTOTOTOTOTOTOTOTOTOTOTOTOTOTOTOTO	121
[wormerra_succinogenes]	DAF BRUARDVVVLDLLLLKRIPR	SFEVELLBIGKUFIVILAFRERVERFENDIIVAFRESPAALUSUERELG	91
[Bacillus_subtilis]	-LVYACSFGIEGIVLIDLI-YKVKK	D-AEIVFLDTGLHFKETYETIERVKERYPGLNIILKKPDLTLEEQAEEHG	109
[Corynebacterium_glutamicum]	IA-VILSMENTVLAELAARHLPE	ADFLFLDTGYHFKETLEVARQVDERYS-QKLVTALPILKRTEQDSIYG	108
[Corynebacterium efficiens]	VA-VTLSMENTVLAELAARHLP	QADFLFLDTGYHFEETLEVARKVDERYSQTLVTALPILTRAEQDSL	136
[Nocardioides sp. JS614]	LC-ATSSMSDAAITHLASSCAPG	LDVLFLDTGYHFPETLAFRDQVAAAFD-VRVVDLRSKPAPGNQ	105
[Chloroflexus aurantiacus]	-LLMTSAFNLNGVVLIELA-AQAG	-FA-GEVVFVDTGYHFPETLATRDHLITRYPOMOFVTLSADLPPEPWGE	94
[Deinococcus geothermalis]	-LLMPSAFNLNGVVLLDLA-ARAG	- YR - GEVVEVDTGFHFPETLATEDRLAARYPEMTFUTLHAGAHPEDGOTFP-	111
[Deinococcus radiodurans]	-TLIMPSAENLNGVVLLDLA-AOAG	- VR - GEVVEVDTGVHEPETLATEDRIESEVPELTEVTLNAGASPDDGOTPP-	127
[Nitrobacter_winogradekyi]	PLATWEEFCTESAALLKUM-ADVDP	TOUTELDTOWLFFETLTYPDTLTANLOLKDURSTOPSEDALKOEDEDR	129
[Mitrobacter_winogradsky1]	DT AT WOOPOTPOALATT FUM ADVDT	A TOUTPI DECUI PEPET AND THINK OF TO AND THE TOP ON THE	145
(Mitrobacter hamburgensis)	KUALVBBEGTESKALUKVII-ADVDE		145
[Cycopnaga_nutchinsonii]	SSPQTHSIVLLHIL-SEPDP	S-IPVIFINIGIHFPEIVLIRDQIADLFNLKNIKVVSSATPRNMQKDABG	91
Salmonella_typhimurium	- YVLSSSFGIQAAVSLHLV-NQIR	PDIPVILTDTGYLFPETYQFIDELTDKLKLNLKVYRAGESPAWQEAR	116
Escherichia_coli	-WVLSSSFGIQAAVSLHLV-NQIR	PDIPVILTDTGYLFPETYRFIDELTDKLKLNLKVYRATESAAWQEAR	116
Erwinia carotovora	-FVLSSSFGIQAAISLHLV-TQQR	PDIPVILTDTGYLFPETYQFIDALTEQLKLNLHVYRAAESPAWQESR	116
Yersinia pseudotuberculosis	-FVLSSSFGIQAAVCLHLV-TRQR	PDIPVILTDTGYLFPETYRFIDDLTEKLQLNLQVFRAAHSPAWQEAR	116
Zymomonas mobilis	-FVLSSSFGIOAAVMLHLV-TRKK	ADIPVILTDTGYLFPETYRFIDALTEKLDLNLOVFRAALSPAWOEAR	116
Photorhabdus luminescens	-FVLSSSFGIOAAVCLHLV-TOEY	PDT PVILTDTGYLEPETYOFIDKLTTOLKLNLOVESAEHSPAWOBAR	116
Vibrio parahaemolyticus	- FAVSSSFGIGAAVMLHLV-TOEK	PDTPTTLTDTGYLFARTVRFTDELTEKLNLNLKVYRAEOSAOWORAR	121
Vibrio milnificus	HAVE CERTON AVALUT V SKOO	ADT DUTL TOTONI POPTYOFT DEL TYCININ	121
Vibrie fichani	DAL GOODOTOA AUNI OLU TRUK		116
Vibrio_fischeri	-PADSSFGIQAAVALQUV-TEVK	SDIPTILIDIGILFPETICFIDGLIDKLNENLHVFIADESPNWQEAK	110
Vibrio_cholerae	-HALSSSFGIQAAVMLHLL-TSVK	SDIPVVLTDTGTLFPETYQFIDELTERLNLNLKVYSAPVSAAWQEAR	119
Photobacterium profundum	-FALASSFGIQSAVMLHLV-TNES	PKVPVILTDTGYLPPETYQFIDQLTLRLSLNLYVYRAEISSAWQEAR	116
Candidatus_Blochmannia	-AILSSSFGIQSSVSLHLT-THYY	PNIPIILIDTGYLFPETYRFIDRLTEKMQLNLHIFSPNQSAAWQEAR	124
ndidatus Blochmannia florida	-LVLSSSFGIHSYVSLHLM-TNYY	PSVPVILIDTGYLFPETYQFIDRLTEKMKLNLHVFRPDKSPAWQEAR	117
Buchnera aphidicola	-QIMSSSFGIQSTVLLHLI-IKKK	PDIPVILIDTGYLFPETYNFIDFLTNKFHLNLKVFRSTISSAWQEAR	116
Xanthomonas oryzae	-AALSSSFGAOSAVTLHLL-SOOR	PDIPVILIDTGYLFPETYRFADALTERLKLNLKVYRPLVSRAWMEAR	112
Xanthomonas campestris	-AALSSSFGAOSAVTLHLL-TOOR	PDIPVILIDTGYLFPETYRFADALTERLSLNLOVYRPLVSRAWMEAR	112
Xvlella fastidiosa	-PALSSECAOSAVMLHLL-TREA	PDT PVTLUDTGVLEPETVEFADTLTERIKIN LKUVOPLESGAWTEAR	108
Thiogenes researching	HUL GOODOCAUMT HLU CDOM	PETPUTLUDTONI PRETVOLUDAL TOPECIN	112
audealteremenas balenlanktis	UPL COOPOTOL LINE ULT TTOO	DETENTION PROPERTY AND A CAN AND A C	114
eudoalceromonas haloplankeis	-HELSSSFGIQAAVALALI-TLQQ	PDIPVVLIDIGILEPUTIOFILOMIARLSLNLKVIKSLLSPAWQBAK	114
Shewanella oneidensis	-HALSSSFGIQAAVMLHMV-SQVQ	SDIPVILIDIGILFPETYQFIDQLIERLSLNLKVYQAPITSAWQHAR	110
Shewanella_trigidimarina	-HALSSSFGIQGAVMLHMV-TQAQ	SDIPVILTDTGYLFAETYQFIDQLTERLNLNLKVYQAGQTQAWQHAR	134
Shewanella_denitrificans	-HGLSSSFGIQAAVMLHLV-TQVQ	SDIPVILTDTGYLFPETYGFIDELTERLNLNLKVYQAGQTSAWQEAR	130
Shewanella amazonensis	-HILSSSFGIQAAVMLHLV-THVK	SDVPVVLTDTGYLFPETYRFIDELTERLKLNLKVYRAPMTAAWQEAR	113
Colwellia psychrerythraea	-FVLSSSFGIQSAVMLHLL-TQVD	SNIPVLITDTGHLFPETYRFIEQLTDRLNLNLQVYQAKESAAWQQAK	121
Crocosphaera watsonii	-LVMSTSFGIOAAVMLHLV-TEVV	PDIPVIWVDTGYLPAETYRFAEELTERLKLNLKVYOSPFTPARMEAL	132
Prochlorococcus marinus	- FVLTTSFGTOSAVLLHML - HGLDGG	SVVPVVVDTGVLPARTYFVADOLTROLGLDLKVVOSTMSPARMEAT	130
Trichodesmium erythraeum	-LVMSTSFGIOAAVMLHLV-TSVT	PNIPVIWIDTGYLPARTYK PARELIERLKLNVKVVOSHTSDOPMENT	123
Superborogging alongature	TWI STERATOSAWALHI A TOVO	DTDUTWTDTOVI DTETVDEA BELTEDI KIN IKUVOCETCDADMEAL	103
Synechococcus_eroligatus	THOUGH PAULODI WE WITH HORE	OPUPUT PT DET PERCONT PT UN Y A TANKA	103
Anabaena_variabilis	- DY TRAFRY DDLY ITHILY SELK	VEVEVEFUELDEDVAKAKEIINDDUTYKTPDVDTR-EAF	111
schizosaccharomyces_pombe	-BFQTBALGLSGLVIMDML-SKMD	MNVPLIFINTLHHFPETLDLLEKVKTKVPNVPVHVYRCAEAANEKEF	116
Saccharomyces_cerevisiae	-LFQTTAFGLTGLVTIDML-SKLSE	KYYMPELLFIDTLHHFPQTLTLKNEIEKKYYQPKNQTIHVYKPDGCESEADF	118
hermosynechococcus_elongatus	-LVQTSAFNVDDMVITDLLYRDLRP	-NPPVPVLFLDTLHHFAETLAFVQQAKEKYSLDLRTYKIP-DVDSREAF	116
Neurospora crassa	-LFQTTAFGLTGLVTLDML-SKLAKESP	DAAPVDLIFLDTLYHFKETYALVDRVQERYPNIRLHVFKPADVSNVDEF	152
Aspergillus fumigatus	-LFQTTAFGLTGLVTLDML-SKLQVP	RPOMVDLIFLDTLHHFPETLALVDRVRKRYPLNNIHIYKPAGIETAEEF	146
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[Pseudom [Pseud [Pseudomo [Pse [Azotoba [Methyloco [Microbul [Ceratop [Polygo [Lycopers [Cath [Arabi [Chlamydomo [Nocardioi [Desulfitobact [Pyrobac [Clostridi [Bordetell [Burk [Ral [Ralston [Dechloro [Sinorhi [Wolinel [Ba [Corynebacte [Corynebact [Nocardi [Chlorofle [Deinococc [Deinococ [Nitrobact Nitrobact [Cytopha Salmon Br Yersinia_ps z Photorha Vibrio y Photobac Candid Candidatus Blo Buch Xa Xantho Xy Thiocaps Pseudoalteromo Shewa Shewanel Shewanel Shewan Colwellia Croco Prochlo Trichode Synecho Ana

Schizosace Saccharo Thermosynecho N Asper

CLUSTAL X (1.81) MULTIPLE SEQUENCE ALIGNMENT

Page 3 of APSr and PAPSr sequences .

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[Mycobacterium tuberculosis]	GENT PAR	193
[Bacudomonag peruginoga]	TREPT A HEROTATE TED VILLA VEND CORDAN FOR COUNT BID A BOTTON OF THE COUNT OF THE C	100
[Pseudomonas_aeruginosa]		170
[Pseudomonas_syringae]	LFSFIRDGHGEUCGIRKIEPLRKKLSSVKAWAIGQRKDOSPGIRSAVAALEVDSAFSTAESTLIKFNPLAQ	1/5
[Pseudomonas_fluorescens]	LFSFYKDGHGECCGIRKIEPLRKLSGVKAWATGORRDOSPGTRSAVAVMEIDTAFSTPERTLYKFMPLAQ	175
[Pseudomonas_putida]	LFSFYKDGHGECCGIRKIEPLRKLATVSAWATGQRRDQSPGTRSQVAALEIDSAFSTPERTLYKFNPLAQ	175
[Azotobacter_vinelandii]	LFSFYKDGHGECCGIRKIEPLKRKLATVSAWATGQRRDQSPGTRSQVAEFEVDGAFSTPDRPLYKFNPLAQ	175
[Methylococcus_capsulatus]	LFSFYRDGHQECCGIRKVRPLRRKLATLDAWITGQRRDQNP-TRQAVPEVEADPAFATGHHPLVKFNPLAA	172
[Microbulbifer degradans]	LFSFFEDNHQECCGIRKVGPLRKKLLEVDAWVTGQRKDQSPGTRASIPVIONDRVFARPGDTLTKFNPLVN	174
[Ceratopteris richardii]	MFSFYEDGHQECCRIRKVRPLRRALKGLKAWITGORRDOSPGTRSNIPIVOIDPSFEGMDGGPGSLIKWNPVAN	256
[Polygonatum pubescens]	LFSFYRDGHOECCRVRKVRPLRRALKGLRAWITGORKDOSPGTRAHIPVVDVDPSFEGLDGGAGSLIKWNPVAN	259
[Zea mays]	LESEVED GHOECCEVERVEPLERALEG LEAWITGOREDOSPOTRAST PLUOVDPSFEGLOGGAGSLVEWPVAN	257
[Twoopersicon esculentum]	LESEVED	260
[Catharanthus resourd]	LECEVED AUGCOUDEVED LEDALYC LEDALTCOPPOSE CET DUI OUD VER AND OUCCUVEN DUAN	250
[Cacharanchus_roseus]		261
[Brassica_juncea]	LFSFIEDGHQECCKIRKVRPLRRALKGLRAWIIGQRKDQSPGIRSEIPVVQVDPVFEGLDGGAGSLVKWNPVAR	201
[Arabidopsis_thaliana]	LFSFYEDGHQECCRIRKVRPLRALKGLRAWITGQRKDQSPGTRSEIPVVQVDPVFEGLDGGVGSLVKWNPVAN	256
[Lemna_minor]	LFSFYEDGHQECCRVRKVRPLRRALRGLRAWITGQRKDQSPGTRASVPTVQVDPSFEGFEGGTGSLIKWNPVAN	261
[Chlamydomonas_reinhardtii]	LYSFYEDGHTECCRIRKVKPLRKQLKVYKAWITGQRKDQSPGTRTEVPVVQVDPVFEGVTGGPGSLIKYNPLSN	215
[Nocardioides sp. JS614 2]	YGTDLYXTDPDLCCALREVOPLADALEGYDAWATGLRRAETQ-HEVIAPVIGWDAKRGKVKVSPLAR	215
[Desulfitobacterium hafniense]	PNAFYDKVELRQKCCELRKVRSLQRVLSTVDGWICGLRKEQSP-TRQEIGLLEWDSRHAIYKVNPLAA	218
[Pyrobaculum aerophilum]	INPFYRDVELRHLCCKIRKVNPLLRALSGLDAWITGLRRDOFP-TRATTRKIGIDHDHYGILKISPICD	177
[Clostridium thermocellum]	PNFFYESVELEKKCCETEKVNPLKEVLSTVDAWTCGLEBEOSL-TROELNTPEWDGLESTVKINPTVF	172
[Bordetella parapartuggig]	MHARYES - VALPEACCETDEVEDLOBALAG - POANT COPPAGE - TOGELDDEPDD DVEG LVEPNDLAA	176
[Burkholderia mallei]	INA PYPE UF I PYPE PYPE INDATAG UDANUT CODDOUV TOAPI POPUT ADD	170
[Beletende autrenhe]		100
[Raistonia_eutropha]	LNAFTDSIDLRKDCCGIRKVEPLNKALSHADAWMIGQRKBOAV-IRSELFFEEDDEARAIPKFNPLAD	199
[Raistonia_solanacearum]	LNAFYDSIELRKACCTIRKVVPLNRALRDADAWLSGQRREQAV-TRADLPFVEEDDTRGIAKYNPLFD	198
[Dechloromonas_aromatica]	INAFYES VTLRKACCYARKVEPLKRALAG KRAWITGMRAEQAA-TRGNLAIREYD EGNG LEKFNPLSD	175
[Sinorhizobium_meliloti]	MNGFYESVEARHACCGVRKLKPLARALDGASYWITGLRRGQSG-NRATTPFAEADVERGLIKINPLAD	188
[Wolinella_succinogenes]	EWGMRESLENRHRCCRVRKIEPLKEALKGKSAWVTGLRAEQSI-TRAELKSVEFDETFDLLKFNPLST	158
[Bacillus subtilis]	DKLWEREPNOCCYLRKVVPLREALSGHPAWLSGLRRDOGP-SRANTNFLNKDEKFKSVKVCPLIH	173
[Corvnebacterium glutamicum]	LNLYRSNPAACCRMRKVEPLAASLSPYAGWITGLRRADGP-TRAOAPALSLDAT-GRLKISPIIT	171
[Corvnebacterium efficiens]	VGKNLYRSNPTACCRMRKVEPLAASISPYAGWITGLRESDGP-TRACAPALSLDATGRLEISPITT	201
[Nocardioides an JS614]	TREWST	169
[Chloroflowig ourortiogual		156
[Chioroffexus_aurantiacus]		130
[Deinococcus_geothermalis]	-DLTASDPDACCAVRKVTPLQATLRKKAPSALLNARSRDQAA-TRADIPFVETGGTRVKINPLAI	1/4
[Deinococcus_radiodurans]	-DLYASDPDACCAARKVDPLORYLKEQGPSALLNARSRDOAS-TRADIPFVEEGGARREVNPLAH	190
[Nitrobacter_winogradskyi]	-ELWFSDPDSCCRIRKVEPLARALQPFAAWINGRKRFQGG-LRADLPMVEDDGARLKFNPFAN	190
[Nitrobacter_hamburgensis]	DRDLWFSDPDACCRIRKVEPLVRALQPFDAWINGRKRFQGG-LRADIPFVEDDGARLKFNPFAN	208
[Cytophaga hutchinsonii]	RLLFTSDPDYCCYLNKVOPLDAVLPDTDIWINGVRADOSA-VRKSFSVEOPAPHDTIRFHPMLD	154
Salmonella typhimurium	YGKLWEQ-GVEGIEKYNEINKVEPMNRALKELKAQTWFAGLREEQSG-SRAHLPVLAIQRGVFKVLPIID	184
Escherichia coli	YGKLWEO-GVEGIEK YNDINKVEPMNRALKELNAOTWFAGLRREOSG-SRANLPVLAIORGVFKVLPIID	184
Brwinia carotovora	YGKLWDO-GVEGTER YNLLNKVEPMNRALSELNAGTWFAGLEREOSG-SEGELPVLATO	184
Versinia pseudotuberculosis	YGKINEO-GUEGTERWATINEVERMARATEALGAOTWEAGIRREOSG-GESOLDUTALO	184
Zimomonas mobilis		104
Deterbel due luminos		104
Photornabdus luminescens	ICKLWEQ-GVEGIERINQINKVEPMNRALKNIKAQSWFAGLKRQOBE-BRSKLPVLAVQ	104
Vibrio parahaemolyticus	YGKLWEQ-GVEGIEKYNKINKVEPMRRALKELNVGTWFSGLRREQSK-SRAGLPILSIQNGVFKFLPVID	189
Vibrio_vulnificus	YGKLWEQ-GIEGIEKYNKLNKVEPMRRALNELNVKTWFSGLRREQSQ-SRAGLPILSIQNGVFKFLPVVD	189
Vibrio_fischeri	YGKLWEQ-GVEGIEKYNKLNKVQPMRRALDQLEIGVWFSGLRREQSG-SRANLPILSIQNGVFKFLPVLD	184
Vibrio_cholerae	YGKLWEQ-GVEGIERYNQINKVEPMRRALDELNIGTWFSGLRREQSQ-SRASLPILSVQNGVFKFLPVID	187
Photobacterium profundum	HGKLWEQ-GVDGIKQYNRLNKVEPMRRALDELNVSAWFGGLRREQSS-SRASLPVLAIQNGVFKFLPLID	184
Candidatus Blochmannia	YGKLWTQ-GIKGIOKYNAINKVEPMHRALRTLKVETWFAGLRRNQSD-SRKKLPIITIQNGIFKFLPIVD	192
Candidatus Blochmannia florida	YGKLWEO-GIEGIROVNFMNKVKPMRYALKKLKVRVVVSGLRRYOSC-SRKNLLILDMONGMFKFLPIVD	185
Buchnera aphidicola	YGKLWEK-GT EGT DE YNNTNKYOPMNEALNEL SYOTWEAGLEHDOSK-SENIL EYLSTKKGT FKTLPTLD	184
Yanthomonag orvzae	HART WRO - CHURT DOWNNT PRUPERNERAL DEL NUCHWARTAL PROPAGE OF A OT DUGY P	180
Yanthomonas campestris		180
Nanchomonas Campescris		176
Ayrerra_rastrarosa	HCKLWAQ-GIDGINGINILAKVEPARKALBELQVGIWFIGLRRGGS-IKIQISIVQR	170
Thiocapsa_roseopersicina	LGRLWEQ-GADGIERWINRLNKIDPMERALRDLDAGIWFAGLRRQQAN-BRAELPVLRRQDGRIKFHPIID	180
Pseudoalteromonas_haloplanktis	FGKLWEQ-GEAGIKQYNQLNKVEPMTRALKEINAGTWEGLRRDQAS-SRADKQVIEISRGTVKVYPIIE	182
Shewanella_oneidensis	FGQLWEQ-GVEGLERYNRLNKVEPMQRALAELEVGTWFAGLRRSQSS-TREELPILAIHGSRFKLLPIIE	184
Shewanella_frigidimarina	FGKLWEQ-GIDGLEKYNRINKVEPMQRALAELNVGTWFAGLRESQSS-TREALPILAISGKRFKLLPIIE	202
Shewanella denitrificans	FGKLWEQ-GLDGLEQYNRINKVEPMORALGELNIGTWFAGLRRTQAS-SREALPILAIHGSRFKLLPIIE	198
Shewanella amazonensis	FGKLWEO-GLDGLERYNRINKVEPMORALEELDVGTWFAGLRRTOAS-TREALPILATHGKRFKLLPIIE	181
Colwellia psychrerythraea	YGDEWAO-SDDALKAYNRRNKVEPLERGLSDLNANTWFSGVRROOSA-HREGLSVVGTLRGRYKVHPIID	189
Crocosphaera watsonii	YGKLWNOHDLNSLNRYDFLRKYEPMORALKELOATAWLAGLEROOTE-HROSLEKVELOG	201
Prochlorococcus marinus	VGPLWETGSVKDLETVHPTPKVEPLEGALDDLEVHCWAGGVPSDOTD-VPPSMAFLDLT	199
Trichodesmium eruthracum	VGPLWARKDLRALNRVDOTRKVERMORALRELKATAWLAGI.BGPDVK, HDOGI.DETGVO	192
finachagagaug alaratus		172
synechococcus_eiongatus	REALWESDOVED FIRE TO THE TO THE AVER THE AVER OF THE AVER OF THE AVERAGE AVERAG	1/2
Anabaena_variabilis	TAKTGRALWDKDITQFHQVTKIEPLQKGLDELNEVAWITGRRRDQAV-TRANMPVFELDGKGRLKVNPLAT	181
Schizosaccharomyces_pombe	AQKFGEKLWETDESREDFLVKVEPASRAYSDLNVLAVFTGRRRSQGG-ERGSLPIVQLDGPVLKINPLAN	185
Saccharomyces_cerevisiae	ASKYGDFLWEKDDDKYDYLAKVEPAHRAYKELHISAVFTGRRKSQGS-ARSQLSIIEIDELNGILKINPLIN	189
Thermosynechococcus_elongatus	AARYGDKLWETNVEQFHHLTKIEPLQRGLAELHTVAWITGRRRDQAL-TRANMPVVELDKEGRLKINPLAA	186
Neurospora crassa	EALYGEKLYETSEELYDWIAKVEPONRAYQELKVAAVFTGRRRSOGG-ARGSIPVIELDEERGIVKINPLVN	223
Aspergillus fumigatus	SKKYGAKLWETDDOFYDWVAKVEPAORAYRELNVHAVLTGRRRSOGG-KRGDLDVIEVDEAGLIKVNPLAN	216
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CLUSTAL X (1.81) MULTIPLE SEQUENCE ALIGNMENT

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[Mycobacterium tuberculosis]	WIDODVOEN
[Pseudomonas aeruginosa]	MISEEVWGY
[Pseudomonas syringae]	MSSEEIWGY
[Pseudomonas fluorescens]	MTSEEIWGY
[Pseudomonas_putida]	MSSEEVWGY
[Azotobacter vinelandii]	MSSEEIWGY
[Methylococcus_capsulatus]	WTSAQVWDY
[Microbulbifer_degradans]	WTSQQVWDY
[Ceratopteris_richardii]	VTGTDIWSF
[Polygonatum_pubescens]	VDGKDVWNF
[Zea_mays]	VDGKDIWTF
[Lycopersicon_esculentum]	VDGKDIWNF
[Catharanthus_roseus]	VEGKDIWNF
[Brassica_juncea]	VEGNDVWNF
[Arabidopsis thaliana]	VEGNOVWNF
[Lemna_minor]	VDGQDIWRF
[Chiamydomonas_reinhardtii]	MISAEVWNE
[Nocardioides_sp05014_2]	WERFOUNA
[Desullicobaccellum aerophilum]	WWWDWUWOW
[Clostridium thermocellum]	WSEDRUWEY
[Bordetella parapertussis]	WNEADVWSU
[Burkholderia mallei]	WTESDYWAY
[Ralstonia eutropha]	WTEAEVWAY
[Ralstonia solanacearum]	WTEAEVWAY
[Dechloromonas aromatica]	WTEKEVWTY
[Sinorhizobium meliloti]	WGIETIQAH
[Wolinella succinogenes]	WSETEVFEY
[Bacillus_subtilis]	WTWKDIWRY
[Corynebacterium_glutamicum]	WSLEETNEF
[Corynebacterium_efficiens]	WELEETNRE
[Nocardioides_spJS614]	WTDEALAAY
[Chloroflexus_aurantiacus]	WRREQIEAR
[Deinococcus_geothermalis]	WTRERLETY
[Deinococcus_radiodurans]	WTREQLEAY
[Nitrobacter_winogradsky1]	ASKEDIEAL
[Nitrobacter hamburgensis]	ASKADI SAL
Salmonella tunhimurium	WDNRTUYO
Escherichia coli	WDNRTTYON
Erwinia carotovora	WDNRTVYOY
Yersinia pseudotuberculosis	WDNRQVYQY
Zymomonas mobilis	WDNRKIHYY
Photorhabdus luminescens	WDNRRVHQY
Vibrio_parahaemolyticus	WINKDVHYY
Vibrio_vulnificus	WSNKDVHYY
Vibrio_fischeri	WINKEVHYF
Vibrio_cholerae	WINKEVHYY
Photobacterium_profundum	WIDNDIEQU
Candidatus Blochmannia	WNSLQIHKY
Candidatus Biochmannia fiorida	WENDYTYD
Yanthomonas orvisa	WTDPPWWOY
Xanthomonas campestris	WTDRDWWOW
Xvlella fastidiosa	WTDRDTWEY
Thiocapsa roseopersicina	WHRPRRARY
Pseudoalteromonas haloplanktis	WINRDVYON
Shewanella oneidensis	WSNKDVHLY
Shewanella frigidimarina	WSNKDVHMY
Shewanella_denitrificans	WSNKDVHEY
Shewanella_amazonensis	WSNKDVHEY
Colwellia_psychrerythraea	WSNKDVHE
Crocosphaera_watsonii	WDSRDIYHY
Prochlorococcus_marinus	WTPKEVFYY
Trichodesmium_erythraeum	WNARDIYOY
Synechococcus_eiongatus	WERODOWN
Schizosaccharomuces nombe	WEFTEUHN
Saccharomyces cerevisiae	WTEEOVEOV
Thermosynechococcus elongatus	WTRKOTWAY
Neurospora crassa	WSFQOVKAY
Aspergillus_fumigatus	WTFEQVKOY

5]	WTDQDVQEYIADNDVLVNPLVREGYP	IGCAPC	A-KPAEG-	ADPRSGRWQGL	AKTECGLHAS	254
a]	MTSEEVWGYIRMLELPYNSLHERGYI	IGCEPC	R-PVLP	NOHEREGRWWWE - EA	THKECGLHAGNLISKA	267
e]	MSSEEIWGYIRMLELPYNSLHERGFI	IGCEPC	R-PVLP	NOHEREGRWWWE-EA	TOKECGLHAGNLIVRD	244
s]	MTSEEIWGYIRMLELPYNSLHERGFI	IGCEPC	R-PVLP	NOHEREGRWWWE - EA	TOKECGLHAGNIISKA	244
	MSSEEVWGYIRMLELPYNSLHERGFI	IGCEPC.	R-PVLP	NOHEREGRWWWE-ES	TOKECGLHAGNLISKA	244
1	MSSEEIWGYIRMLEIPYNSLHERGYI	IGCEPC	R-PVLP	NOHEREGRWWWE-EA	THRECGLHAGNLIGKS	244
8]	WTSAQVWDTTTACETPFNALHLKGIV	ICCEPC	R-PVRP	GUHEREGRWWWE-DA	LKKECGLHIGDASRV-	240
8 J	WTSQQVWDTIRSEGVPINSLHDRGFV	TOCEPC	R-PIGP	MURPEORWWE-EA	TAKECGLEATNVKK	291
	VDCK DUWNET PTMEVDUNCT PTOCYU	TECEPC	P-PULPG-		VAN POOL BYONT SPDP	323
1	VDGKDTWTEL ETMDYPVNAL HAOGYUS	TOCEPC	P-PULPG-	OHEREGRWWWED - A	KAKECGLEKGNIDKDG	326
nl	VDGKDIWNFLRAMNVPVNSLHSOGYV	IGCEPC	R-PVLPG-	OHEREGRWWWED - A	KAKECGLHKGNIKDET	329
sì	VEGKDIWNFLRAMDVPVNTLHSOGYV	IGCEPC	R-PVLPG-	OHEREGRWWWED-A	KAKECGLHKGNIKEET	328
aj	VEGNDVWNFLRTMDVPVNTLHAAGYV	IGCEPC	R-AVLPG-	OHEREGRWWWED - A	KAKECGLHKGNIKESS	330
a]	VEGNDVWNFLRTMDVPVNTLHAAGYV	IGCEPC	R-AVLPG-	QHEREGRWWWED-A	KAKECGLHKGNIKENT	325
r]	VDGQDIWRFLRTMAVPVNSLHSQGYVS	IGCEPC	R-PVLPG-	QHEREGRWWWED-A	KA <mark>KECGLH</mark> KGNIKQDE	330
i]	MTSAEVWNFLRVMKVPTNKLHNCGYI	IGCEPC	R-PVLPN-	QAEREGRWWWED-A	AA <mark>KECGLH</mark> SGNIKKAD	284
2]	WIDEEIERYIADHDVLVNPLAYDGYP	IGCWPC	R-RVAPG-	EDPRSGRWAGT	TKTECGIHS	275
e]	WSEEQVWAYIRRENIPYSSLYNKGFR	IGCQPC	R-AVAPG-	ADVRSGRWWWE-DP	SKKECGLHPSLNPPAK	287
n]	WTWDEVWQYIKKYNLPYCKLYDRGYT	IGCEPC	R-PTFVSL	GDVTGEELRKGRWWWE-KN	APKECGIHCSLEAGS-	251
n]	WSEDRVWEYIKKYNIPYNSLYSKGFR	ICCOPC	R-AVRP	GEDVRSGRWWWE-DP	DKKECGLHARVGQV	239
sj	WNEADVWSVIRALGIPYNPLHDOGYP	IGCEPC	R-AVRP	GEDVRAGRWWWE-SS	DSKECGLHAGNRV1Q1	245
1	WTESDVWATLNAFDVPVNPLHARGIP	TOCEPC	R-AIRP	GEDSRAGRWWWE-SR	DIRECGLHITITPVP-	238
1	WIEABUWAYI PPHAUDUNAT UDYOYD	TOCRDC	P-AUPAG-	POURAGRAMME-SK	Der Boot WATHLOUVE	200
1	WTEREVWTYTKONAVPVNALHDREVP	TCCAPC	P-ATSP-	CEDTRACRWWE-NP	ESKECGLHVKG	239
ii.	WGIETTOAHVAAEGTPUNPLHSRCVP	IGCEPC	R-ATKP	GEPERAGRWWWE -ND	EKRECGLHVPEAASST	257
si	WSETEVFEYIKEHALPLHPLYTEGYR	IGCSPC	R-ATEAG-	BELRAGRWWWE-NP	EHKECGLHLKGAOWTS	227
si	WTWKDIWRYTSRNELDYNPLHDOGYPS	IGCAPC	S-PAFTA-	EDLRSGRWNGM	AKTECGLHE	233
n]	WSLEETNEFIADNNLIDHPLTHOGYPS	IGCETC	L-PVAEG-	ODPRAGRWAGN	AKTECGLHS	231
s]	WSLEETNRFITDNNLIDHPLTHOGYPS	IGCATC	L-PVAEG-	ODPRAGRWAGN	AKTECGLHS	261
4]	WTDEALAAYLHEHALPGNRLTELGYPE	IGCEPC	SLPV-	GTDARSGRWPGQ	HKTECGLHAS	229
s]	WRREQIEAFAQAHHLPVNPLYWAGFK	IGCWPC	R-AVRPG-	EDVRAGRWSGK	GKIECGLWIGEQSL	221
s]	WTRERLETYAREHDLPVNPLYWDGFL	IGCWPCI	R-AVRPG-	EDARAGRWAGK	GKTECGLWAGDNRL	239
s]	WTREQLEAYAAEHDLPVNPLYFDGFL	IGCWPC	R-AVKPG-	EDARAGRWAGK	GKTECGLWQGENKL	255
1]	ASREDIEAIYASAKLPPHPLVASGFR	VGCMPC	S-RGQAG-	EDARAGRWRG R	AKTECGIHTMKIP	254
sj	ASRADIEAIYASAKLPPHPLVASGFL	VGCMPC	S-RAQAG-	EDARAGRWRDR	AKTECGIHTMKTS	272
11	WDVRMIEKYIKEHKIPRHPLEEKGIL	ICCEPCI	R-KFDLET	MARRED PROT	NATECGLNTELVSKK-	220
1.4	WDNRTVIQILQKHGLXINPLWDQGIL	UCDTHT	R-KWEPG-	MAREPTREET	VPROGLARG	244
ra	WONRTY YOYLKENGLSYHPLWDOGYL	VCDTHT	P-KWEPG-	MSERETPERGI	KPRCGLWRG	244
is	WDNROVYOYL TOHGLS YHPLWEOGYL	VGDTHT	R-KWEPG-	MSEETREEGL	-KRECGLHEG	244
is	WDNRKIHYYLKEHDLPYHPLWDEGYL	VGDTHT	R-KWEEG-	MSEEETRFFGL	-KRECGLHEG	244
ns	WDNRRVHOYLTKHGLEYHPLWEOGYL	VGDIHT	O-KWEPG-	MSBEOTRFFGL	-KRECGLHEN	244
us	WINKDVHYYLEQHGLTYHPLWEEGYL	VGDTHT	R-KWEPG-	MSEEETRFFGL	-KRECGLHEDDGNEQD	255
us	WSNKDVHYYLKEHGLSYHPLWEQGYLS	VGDTHT	Q-KWEPG-	MSEEETRFFGL	- KRECGLHEED - NEQD	254
ri	WINKEVHYFLKEYDLPYHPLWDQGYLS	VGDTHT	Q-KWEPG-	MSEEETRFFGL	- KRECGLHEDD - GELD	249
ae	WINKEVHYYLKDNDLPYHPLWEQGYL	VGDTHT	Q-KWQPG-	MNEEQTRFFGL	- KRECGLHEDH - NDTH	252
m	WIDNDIEQYLNQYDLPYHPLRDEGYL	MODTHT	K-KWEPG-	MTBEETRFFGL	-KRECGLHEED-AESD	249
ia	WNSLQIHRYIEKHSLEYHPLWQQGYV	ICDVHT	R-KWEPG-	MKEEDTRFFGL	-ORECGLHIIE	253
da	WDHNQVHQYIKRYDLEYHPLWKQGYV	COVHT	I-KKEPG-	MKDEETRFFGL	-ORECGLEVID	246
la	WERDKIKDILKENNLDIHPLINNGIS	TOPRHT	R-KHMPG-	MEREDTREEST	- KRECGLEBN	244
ie	WTDRDWWOYLOAHAL DYHDI WFOGYUS	TOPPHT	P-PWFPG-	MEREDIRFEGL	*PECCIMEDI	241
sa	WTDRDIWEYMKHHDLPYHPLWEOGYVS	TODIHT	R-PLEPG-	MREEDTREEGE	-KRECGINENI	237
na	WHRPRRARYLRRHDLPDHPLRDOGYV	GDVHT	V-PLLPG-	MLEEETRFFGI	-KRECGLHR	239
is	WINRDVYOYLTKHNLPYHPLWEEGYV	MGDIHTT	R-KLEPG-	MTEEETRFFGL	-NRECGLHIDGDGI	246
is	WSNKDVHLYLTQFDLPYHPLWEQGYVS	VGDTHS	K-PLELG-	MTEEETRFNGL	-KRECGLHYEI	245
na	WSNKDVHMYLTEHDLPYHPLWEQGYVS	VGDTHS	K-PLELG-	MSEEETRFNGL	- QRECGLHFDI	263
ns	WSNKDVHEYLTLHGLPYHPLWEQGYV	VGDTHS	K-PLELG-	MSEEDTRFNGL	-KRECGLHYEI	259
is	WSNKDVHEYLTKFGLPYHPLWEQGYVS	VGDTHS	K-PLELG-	MSEEDTRFNGM	-KRECGLHYEI	242
ea	WSNKDVHEYLTKHNLPYHPLWDEGYV	VGDVHS	K-PLTLG-	MSESDTRFGGG	- QRECGLHTDGDGI	253
11	WDSRDIYHYLTAHDLPYHPYFDKGYV	VGDWHS	RPLMADD -	ESERDSRFHGV	- KQECGLHLNLSPGAA	268
us	WTPKEVFYYMQEHQLPQHPLFEKGYST	VGDWHSS	A - PDGGG -	ASGRETRFGGL	- KOECGLHLPGVLGEG	265
m	WNARDI IQILTAHNLPIHPFPDLGIT	VGDWHS	R-PLTTDL	UDERDTRFYGL	- KOECGLHLPQTQEBA	259
is	WTPODSWAYVAFHOVI VNDI HDOGYD	TODEPT	T-KUGEC	EDEPAGEWPCCH	KTRCGTHT	241
De	WSFTEVHNYTTTNNYPYNELLNKGYD	VCDWHS	O-PVREG	EDERAGRWRGDE	-KTRCGLHSHPOSKPA	252
10	WTFEOVKOYIDANNVPYNELLDLOVP	GDYHS	O-PVKEG-	EDERAGRWKGKA	-KTECGIHEASRFAOF	256
us	WTRKOTWAYVMEHOVIYNPLHDRGYA	GDEPL	T-PLANG-	EDERAGRWRGMG	-KTECGIHI	246
sa	WSFQQVKAYIDEHQVPYNELLDKGYK	GDWHS	S-PVAAG-	EDERAGRWKGK	NKTECGIHNKQSRYAQ	290
us	WTFEQVKQYIKENDVPYNELLDRGYK	IGDWHST	Q-PVKEN-	EDERSGRWKGQ	EKTECGIHNPRSKYAQ	283
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CLUSTAL X (1.81) MULTIPLE SEQUENCE ALIGNMENT





Supplementary Fig. 4a



Supplementary Fig. 6



Supplementary Figs. 4 b and c







Supplementary Fig. 5