#### Results

## Normal morphology and unchanged macrophage marker in MORE-PGKO aortas

Aortas from MORE-PGKO mice did not show significant morphological changes compared to LC (Online Figure I A). Inflammation has been considered as an important factor in blood pressure regulation and PPAR-γ is associated with both inflammation and blood pressure regulation (1). We stained aortas with antibodies recognizing macrophage markers. The results showed that F4/80 staining was entirely absent in aortas of both LC and MORE-PGKO mice while the spleen from LC mice served as a positive control with macrophage stained in reddish brown (Online Figure I B). Comparable results were observed using the BM-8 antibody (data not shown) that detects a similar but non-identical macrophage population. Staining with two additional antibodies that detect macrophage, Mac3 and EH-HR3, did not reveal differences between MORE-PGKO and LC aortas either (data not shown). Further, F4/80 protein and mRNA expression levels were not changed in MORE-PGKO aortas compared with LC mice (Online Figure I C~E).

#### Methods

## Histologic analysis

For immunohistochemical staining, sections were incubated at 58°C for one hour, deparaffinized and incubated in antigen retrieval solution (Dako) at 100°C for 14 min. Sections were blocked sequentially with serum, avidin and biotin solutions followed by incubation with primary antibodies against F4/80 (8.33 µg/ml, Bachem), BM-8 (5 µg/ml, Bachem), Mac3 (0.3 µg/ml, BD-Pharmingen), or EH-HR3 (10 µg/ml, Bachem) at 4°C overnight. Staining was visualized using streptavidin-peroxidase and DAB and the sections were counterstained with methyl green/alcian blue. Digital images were obtained using an Olympus BX51 microscope (Olympus America Inc.).

#### Gene expression analysis

F4/80 gene was assayed using reverse transcription quantitative PCR (RT-QPCR) as described (2). All primer-probe sets were purchased from Applied Biosystems.

#### Western blot analysis

Total protein was isolated from aortas, subjected to electrophoresis, and transferred to PVDF membranes as described before (3, 4). Membranes were incubated with primary antibodies recognizing F4/80 (Abcam Inc.), and Actin (Santa Cruz Biotechnology Inc.). The membranes were then incubated in secondary antiserums conjugated with horseradish peroxidase and detected as before (4).

# **Statistics**

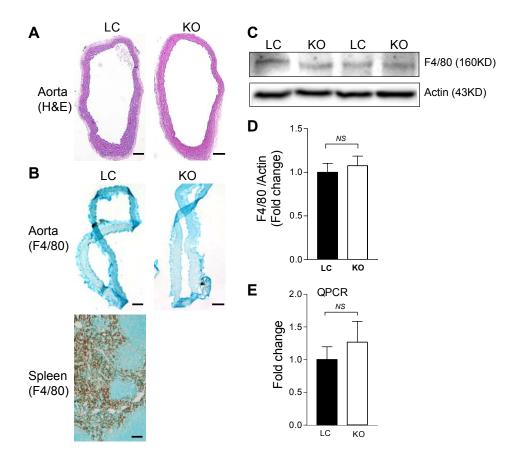
Mean  $\pm$  standard error (SE) values were analyzed using Prism (GraphPad Software Inc.). Statistical comparisons between groups were performed by Student's t test. Groups were considered significantly different if P values were  $\leq 0.05$ .

# References

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# Figure legends

Online Figure I. Normal morphology and unchanged macrophage marker in MORE-PGKO aortas. A. Representative H&E histochemistry of aortas from male LC and MORE-PGKO mice. LC: littermate control; KO: MORE-PGKO. The bars indicate 20 m. B. Representative immunohistochemical staining of F4/80 (a macrophage marker) in male mice showing negative results in aortas. Spleen was used as a positive control with reddish brown staining of macrophages. The bars indicate 20 m. C. Representative Western blots of F4/80 in aortas from male LC and MORE-PGKO mice. Actin was used as a loading control. D. Quantification of F4/80 over Actin. n = 4 for each group. NS: not significant. E. RT-QPCR result of F4/80 expression in male LC and MORE-PGKO aortas. β-actin was used as an endogenous control. n = 4 for each group. NS: not significant. The results from female mice were similar for A~E.



Online Figure I