Comparative In Vitro Activity of Ceftizoxime, Cefoperazone, and Cefoxitin Against Anaerobic Bacteria

MURRAY W. DRULAK AND ANTHONY W. CHOW†*

Division of Infectious Disease, Department of Medicine, University of British Columbia and Vancouver General Hospital, Vancouver, British Columbia, V57 1M9

Received 15 May 1981/Accepted 11 August 1981

Against 482 obligate anaerobes studied by the agar dilution technique, ceftizoxime was significantly more active than both cefoxitin and cefoperazone (P < 0.001); the latter two agents were comparable in activity. The enhanced activity of ceftizoxime, as compared with the activity of cefoxitin, was against both grampositive and gram-negative anaerobes (especially Lactobacillus and Bacteroides spp.). Cefoperazone, however, was more active than cefoxitin against gram-positive anaerobes (particularly Lactobacillus spp.) but was less active than cefoxitin against gram-negative anaerobes (particularly Bacteroides fragilis and Veillonella spp.).

Ceftizoxime (FK-749) and cefoperazone are two newer cephalosporins with enhanced in vitro activity against a wide variety of gram-positive and gram-negative organisms, including nosocomial pathogens such as Serratia and Enterobacter spp. (6, 7). Both antimicrobial agents exhibit stability to beta-lactamase activity, although ceftizoxime has been reported to be more stable than cefoperazone to beta-lactamase hydrolysis (1, 9). Their in vitro activities against clinically important anaerobic bacteria, however, have not been extensively investigated (1, 3, 5, 7, 8). We report here the in vitro activities of ceftizoxime and cefoperazone against 482 isolates of anaerobic bacteria, as determined by an agar dilution technique; the activities of both agents were compared with that of cefoxitin. These isolates were obtained from hospitalized patients of Harbor-University of California at Los Angeles Medical Center and Vancouver General Hospital, British Columbia, Canada, during 1975 to 1980. Identification to the species level was carried out in prereduced, anaerobically sterilized media by the methods of Holdeman et al. (4). The isolates were stored in 20% skim milk and frozen at -75°C until ready for antimicrobial agent susceptibility testing. Susceptibilities to ceftizoxime, cefoperazone, and cefoxitin were determined in Wilkins-Chalgren media (Difco Laboratories, Detroit, Mich.) by the method of Sutter et al. (11), with minor modifications. Twofold serial dilutions of sensitivity powder were added, and the final anti-

† Address reprint requests to: Division of Infectious Disease, G. F. Strong Research Laboratories, Vancouver General Hospital, Vancouver, British Columbia, Canada V57 1M9.

biotic concentrations ranged from 0.25 to 128 μg/ml. A 48-h subculture of the test organism in prereduced, anaerobically sterilized thioglycolate broth was adjusted to a McFarland number 1 nephelometer standard (2), previously determined to approximate 10⁶ to 10⁷ organisms per milliliter, as determined by colony count in roll tubes and prereduced, anaerobically, sterilized media. Inocula (0.0025 ml) were delivered with a Steers replicator (10). Plates were incubated at 37°C in anaerobic jars after air had been evacuated and replaced with a gas mixture of 80% nitrogen, 10% carbon dioxide, and 10% hydrogen. Anaerobic, microaerophilic (incubated in candle jars), and aerobic plates without antibiotics were used for growth and contamination controls, and two reference strains (Bacteroides fragilis 3186 and Clostridium perfringens 3000) with known minimal inhibitory concentrations (MICs) were included in each test for determining reproducibility. All results were read at 48 h, and the MIC recorded was the lowest antibiotic concentration which yielded no visible growth. We compared the in vitro activities by geometric mean MICs, using the Student's t test (two tailed) with a significance level of P < 0.01.

MICs of the control strains (B. fragilis 3186 and C. perfringens 3000) determined in 15 separate tests agreed with the modal MIC \pm 1 log₂ dilution from 87 to 100% for all three antibiotics studied. The antibiotic concentrations required to inhibit 50 (MIC₅₀) and 90% (MIC₉₀) of the strains and the MIC ranges of the study drugs are summarized in Table 1. Overall, ceftizoxime (geometric mean MIC, 1.13 μ g/ml) was more active than cefoxitin (geometric mean MIC, 1.84

Table 1. Comparative in vitro activities of ceftizoxime, cefoxitin, and cefoperazone against anaerobic bacteria

Organism (no.)	Antibiotic	MIC ₅₀ (μg/ml)	$MIC_{90} (\mu g/ml)$	MIC range (μg/m
Bacteroides fragilis (31)	Ceftizoxime	7.4	52	1->128
	Cefoxitin	12	16	8-32
	Cefoperazone	39	>128	1->128
B. vulgatus (19)	Ceftizoxime	0.4	4	0.25-64
	Cefoxitin	3.1	13	0.25-8
	Cefoperazone	14	30	0.25->128
B. distasonis (17)	Ceftizoxime	0.4	16	0.25-64
	Cefoxitin	10	37	1-64
	Cefoperazone	18	90	0.25-128
B. thetaiotaomicron (11)	Ceftizoxime	15	46	0,25-64
	Cefoxitin	25	52	16-64
	Cefoperazone	58	114	8-128
B. ruminicola (14)	Ceftizoxime	6.7	41	0.05.64
	Cefoxitin	11	41 52	0.25-64
	Cefoperazone	26	52 41	0.25-64 0.25-128
	coroporazone	20	41	0.20-126
B. corrodens (10)	Ceftizoxime	< 0.25	1.0	0.25->128
	Cefoxitin	0.5	1.5	0.25-2
	Cefoperazone	< 0.25	16	0.25->8
Other Bacteroides spp. (23)	Ceftizoxime	0.4	20	0.25->128
	Cefoxitin	2	22	0.25-32
	Cefoperazone	0.5	53	0.25-64
Fusobacterium spp. (9)	Cefitzoxime	0.7	35	0.25-64
	Cefoxitin	2.5	18	0.25-32
	Cefoperazone	0.7	26	0.25-128
Veillonella spp. (14)	Ceftizoxime	0.7	1.8	0.25-32
	Cefoxitin	0.5	2.5	0.25-4
	Cefoperazone	4.0	14	0.25-64
Peptostreptococcus spp. (29)	Ceftizoxime	< 0.25	2.0	0.25-64
	Cefoxitin	< 0.25	3.4	0.25-64
	Cefoperazone	0.3	2.0	0.25-64
Peptococcus spp. (84)	Ceftizoxime	<0,25	1.0	0.25-64
	Cefoxitin	<0.25	0.8	0.25-8
	Cefoperazone	<0.25	1.4	0.25-8
Clostridium perfringens (27)	Ceftizoxime	0.7	2.0	
owou waan perji ageno (21)	Cefoxitin	0.7	2.3 1.0	0.25-8 0.25-16
	Cefoperazone	0.3	2.7	0.25-16 0.25-4
Other Classification (00)	0.61			
Other Clostridium spp. (26)	Ceftizoxime Cefoxitin	4.0	30	0.25-128
	Cefoperazone	8.0 8.0	43 53	0.25-64 0.25-64
·6.1.1 4 · · · (0)	- ·			
Bifidobacterium spp. (9)	Ceftizoxime	<0.25	18	0.25-32
	Cefoxitin Cefoperazone	0.7 <0.25	70 2.5	0.25-128 0.25-8
	•			0.20-0
Eubacterium spp. (19)	Ceftizoxime	0.6	22	0.25-32
	Cefoxitin Cefoperazone	3.4	8	0.25-16
	Ceroperazone	0.5	99	0.25->128
Actinomyces spp. (8)	Ceftizoxime	0.25	21	0.25-32
	Cefoxitin Cefoperazone	0.5 0.5	40 1.6	0.25-64
	Ceroperazone	0.5	1.0	0.25–2
Propionibacterium spp. (94)	Ceftizoxime	< 0.25	< 0.25	0.25-1
	Cefoxitin	< 0.25	0.5	0.25-16
	Cefoperazone	< 0.25	0.4	0.25-1
Lactobacillus spp. (38)	Ceftizoxime	36	64	0.5->128
	Cefoxitin	>128	>128	1->128
	Cefoperazone	11	31	0.5-128

 μ g/ml) (P < 0.001), whereas cefoperazone (geometric mean MIC, 1.81 μ g/ml) was comparable to cefoxitin. At concentrations readily achieved in serum for these antibiotics (16 μ g/ml), similar cumulative percentages of the total number of isolates were inhibited by ceftizoxime (85%), cefoperazine (80%), and cefoxitin (85%).

Considerable variation in susceptibility was observed for each antibiotic among different genera or species of anaerobes tested. Among Bacteroides spp., B. fragilis was more resistant than B. vulgatus (P < 0.001) but more susceptible than B. corrodens (P < 0.001) and was comparable in activity to B. ruminicola for all three antibiotics. B. fragilis was more resistant than B. distasonis to ceftizoxime (P < 0.001), whereas B. fragilis was more susceptible than B. thetaiotaomicron (P < 0.001) to cefoxitin. Among anaerobic cocci, Veillonella spp. were more resistant than Peptococcus spp. (P <0.001) to cefoperazone and cefoxitin, whereas no significant difference in resistance to ceftizoxime was noted. Among gram-positive bacilli, C. perfringens was consistently more susceptible than non-perfringens species of Clostridia to all three antibiotics (P < 0.001). Similarly, Lactobacillus spp. were consistently more resistant than other non-spore-forming anaerobic gram-positive bacilli (Propionibacterium, Bifidobacterium, Eubacterium, and Actinomyces spp.) to all three antibiotics (P < 0.005).

The comparative in vitro activities of these agents indicated that ceftizoxime was more active overall than cefoxitin (P < 0.001), and this enhanced activity was against both gram-positive (particularly Lactobacillus spp.) and gramnegative anaerobes (particularly Bacteroides spp.) (P < 0.005). Cefoperazone, on the other hand, was more active than cefoxitin against gram-positive anaerobes (particularly Lactobacillus spp.; P < 0.001) but was less active than cefoxitin against gram-negative anaerobes (particularly B. fragilis and Veillonella spp.; P < 0.005). The enhanced in vitro activities of these newer cephalosporins, particularly ceftizoxime,

against a wide variety of anaerobic bacteria, coupled with their broadened activity against many aerobic organisms, offers considerable promise for their use in single-agent therapy for mixed aerobic-anaerobic infections. Controlled clinical trials are indicated.

LITERATURE CITED

- Brown, J. E., V. F. Del Bene, and C. D. Collins. 1981. In vitro activity of N-formimidoyl thienamycin, moxalactam, and other new beta-lactam agents against Bacteroides fragilis: contribution of beta-lactamase to resistance. Antimicrob. Agents Chemother. 19:248-252.
- Finegold, S. M., W. I. Martin, and E. G. Scott. 1978.
 Baily and Scott's diagnostic microbiology, 5th ed., p. 488–489. C. V. Mosby, St. Louis.
- Fu, K. P., and H. C. Neu. 1980. Antibacterial activity of ceftizoxime, a beta-lactamase-stable cephalosporin. Antimicrob. Agents Chemother. 17:583-590.
- Holdeman, L. V., E. P. Cato, and W. E. C. Moore. 1977. Anaerobe laboratory manual, 4th ed. Virginia Polytechnic Institute Anaerobe Laboratory, Blacksburg.
- Jacobus, N. V., F. P. Tally, M. Barza, and S. L. Gorbach. 1980. Susceptibility of anaerobic bacteria to cefoperazone and other beta-lactam antibiotics. Clin. Ther. 3:34-38.
- Jones, R. N., P. C. Fuchs, A. L. Barry, T. L. Gavan, E. H. Gerlach, and H. M. Sommers. 1980. Antimicrobial activity and spectrum of cefoperazone against recent clinical isolates. Clin. Ther. 3:14-23.
- Kamimura, T., Y. Matsumoto, N. Okada, Y. Mine, M. Nishida, S. Goto, and S. Kuwakara. 1979. Ceftizoxime (FK 749), a new parenteral cephalosporin: in vitro and in vivo antibacterial activities. Antimicrob. Agents Chemother. 16:540-548.
- Kaye, D., W. Dobasa, and K. Kaye. 1980. Susceptibilities of anaerobic bacteria to cefoperazone and other antibiotics. Antimicrob. Agents Chemother. 17:957-960
- Kojo, H., M. Nishida, S. Goto, and S. Kuwahara. 1979. Antibacterial activity of ceftizoxime (FK 749), a new cephalosporin, against cephalosporin-resistant bacteria and its stability to β-lactamase. Antimicrob. Agents Chemother. 16:549-553.
- Steers, E., E. L. Foltz, B. S. Grave, and J. Riden. 1959.
 An inocula replicating apparatus for routine testing of bacterial susceptibility to antibiotics. Antibiotics Chemother. 9:307-311.
- Sutter, V. L., A. L. Barry, T. D. Wilkins, and R. J. Zabransky. 1979. Collaborative evaluation of a proposed reference dilution method of susceptibility testing of anaerobic bacteria. Antimicrob. Agents Chemother. 16:495–502.