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3	LEGENDS TO SUPPLEMETARY FIGURES
4	fig. S1. Tyrosine phosphorylation at Y474 of Tir is required for efficient actin
5	polymerization and the recruitment of ZO-1, but not for disruption of the epithelial barrier.
6	(A) Production of actin-rich pedestals by EPEC is dependent on the tyrosine phosphorylation of
7	Tir at Y474. HeLa cells were infected by $\Delta tir$ containing p99-tir ( $\Delta tir$ /Tir) or p99-tirY474F.
8	( $\Delta tir$ /TirY474F), as described in the Materials and Methods. The cells were fixed and stained
9	for phosphotyrosine with a monoclonal antibody 4G10 (green), for F-actin with
10	rhodamine-phalloidin (red), and for EPEC with anti-intimin antiserum (blue). When the cells
11	were infected with $\Delta tir/Tir$ expressing a wild-type Tir, anti-phosphotyrosine antibody
12	detected phosphorylated Tir at the interface between the bacteria and pedestals (upper row),
13	whereas no phosphorylation of Tir was detected with $\Delta tir/TirY474F$ expressing a mutant Tir
14	with a substitution of Y474 by F474 (lower row). Scale bar, 5 mm.
15	(B) Tyrosine phosphorylation at Y474 is required for ZO-1's recruitment. HeLa cells were
16	infected with a wild-type EPEC (WT), $\Delta tir$ , or $\Delta tir$ /Tir or $\Delta tir$ /TirY474F, as described in the
17	Materials and Methods. The cells were fixed and stained for ZO-1 (green) and F-actin (red),
18	and for bacteria with anti-intimin antiserum (blue). It is notable that no recruitment of ZO-1
19	was observed in the absence of Tir, and even in its presence, the substitution at Y474 of Tir

20 abolished the recruitment. Scale bar, 5  $\mu$ m.

<ol> <li>13</li> <li>14</li> <li>15</li> <li>16</li> <li>17</li> <li>18</li> <li>19</li> </ol>	<ul> <li>HeLa cells expressing NZO-1 (upper row), control mEGFP (middle row), or mEGFP-PRR (lower row) were infected by <i>S. flexneri</i> (panel A) or <i>L. monocytogenes</i> (panel B) as described in the Materials and Methods. F-actin and bacteria were visualized with rhodamine-phalloidin (red) and DAPI (blue), respectively. NZO-1 and PRR were detected with anti-VSVG tag and with the fluorescence of mEGFP, respectively. Scale bars, 5 μm.</li> <li>fig. S3. Clustering of the membrane-targeted Nck SH3 domains induces the recruitment of</li> </ul>
<ol> <li>13</li> <li>14</li> <li>15</li> <li>16</li> <li>17</li> <li>18</li> </ol>	HeLa cells expressing NZO-1 (upper row), control mEGFP (middle row), or mEGFP-PRR (lower row) were infected by <i>S. flexneri</i> (panel A) or <i>L. monocytogenes</i> (panel B) as described in the Materials and Methods. F-actin and bacteria were visualized with rhodamine-phalloidin (red) and DAPI (blue), respectively. NZO-1 and PRR were detected with anti-VSVG tag and with the fluorescence of mEGFP, respectively. Scale bars, 5 µm.
<ol> <li>13</li> <li>14</li> <li>15</li> <li>16</li> <li>17</li> </ol>	HeLa cells expressing NZO-1 (upper row), control mEGFP (middle row), or mEGFP-PRR (lower row) were infected by <i>S. flexneri</i> (panel A) or <i>L. monocytogenes</i> (panel B) as described in the Materials and Methods. F-actin and bacteria were visualized with rhodamine-phalloidin (red) and DAPI (blue), respectively. NZO-1 and PRR were detected with anti-VSVG tag and with the fluorescence of mEGFP, respectively. Scale bars, 5 μm.
<ol> <li>13</li> <li>14</li> <li>15</li> <li>16</li> </ol>	HeLa cells expressing NZO-1 (upper row), control mEGFP (middle row), or mEGFP-PRR (lower row) were infected by <i>S. flexneri</i> (panel A) or <i>L. monocytogenes</i> (panel B) as described in the Materials and Methods. F-actin and bacteria were visualized with rhodamine-phalloidin (red) and DAPI (blue), respectively. NZO-1 and PRR were detected
13 14 15	HeLa cells expressing NZO-1 (upper row), control mEGFP (middle row), or mEGFP-PRR (lower row) were infected by <i>S. flexneri</i> (panel A) or <i>L. monocytogenes</i> (panel B) as described in the Materials and Methods. F-actin and bacteria were visualized with
13 14	HeLa cells expressing NZO-1 (upper row), control mEGFP (middle row), or mEGFP-PRR (lower row) were infected by <i>S. flexneri</i> (panel A) or <i>L. monocytogenes</i> (panel
13	HeLa cells expressing NZO-1 (upper row), control mEGFP (middle row), or
12	mediated through the proline-rich region of the ZO-1 molecule.
11	fig. S2. Recruitment of ZO-1 to the actin tails induced by S. flexneri and L. monocytogenes is
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9	experiments.
8	Tir (Y474F) from p99-tirY474F. Data represent the means (± SD) from three independent
7	triangle, $\Delta tir$ expressing a wild-type Tir from p99-tir; open circle, $\Delta tir$ expressing a mutant
6	(Millipore) every two hours. Closed square, wild-type EPEC; open square, $\Delta tir$ ; open
5	at a MOI of 1 to 3. The TERs across the monolayers were measured with Millicell-ERS
4	5 days were infected by either wild-type EPEC or the mutants expressing Tir from plasmids
3	previously (1). Briefly, Caco/B7 monolayers cultured on filter supports (Transwell COL) for
	EPEPC. The measurement of TER across Caco/B7 monolayers was performed as described
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1	NIH-3T3 cells expressing either Nck SH3(3) (upper row) or Nck SH3(1+2+3) (lower
2	row) were treated with anti-CD16 and Alexa546-conjugated anti-rat IgG antibodies. After
3	fixation, the clustered fusion proteins were observed under a fluorescence microscope (CD16,
4	red). F-actin (panel A) and ZO-1 (panel B) were visualized with Alexa488-phalloidin, and
5	T8-754 and Alexa488-anti-mouse IgG antibodies, respectively (green). Insets are a higher
6	magnification of the regions outlined by white dotted lines. Scale bar, 10 $\mu$ m.
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9	REFERENCE
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14	Res. Commun. <b>337:</b> 922-927.
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 ZO-1 fragment
 F-actin
 Merge

 Listeria
 NZO-1
 Image: Comparison of the second of the secon

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