

# **Additional file 1 – Experimental results for human cell cycle and the biological support of the gene regulations**

-Additional supporting analyses for the article: Jung-Hsien Chiang and Shih-Yi Chao:  
**Modeling human cancer-related regulatory modules by GA-RNN hybrid algorithms**

## **Discussions**

Data from Whitfield *et al.* [35] is downloaded from the reference web site. The goal of human cell cycle analysis from Whitfield *et al.* is to identify >850 genes periodically expressed during the cell cycle, and to show that most of these genes have been previously associated with the proliferation of tumors during the human cell division cycle as well. We adopt this data set to construct cancer-related regulatory modules with feedback or feed-forward controlled target genes which are regulated by some specific TFs.

For the purpose of demonstration, we list experimental results for common regulators with [33]. Since yeast transcriptional regulatory mechanisms are more comprehended than Homo sapiens, we can confirm the accuracy of the connections within the regulatory modules provided by our system according to the published biological literatures which listed in the table below. We also provide simple explanations and descriptions for target genes. Some predicted results of target genes represent by boldface indicate the hypothetical targets. As listed in the table below, for instance, the E2F1 is a transcription factor of CCNA2, CDC25A, c-MYC, RFC3, p130, p107 and cycA, and E2F1 regulates the expression of these genes [4][5][7] [19][20].

## **Conclusions**

Microarray technology has produced high-throughput data for exploring, analyzing, and understanding the phenomena of transcriptional regulatory mechanisms. We introduce a Genetic Algorithm-Recurrent Neural Network (GA-RNN) hybrid method for finding feed-forward regulated genes when given some transcription factors. It has proved that the gene regulations identified by GA-RNN have biological evidence supports, and it is a feasible approach to model human cancer-related regulatory modules.

**Table S6 - Experimental results for human cell cycle and the biological support of the gene regulations**

Regulator	Target predicted by [33] Expression data from Whitfield et al., 2002	Target predicted by our approach Expression data from Whitfield et al., 2002	Biological evidences *	Explanations and descriptions
E2F1	BUB1B, E2F1	E2F1 → E2F1	[1][24]	ChIP analysis demonstrated that E2F1 bound to E2F1 promoter chromatin in the G0 and early G1 cell cycle stage.[24]
		E2F1 → RB → cycA	[2][3][31]	E2F interacts with the CycA promoter and 5-prime UTR as demonstrated by chromatin immunoprecipitation (ChIP) assay. [31]
		E2F1 → RB →E2F3	[2][3][6]	The p16/RB/E2F regulatory pathway, which controls transit through of the G1 restriction point of the cell cycle, is one of the most frequent targets of genetic alterations in human cancer. Any of these alterations results in the deregulated expression of the transcription factor E2F, one of the key mediators of cell cycle progression. [2][3][6]
		E2F1→CDC6→CCNA	[2][18][30]	An interaction between E2F1 and CDC6 promoter was demonstrated by chromatin immunoprecipitation assay. Binding of E2F1 to the CDC6 promoter was reduced in MIF-/- EuMyc lymphomas. [30]
		RB, E2F3, CDC25A, CCNA2, P130, c-MYC, P107, P53, RFC3, cycA	[4][5] [7] [19][20][24]	E2F1 interacts with the CCNA2 promoter.[24] E2F1 is a transcription factor of CCNA2, CDC25A, c-MYC, RFC3, p130, p107 and cycA and regulates the expression of these genes. [4][5] [7] [19][20] An interaction between E2F1 and Cdc25A promoter chromatin was demonstrated by chromatin immunoprecipitation. [24]
		<b>RBL2, cyclinD1, CK2, TBL1, rbp6</b>	<b>hypothetical targets *</b>	

CDC25A	CCNE1, CDC20, CDC25A, STK15, BUB1B	CDC25A → CDC25A	[8]	Cdc25A is an important regulator of the G1/S transition but functions also in the mitotic phase of the human cell cycle.
		CCNE1, E2F1, E2F3, P130, MYC	[9] [10] [11][32]	An interaction between Myc and Cdc25a promoter was demonstrated by chromatin immunoprecipitation (ChIP) assay. [32] p130 interacts with the CDC25A promoter
		<b>CCNA1, CDK2</b>	<b>hypothetical targets *</b>	
CDC6	N/A	CDC6 → c-MYC → ORC2	[25][28]	In addition to its DNA replication activity, CDC6 also has a role as a transcriptional suppressor of c-Myc.[25]
		CDC6 → PCAF → E2F1	[24]	[24] investigate the <i>p107</i> , <i>E2F-1</i> , <i>Cdc25A</i> , <i>Cdc6</i> , <i>B-myb</i> , <i>cyclin A</i> , and <i>Cdc2</i> promoters because each of these promoters has been implicated as a target of the E2F and pRB family on the basis of genetic and/or biochemical criteria.
		CDC6, E2F1, ORC2, c-MYC, P107, P130, RBL2, MCM7, E2F3,	[24][25] [26] [27] [29]	An interaction between p130 and Cdc6 promoter chromatin was demonstrated by chromatin immunoprecipitation.[24] some CDC6 protein is associated with the specific nuclear structure throughout the cell cycle and that major binding sites on chromatin differ between MCM and CDC6. CDC6 protein, a candidate regulator of MCM.[27]
		<b>NSEP1, PTHYA, OCD, GCN5, PCNA, ORC1C, AKAP8L, CCNA, NICD,</b>	<b>hypothetical targets *</b>	
PCAF	N/A	PCAF, p53, P21	[12]	Stimulation of p73-mediated transactivation by PCAF requires the HAT domain of PCAF and the p53-binding site within the p21 promoter. <i>In vivo</i> , coexpression of wild-type, but not HAT-deficient PCAF with p73 markedly increases p21 expression.
		<b>CP1C, BRCA2, CBF1, RAR-alpha, CK2,</b>	<b>hypothetical targets *</b>	

		<b>TBL1,</b>		
HDAC3	N/A	HDAC3, NCOR1, YY1, API2, TBL1, P53	[13][14] [15][21][22]	[13] shows that mammalian histone deacetylase (HDAC)-1, -2, and -3 are all capable of down-regulating p53 function. Down-regulation of p53 activity by HDACs is HDAC dosage-dependent, requires the deacetylase activity of HDACs, and depends on the region of p53 that is acetylated by p300/CREB-binding protein (CBP).
		<b>TGIF, TAB2, P50, GATA2, CK2, RBL2</b>	<b>hypothetical targets *</b>	
PCNA	CDC25B, CK2	RFC3, RB, P53, P50	[16] [17]	Lu et al. found that PCNA co-immunoprecipitated with human p50, as well as calf thymus DNA polymerase d heterodimer, but not with p125 alone, suggesting that PCNA directly interacts with p50 but not with p125.[17]
		<b>APC, COP1, PTK2, PTTG1</b>	<b>hypothetical targets *</b>	

\* The gene names without boldface indicate the true positive (TP) targets that confirmed by searching in databases [34] or biological documents provided below.

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