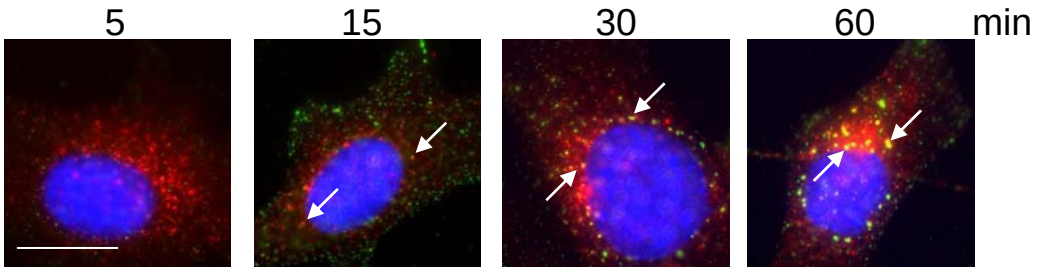
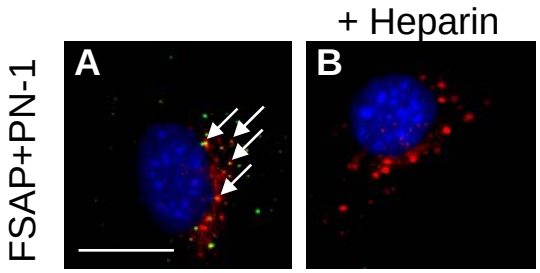
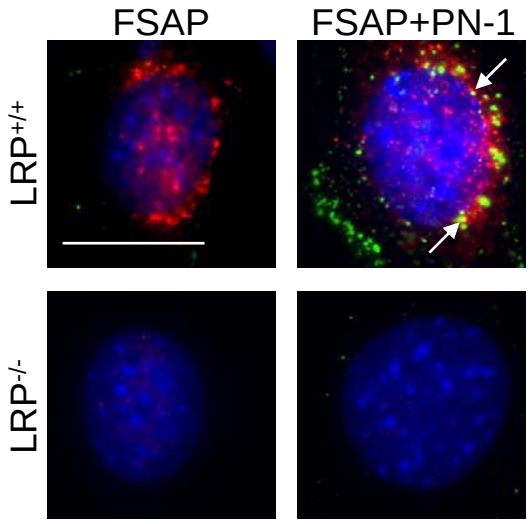


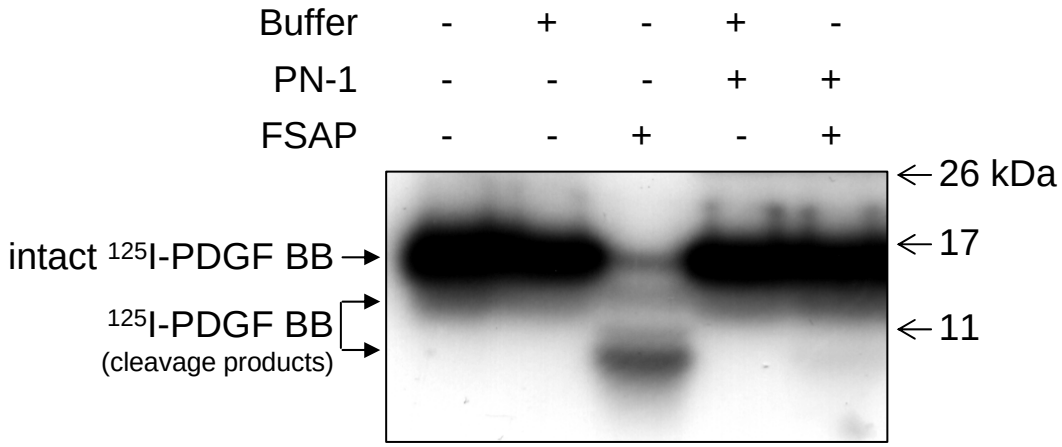
Supplemental data 1: Time-dependent binding and uptake of FSAP-PN-1 complex in VSMC via LRP. 1 µg/ml FSAP and 2 µg/ml PN-1 were preincubated for 30 min and the mixture was added to the cells for indicated times at 37°C. FSAP is stained with FITC-labeled secondary antibody indicated in green, LRP is stained with Rhodamine red-X-labeled secondary antibody indicated in red and nuclei are DAPI stained indicated in blue. Yellow color indicates co-localization of FSAP and LRP. White arrows highlight examples of co-localization. Calibration bars indicates 20 µm. FSAP-PN-1 complex internalization proceeds in a time dependent manner starting at 15 min and reaching a maximum at 60 min. Similar results were obtained in 2 separate experiments.



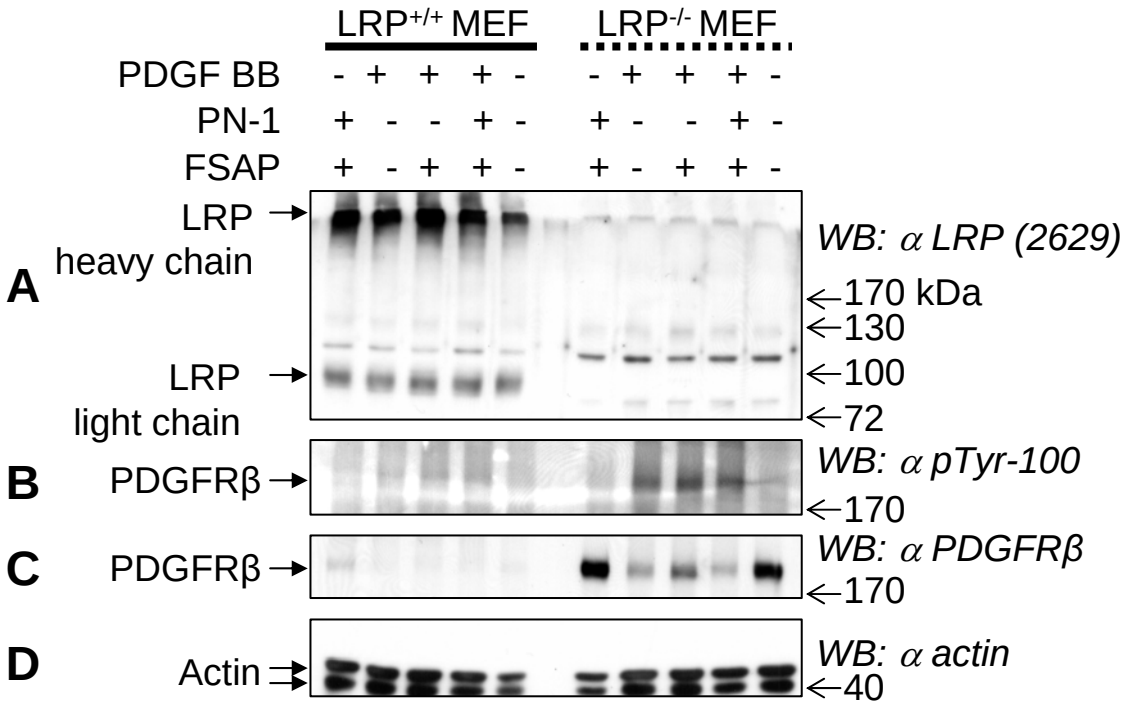
Supplemental data 2: Heparin-dependent binding and internalization of FSAP-PN-1 complex in VSMC via LRP. 1 µg/ml FSAP and 2 µg/ml PN-1 were preincubated for 30 min (A) without or (B) with 10 µg/ml Heparin and the mixture was added to the cells for indicated times at 37°C. FSAP is stained with FITC-labeled secondary antibody indicated in green, LRP is stained with Rhodamine red-X-labeled secondary antibody indicated in red and nuclei are DAPI stained indicated in blue. Yellow color indicates co-localization of FSAP and LRP. White arrows highlight examples of co-localization. Calibration bar indicates 20 µm. FSAP-PN-1 complex interaction with LRP and therefore internalization via LRP is blocked by Heparin.



Supplemental data 3: Internalized FSAP-PN-1 complexes via LRP in mouse embryo fibroblasts (MEF). 1 µg/ml FSAP and 2 µg/ml PN-1 were preincubated for 30 min and the mixture was added to $LRP^{+/+}$ MEF (upper panels) and $LRP^{-/-}$ MEF (lower panels) for 45 min. FSAP is stained with FITC-labeled secondary antibody indicated in green, LRP is stained with Rhodamine red-X-labeled secondary antibody, nuclei are stained with DAPI indicated in blue. Yellow color indicates co-localization of FSAP and LRP. White arrows highlight examples of co-localization. Calibration bar indicate 20 µm. The lack of staining of $LRP^{-/-}$ cells with the α **LRP antibody** indicates specificity of staining **with this antibody**.



Supplemental data 4: ^{125}I-PDGF BB-binding to VSMC: ^{125}I-PDGF BB was incubated with buffer, 1 µg/ml FSAP or FSAP + 2 µg/ml PN-1 for 30 min and then added to VSMC for 60 min on 37°C. After extensively washing the cells the cell associated radioactivity was recovered by lysis and SDS-PAGE was performed under reducing conditions followed by autoradiography. FSAP alone induced cleavage of ^{125}I-PDGF BB and this was inhibited by PN-1. Intact ^{125}I-PDGF BB bound to cells but cleaved ^{125}I-PDGF BB did not.



Supplemental data 5: Effect of FSAP-PN-1 complex on PDGF-BB mediated stimulation of LRP$^{+/+}$ and LRP$^{-/-}$ MEF. 1 µg/ml FSAP and 2 µg/ml PN-1 were preincubated for 30 min. 20 ng/ml PDGF BB was then preincubated with either buffer, FSAP or FSAP-PN-1 for 60 min and the mixtures were used for stimulating cells for 10 min. MEF were lysed with lysis-buffer and processed for Western Blot analysis. Heavy chain and light chain of LRP is expressed in LRP$^{+/+}$ MEF but not in LRP$^{-/-}$ MEF (A). Since LRP internalizes PDGFβR, more PDGFβR is present in LRP$^{-/-}$ than LRP$^{+/+}$ cell (C). Stimulation of cells with PDGF-BB leads to tyrosine phosphorylation of PDGFβR (B) and its internalization and degradation (C). FSAP or FSAP-PN-1 complex do not influence this process. Actin blot shows equal loading of cell extracts in all lanes (D).