

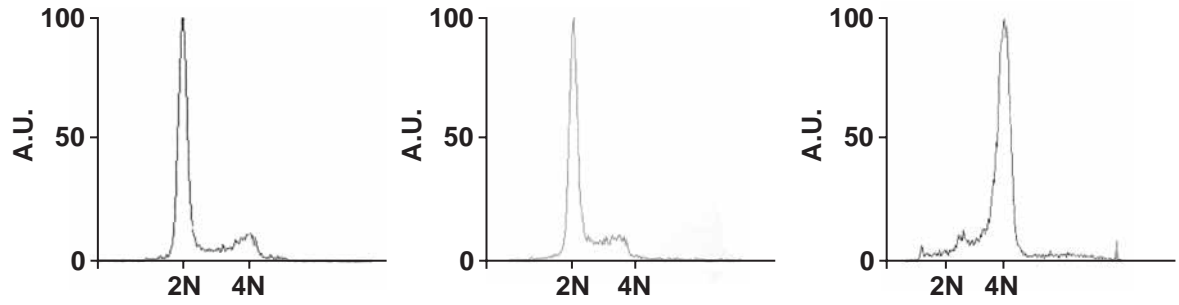
## Supplementary Data

### Supplementary Figure S1. FACS analysis of HeLa cells after synchronization in S and G2.

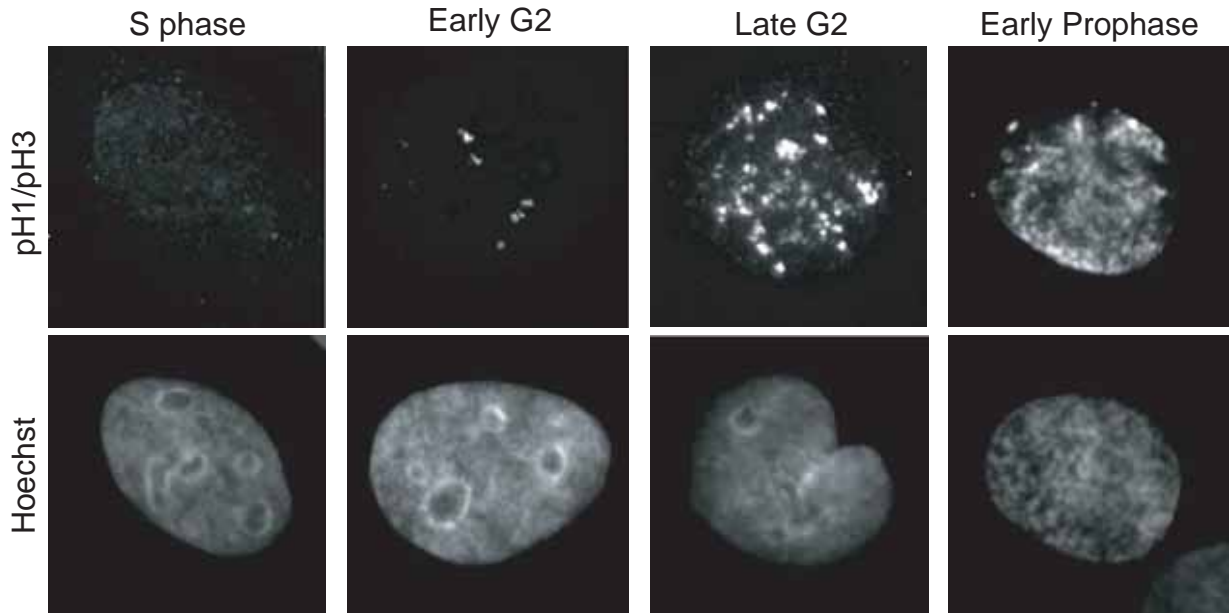
(A) Asynchronously growing HeLa cells (left panel) were arrested in S phase (central panel) or G2 (right panel); see Methods for details. DNA stained with propidium iodide was sorted using a coulter Elite FACScan. (B) HeLa cells were grown on coverslips, treated to be blocked in G2 or arrested in S-phase by double thymidine/aphidicolin block and, at various times after releasing the block, permeabilized by incubating for 2 min at room temperature in 0.1% Triton X-100 in PBS. The cells were then fixed and labelled with anti-phosphohistone-H1 and -H3 polyclonal antibodies (pH1/pH3), as markers of different cell-cycle phases (Hidalgo Carcedo *et al*, 2004), and with Hoechst to label DNA. Images were acquired using an inverted Axiophot equipped with a digital camera. Representative images of cells found in S phase (diffuse nuclear staining, uncondensed DNA), early G2 (faint staining of very few pericentromeric foci, uncondensed DNA), late G2 (strong staining at several centromeric foci, uncondensed DNA) and early prophase (strong staining extended to the chromosome arms, low level of chromosome condensation). (C) Relative percentages of mitotic (white) and G2 (black) populations in untreated HeLa cells (NS), and in HeLa cells treated with bisbenzimidazole (Bis-bi) and aphidicolin (Aph), and following aphidicolin washout, as indicated (see also Methods). More than 300 cells in three different experiments were evaluated as detailed in (B). The standard deviation never exceeded 7%.

Figure S1

**A**



**B**



**C**

