IGF2 Expression is a Marker for Paraganglionic/ SIF Cell Differentiation in Neuroblastoma

Fredrik Hedborg,*^{†‡§} Rolf Ohlsson,^{†‡} Bengt Sandstedt,[¶] Lars Grimelius,* Jeff C Hoehner,* and Sven Påhlman*

From the *Department of Pathology, University Hospital, Uppsala; †Department of Drug Dependence Research, Karolinska Hospital, Stockholm; †Department of Developmental Biology, Institute of Zoology, University of Uppsala, Uppsala; *Department of Pediatrics, University Hospital, Uppsala; and *Department of Pathology, Pediatric Unit, Karolinska Hospital, Stockholm, Sweden

Neuroblastoma is a childbood tumor of the sympathetic nervous system. Observations in the Beckwith-Wiedemann syndrome suggest that sympathetic embryonal cells with an abundant expression of the insulin-like growth factor 2 gene (IGF2) may be involved in the genesis of lowmalignant infant neuroblastomas. We have therefore compared the cell type-specific IGF2 expression of the human sympathetic nervous system during early development with that of neuroblastoma. An abundant expression in normal sympathetic tissue was specific to extra-adrenal chromaffin cells, ie, paraganglia and small intensely fluorescent (SIF) cells, whereas sympathetic neuronal cells were IGF2-negative. A subpopulation of neuroblastomas expressed IGF2, which correlated with an early age at diagnosis, an extraadrenal tumor origin, and severe bemodynamic signs of catecholamine secretion. Histologically IGF2-expressing tumors displayed a lobular growth pattern, and expression was restricted to the most mature and least proliferative cells. Typically, these cells were morphologically and histochemically similar to paraganglia/SIF cells and formed distinct ring-like zones in the center of the lobules around a core of apoptosis-like tumor cells. The similarities found between IGF2expressing neuroblastoma cells and paraganglia/SIF cells in terms of bistological features, anatomical origin, and age-dependent growth suggest a paraganglionic/SIF cell lineage of most infant tumors and also of extra-adrenal tumors diagnosed after infancy. Furthermore, since

paraganglia/SIF cells undergo postnatal involution, the same cellular mechanism may be responsible for spontaneous regression in infant neuroblastoma. (Am J Pathol 1995, 146:833–847)

Neuroblastoma is an embryonal tumor of early childhood originating from the developing sympathetic nervous system.¹ Two major tumor types have been well described.¹⁻³ One prognostically unfavorable type usually affects children more than 2 years of age, with invasive growth at diagnosis, and a biological marker profile including a near-diploid DNA content, chromosome 1p deletions, an amplified N-myc gene, and low expression of the c-trk proto-oncogene, coding for the high-affinity receptor for nerve growth factor (NGF). The second type has an excellent prognosis, is usually diagnosed at an age of less than 18 months, grows locally, has a hyperdiploid DNAcontent, and expresses the c-trk proto-oncogene abundantly but lacks 1p deletions and N-myc amplification. Moreover, these two forms differ significantly in their sites of origin. Aggressive tumors are usually localized to the adrenal area, whereas prognostically favorable infant tumors are much more frequently found at extra-adrenal sites,4,5 suggesting an origin from the sympathetic trunk or from prevertebral sympathetic structures in the retroperitoneum. Histological signs of maturation are more frequent in prognostically favorable infant tumors than in the older age group, when criteria such as nuclear enlargement, increased cytoplasm, cell processes,5-7 and presence of fibrillar material (neuropil) are used.⁸ However, the majority of these infant tumors are still mor-

Accepted for publication January 6, 1995.

Address reprint requests to Dr. Fredrik Hedborg, Department of Pediatrics, University Hospital, S-751 85, Uppsala, Sweden.

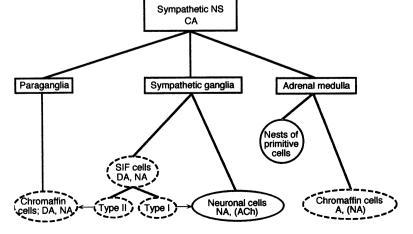
Supported by grants from the Children's Cancer Foundation of Sweden (grants 91.053 and 91.058), the Swedish Cancer Society (grants 1745-B94-06XCC and 1955), the Swedish Medical Research Council (grant 102), HKH Kronprinsessan Lovisas förening för barnasjukvård, Hans von Kantzows and Ollie och Elof Ericssons stiftelser, Research Funds of the Karolinska Institute, Lions Cancer Research Foundation at Uppsala University Hospital, and Gillbergska stiftelsen.

phologically primitive,⁵⁻⁷ although biochemical evidence (c-src splice variant, neuron-specific enolase, and synaptophysin expression) indicates a more mature phenotype than in aggressive neuroblastoma.⁹ Some neuroblastomas display more overt evidence of maturation because of their content of ganglion-like cells, which have a more abundant cytoplasm and a more pronounced nuclear enlargement with a distinct nucleolus. Such evidence of neuronal differentiation is usually accompanied by the presence of Schwann cell-like elements. These tumors, frequently referred to as ganglioneuroblastomas,^{5–7} are usually associated with a diagnosis after infancy and have a better prognosis than histologically immature tumors of the older age group.^{5,7} Neuroblastoma has the highest frequency of spontaneous regression of all human cancers,¹⁰ which is primarily associated with infant disease, particularly metastatic tumors of the 4S type.¹ However, in widespread disease of children more than 1 year old at diagnosis, spontaneous regression is a rare event (reviewed in ref 1). This unique capacity of neuroblastoma may also explain why nonradical surgery usually is curative in regional infant disease,¹¹ and why clinically silent tumors (neuroblastoma in situ) are a frequent autopsy finding in neonates who have died from nonneoplastic causes.12

The constituents and hormone/neurotransmitter production of the sympathetic nervous system are schematically shown in Figure 1. Cells of the sympathetic nervous system include neuronal, paraganglionic, small intensely fluorescent (SIF), and adrenal chromaffin cells. Paraganglia are discrete encapsulated neuroendocrine structures adjacent to sympathetic ganglia.¹³ These structures appear during early sympathetic organogenesis and are of significant size during prenatal development, the largest of which is called the "organ of Zuckerkandl", located along the abdominal aorta.14 In addition, small collections of microscopically and histochemically similar cells are found within all sympathetic ganglia.¹⁵ These cells are denoted SIF cells, a term based upon their intense formaldehyde-induced catecholamine fluorescence. These purely neuroendocrine, paraganglia-like SIF cells are denoted type II, as opposed to another type of SIF cell (type I), which shows mixed neuroendocrine/interneuron features and exists as solitary cells within sympathetic ganglia.¹⁶ In the fetus and the neonate, the predominant source of catecholamine production and release is these extraadrenal sympathetic neuroendocrine cell types, 17, 18 whereas the adrenal medulla is poorly developed both in size^{14,15} and function.¹⁷ During childhood, however, the size of paraganglia and the number of SIF cells decrease significantly.¹⁵ Postnatal paraganglionic involution is observed in several mammalian species,14,18,19 and in the rat this is an early event occurring during the first weeks of life, involving signs of chromaffin cell degeneration such as nuclear irregularity and pyknosis.¹⁹ In contrast, adrenal chromaffin cell growth accelerates after birth and continues until puberty.¹⁵ Catecholamines produced by sympathetic neuroendocrine cells are stored in granular vesicles. These organelles are rich in chromogranins²⁰ and are also responsible for the chromaffin reaction, specific for sympathetic neuroendocrine cells. Adrenal chromaffin cells differ from their extra-adrenal counterparts in that they produce adrenalin, which is the major catecholamine of the adrenal medulla from midgestation.14

Insulin-like growth factor 2 (IGF-2) exerts mitogenic, trophic, and differentiation-promoting effects on neuroblastoma-derived cell lines.^{21–23} This growth factor may also mediate signals for cell survival of

Figure 1. Schematic representation of the constituents of the sympathetic nervous system and their neurotransmitter/bormone production. Neuroendocrine cell types are represented by dashed lines. Similarities in phenotype between SIF type II cells and paraganglia cells, as well as SIF type I cells and sympathetic neurons, are indicated by arrows. Neuroglial cells are omitted. The scheme is based on data presented in refs. 13 to 16. Abbreviations used: A, adrenalin; ACb, acetylcholine; CA, catecholamines; DA, dopamine; NA: noradrenalin; NS: nervous system. Minor production is shown within parentheses.



embryonic sympathetic cells,²⁴ presumably counteracting programmed cell death (apoptosis), as is shown to be the case for IGF-1 in oligodendrocyte survival in fetal rat.²⁵ The study of transgenic mice deficient for *IGF2* expression indicates that regulation of prenatal growth is the major function of IGF-2.²⁶ An overactive *IGF2* gene has been suggested²⁷⁻³⁰ as a mechanism for the fetal overgrowth³¹ and predisposition for embryonal tumors³² of the Beckwith-Wiedemann syndrome (BWS). A striking correlation between cell types with an abundant *IGF2* expression during normal development and the cell type-specific pattern of hyperplasia and tumor formation of the BWS has been observed.³⁰ The combination of paraganglionic hyperplasia³¹ and an apparent predispo-

sition for extra-adrenal infant neuroblastoma/ ganglioneuroma^{33–37} in this syndrome may suggest an extra-adrenal chromaffin origin of these tumors. The existence of an abundant *IGF2* expression in these normal and neoplastic cell types would provide further support for this hypothesis. The purpose of this study was to evaluate the utility

of IGF2 expression as a lineage marker in neuroblastoma. During normal development of the sympathetic nervous system chromaffin cells with an extra-adrenal location displayed an abundant IGF2 expression in contrast both to their adrenal counterparts and to sympathetic neurons. In neuroblastoma, IGF2 expression was present primarily in extra-adrenal tumors, and IGF2-expressing tumor cells were morphologically and histochemically similar to paraganglia/ SIF cells. We submit that this evidence of extraadrenal chromaffin differentiation in neuroblastoma provides new insights into the marked clinical heterogeneity of this tumor. A paraganglionic/SIF cell lineage of a subset of neuroblastomas may thus explain differences in anatomical origin, age at diagnosis, endocrine activity, histology, and tendency for spontaneous regression and maturation.

Materials and Methods

Embryonic and Fetal Tissue Specimens

Tissue specimens from first- and second-trimester pregnancies were obtained at therapeutic terminations, early tubal pregnancies, and late spontaneous abortions with the permission of the local Medical Ethics Committee at Karolinska Hospital (93–216). Age was determined by ultrasound analysis, crown-torump length (18 to 220 mm), foot length, and anamnestic information. Developmental (postfertilization) age was used instead of gestational age. Eight conceptuses of the following ages were analyzed: 7.0, 7.5, 7.5, 10.5, 11.0, 13.0, 19.0, and 23 weeks. The first two and the last two specimens represented abnormal pregnancies but were accepted for evaluation of normal development as they appeared histologically normal, and given that results from analyses of these specimens were concordant with those from normal pregnancies.

Tumor Specimens

32 neuroblastomas, 1 ganglioneuroblastoma, and 2 ganglioneuromas were included in the study. They were chosen from a national study of children treated for these tumors in Sweden during a 7-year period (1986 to 1993). The international criteria for neuroblastoma diagnosis, staging, and response to treatment³ were used for clinical classification of the tumors. Most low-stage (stages 1 to 2b) tumor specimens were collected before chemotherapy. Surgical biopsies were usually not taken in high-stage disease (stages 3 and 4) before treatment, but two such biopsies (cases 41 and 110) and four core needle biopsies (cases 105, 108, 115, 149) taken before treatment were included in the material. Because of the relative scarcity of extra-adrenal tumors diagnosed after 2 years of age in the total national material, a disproportionately large number of these tumors were sought in this study to better evaluate this tumor subtype. The specimens examined represented the primary tumor with one exception, in which a biopsy from a skin metastasis appearing during chemotherapy was utilized (case 165). Follow-up times for survivors ranged from 20 months to 6.5 years, and for children with fatal disease from 2 weeks to 17 months.

Tissue Processing

Tissue specimens were fixed in buffered formalin and routinely processed to paraffin. $5-\mu$ thick sections were collected on silanized slides and stained with hematoxylin and eosin or used for *in situ* hybridizations and immunohistochemical stainings.

RNA Probes

A ³⁵S-labeled antisense riboprobe used for *in situ* hybridization analysis (see below) was made from a 680-bp Hinf1-Pst1 human *IGF2* cDNA insert, cloned into pGem-3 (SP6; ref. 38). The probe was transcribed from supercoiled plasmids, yielding a specific activity of approximately 250 Ci/mmol. A sense probe from the same plasmid, with a similar specific activity, was employed as negative control.

In situ Hybridization Analysis

Tissue sections were hybridized to riboprobes at 56 C overnight and washed stringently before RNAse treatment as described previously.³⁸ After application of NTBII (Eastman Kodak Co., Rochester, NY) photographic emulsion (diluted 1:1 in 2% glycerol in H_2O) and exposure for 3 to 7 days at 4 C, sections were counterstained with Mayer's hematoxylin and mounted.

Immunohistochemical Characterization

Antibodies with specificity for the following antigens were used in consecutive sections: neuron-specific enolase (NSE; polyclonal antibody, BioGenex Laboratories, San Ramon, CA), chromogranins A and B (monoclonal antibody, BioGenex Laboratories; polyclonal antibody, Milab, Malmö, Sweden) and Ki 67 (monoclonal Mib 1 antibody, Immunotech S.A., Marseille, France). These antibodies were chosen as markers for neuronal and neuroendocrine differentiation and proliferation, respectively. Human leukocyte common antigen (monoclonal CD45 antibody, Dakopatts, Glostrup, Denmark) was used for detection of retroperitoneal lymphatic cells during normal development. The avidin-biotin-peroxidase complex method with diaminobenzidine as chromogen was used according to the manufacturer's manual (ABC kit, Dakopatts).

Results

Morphological and Histochemical Characterization of Human Sympathetic Nervous System Development

Sympathetic Development from Week 8 to Week 23

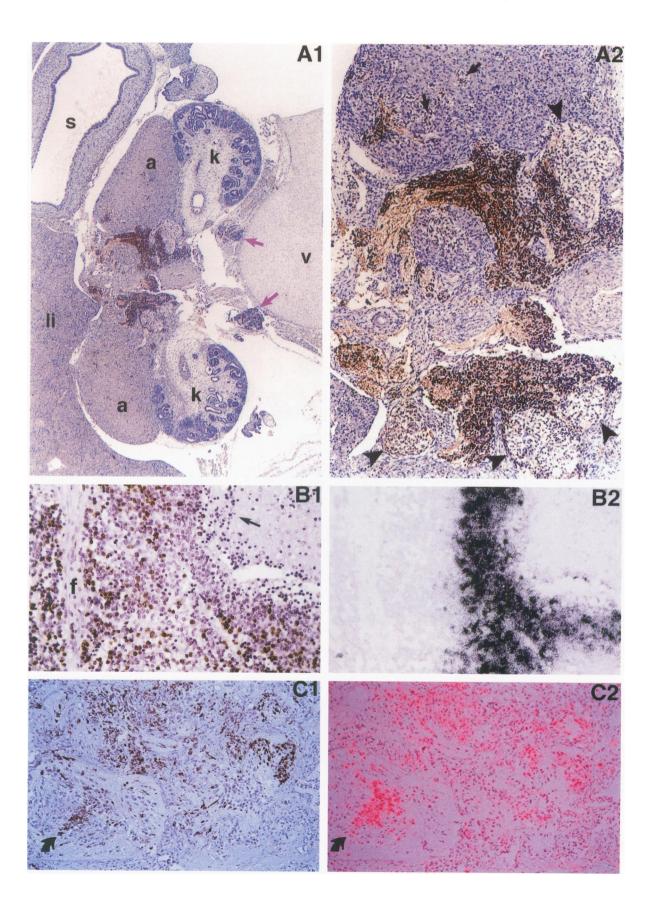
From week 8 of development, sympathetic cells were abundant in the retroperitoneal area, particularly at the adrenal level (Figure 2A). Chromaffin differentiation, represented by paraganglia, adrenal chromaffin cells (Figure 2A2), and SIF cells in the sympathetic trunk (Figure 3, A to C) was present already in the earliest specimens (8th week), whereas sympathetic neurons were present but morphologically primitive (Figures 2A2 and 3, C and D). During the period studied, paraganglia were of considerable size and particularly abundant in the juxta-adrenal area (Figure 2A), contrasting with relatively low numbers of adrenal chromaffin cells (Figure 2A2).

IGF2 Expression

Sympathetic neuronal cells did not express IGF2 at any stage of development investigated (Figure 3, A, E, and G). Paraganglia, however, expressed the gene abundantly throughout development (Figure 3, A and G). Type II SIF cells (Figure 3, A and E), were also strongly IGF2-positive. A centralized location of such SIF cells within the sympathetic trunk was a consistent finding (Figure 3A). These cells were particularly prominent in the earliest specimens but were also present at later stages. Type I SIF cells appearing as solitary cells within sympathetic ganglia were identified from the 19th week onward. It was possible to demonstrate with reasonable certainty that these cells also expressed IGF2 abundantly by superimposing photographs of consecutive sections utilized for IGF2 in situ hybridization and chromogranin immunohistochemistry (cf Figure 3, G and H).

Two types of adrenal medullary cells were seen. One type consisted of nests of primitive sympathetic cells, located mostly in the center of the gland with no detectable *IGF2* expression (Figure 3I). The other type consisted of chromaffin cells interspersed with fetal cortical cells, either on the medial side of the gland during early stages (until the 11th week; Figure 2A), or with a more central position during later stages (not shown). Whether *IGF2* expression in adrenal chromaffin cells was low or nonexistent could not be

Figure 2. Immunobistochemical characterization of embryonic sympathetic development and tumor specimens. (A) Early sympathetic development in the retroperitoneal area. Abdominal cross-section at the adrenal level of a 7.5-week embryo. NSE immunoreactive (brown) structures represent sympathetic cells and peripheral nerves. Cbromaffin differentiation is represented by paraganglia (arrowheads, A2) and scattered adrenal cells (arrows, A2). Sympathetic neuronal cells are abundant but morphologically primitive (A2). Other symbols: a. adrenal: k. kidney; li, liver; s. stomach; v. vertebra; colored arrow, sympathetic trunk. Magnification: A1, 30×; A2, 75×. **2B**: Zones of proliferating, IGF2-expressing, and apoptosis-like cells in a neuroblastoma tumor lobule. Consecutive sections from a neuroblastoma of a newborn child with particularly abundant IGF2 expressing cald apoptosis-like bodies toward the center of the lobule (arrow; same area shown in bigb-power view in Figure 4H), neigbboring the zone of IGF2-expressing cells, illustrated by panel B2 (brightfield view). Note the slightly brighter nuclei of IGF2-expressing the zone of chromogranin A+B and IGF2 in neuroblastoma. Consecutive sections from a malignant neuroblastoma of thoracic origin diagnosed at 3 years of age, causing severe hypertension (case 165, Table 1). (C1) shows chromogranin A+B and IGF2 in neuroblastoma. Consecutive sections from a malignant neuroblastoma of thoracic origin diagnosed at 3 years of age, causing severe hypertension (62×. All sections were counterstained with bematoxylin.



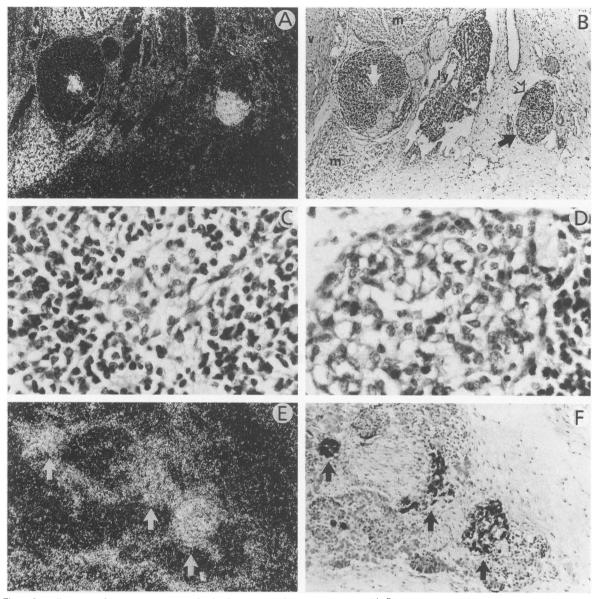
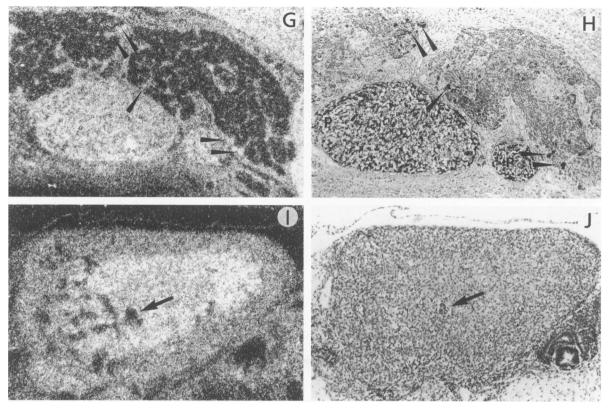


Figure 3. Cell type-specific IGF2 expression in the developing sympathetic nervous system: (A, B) IGF2 is expressed in paraganglia and SIF type II cells of the sympathetic trunk, but not in sympathetic neuronal cells. Abdominal cross section of a 10.5-week fetus showing (A) IGF2 in situ hybridization (darkfield view) and (B) bematoxylin and eosin staining of a consecutive section. Symbols: ly, lymphoid tissue; m, muscle; black solid arrow, paraganglion; open arrow, sympathetic ganglion; white arrow, SIF cells in the center of the sympathetic runk. Magnification, $33 \times .$ (C, D) Nuclear morphology of SIF cells (C) and paraganglia cells (D) compared with neighboring primitive sympathetic ganglion (19.5 weeks): Consecutive sections showing IGF2 expression in SIF type II cells of a juxta-adrenal sympathetic ganglion (19.5 weeks): Consecutive sections showing IGF2 expression (E, darkfield view) and chromogranin A+B immunoreactivity in bematoxylin counterstain (F). Arrows indicate SIF cells. Magnification, $63 \times$.

distinguished conclusively because of interfering signals from the strongly *IGF2*-positive cells of the fetal adrenal cortex (Figure 3I).

Immunohistochemical Characterization: Chromogranin A+B

The neuroendocrine nature of SIF cells and paraganglia was verified by intense chromogranin immunoreactivity (Figure 3, F and H). Adrenal chromogranin immunoreactivity was intense in the chromaffin cell type, whereas the nests of primitive sympathetic cells stained either negatively or weakly positively in areas of neuropil formation (not shown). Primitive sympathetic neuronal cells of sympathetic ganglia also stained negatively, with the exception of neuropil in the later stages (from 19 weeks; not shown). Morphologically more mature sympathetic neurons



(G,) IGF2 expression in type I SIF cells. Other area of same sections as shown in (E) and (F). Type I SIF cells stain intensely for chromogranins (arrowheads, panel H). IGF2 expression of the same areas are indicated in panel G (darkfield view). P, paraganglion. Magnification, $33 \times .$ (I, J). Adrenal IGF2 expression. An adrenal gland of a 7.5-week embryo in a cross-abdominal section showing intense expression in cortical cells and low or nonexistent expression in chromaffin cells (mostly located on the medial/left side) and in a nest of primitive sympathetic cells (arrow). (I) Darkfield view. (J) Consecutive section stained with bematoxylin and eosin. (For further morphological information see Figure 2A2 showing NSE immuno-reactivity of chromaffin cells in an adjacent section). Magnification, $40 \times .$

with increasing cytoplasm, nuclear enlargement, and distinct nucleoli were seen from the 19th week. These cells displayed weak chromogranin immunoreactivity (Figure 3F).

Other Markers

The neuronal character of retroperitoneal and adrenal sympathetic cells was confirmed by NSE immunoreactivity (Figure 2A). Cells of the lymphoid system were frequently identified in the retroperitoneal area at all stages of development (Figure 3B), but could be distinguished from the morphologically similar primitive sympathetic cells by a positive CD45 immunohistochemical reaction (not shown). Proliferation of primitive sympathetic neuronal, paraganglionic, and type II SIF cells was evident by a high cellular density of Ki 67 immunoreactivity (not shown).

IGF2 Expression in Neuroblastoma

Moderate *IGF2* expression of fibrovascular septa was identified in all specimens, but only those containing

IGF2-expressing neuroblastic cells will subsequently be referred to as positive tumors. The data are summarized in Table 1.

Extra-Adrenal Tumors

Two distinct clinical forms were seen. All 13 tumors diagnosed before 2.5 years were locally growing and, with one exception, prognostically favorable, whereas most (4/5) older children presented with advanced tumor growth at diagnosis and died from tumor progression. One newborn child died from cardiogenic shock and hepatic thrombosis, presumably because of massive tumor secretion of catecholamines (case 157, Table 1). One 3.5-year old child with multiple skeletal tumor foci is in complete long-term remission (case 111, Table 1). IGF2 expression was a frequent finding in both of these age groups (9/13 and 3/5 respectively, Table 1). In most tumors characteristic ringlike patterns of IGF2 expression were found (Figure 4A), corresponding to zones of cells in tumor areas with a lobular pattern (Figure 4B). These zones had a central position within lobules and surrounded a core of cellular debris (cf.

| Age | Site | Case | Stage | Outcome* | IGF2 expression | | Apoptosis- like bodies |
|--------------------------------------|----------------------|------|----------------|------------------|--------------------|---|---------------------------|
| | | no. | | | | | |
| Extra-adrenal tumors (age-sequenced) | | | | | | | |
| 0 | Pararenal | 148 | 1 | NED | + | + | - |
| 0 | Pararenal | 157 | 3 | DOD [†] | + | + | + |
| 1 month | Thoracic | 85 | 3 | NED | - | - | - |
| 1 month | Pelvic | 118 | 1 | NED | + | - | - |
| 2 months | Pararenal | 84 | 1 | NED | + | + | - |
| 2 months | Pararenal | 124 | 1 | NED | + | + | + |
| 3 months | Thoracic | 51 | 2b | NED | + | + | + |
| 7 months | Pelvic | 128 | 3 | NED | + | + | + |
| 12 months | Thoracic | 146 | 3 | NED | - | _ | - |
| 12 months | Para-adrenal | 153 | 3 | NED | - | + | - |
| 13 months | Para-adrenal | 78 | 3 | NED | + | + | + |
| 18 months | Thoracic | 87 | 2a | NED | _ | _ | - |
| 2.5 years | Pelvic | 138 | 2b | NED | + | + | _ |
| 3 years | Thoracic | 165 | -~ | DOD | + | + | _ |
| 3.5 years | Abd, paravertebral | 111 | 4 [‡] | NED | _ | + | + |
| 4 years | Thoracic | 130 | .4 | DOD | _ | _ | _ |
| 5 years | Thoracic | 69 | 3 | DOD | + | + | + |
| 6.5 years | Thoracic | 41 | 4 | DOD | + | _ | _ |
| Adrenal tumors (age-sequenced) | meraele | | • | 202 | | | |
| 0 | Adrenal [§] | 39 | 4S | DOD | + | + | + |
| 0 | Adrenal | 115 | 4S | DOD | _ | _ | _ |
| 0 | Adrenal [§] | 125 | 4S | NED | + | + | + |
| 0 | Adrenal [§] | 103 | 1 | NED | _ | + | - |
| 10 months | Adrenal | 106 | 4 | NED | _ | _ | _ |
| 11 months | Adrenal | 108 | 4‡ | NED | (+)¶ | - | _ |
| 21 months | Adrenal | 135 | 3 | DOD | + | + | + |
| 3 years | Adrenal | 149 | 3 | DOD | _ | _ | _ |
| 3.5 years | Adrenal | 68 | 4 | DOD | _ | _ | _ |
| 3.5 years | Adrenal | 105 | 4 [‡] | NED | _ | _ | _ |
| 3.5 years | Adrenal | 155 | 4 | DOD | _ | _ | _ |
| 4.5 years | Adrenal | 110 | 4 [‡] | dead# | _ | + | + |
| 5 years | Adrenal | 32 | 4 | DOD | _ | · | <u>'</u> |
| 6 years | Adrenal | 127 | 2b | NED | - + | + | + |
| 8.5 years | | 126 | 20 | DOD | т | т | т |
| 11.5 years | Adrenal | 120 | 4 | 000 | - | - | - |
| Ganglioneuroma | Thorpoin | 70 | | NED | | | |
| 2.5 years | Thoracic | 73 | | | - | - | - |
| 11 years | Pararenal | 59 | | NED | - | - | - |

Table 1. IGF2 Expression in Neuroblastomas and Ganglioneuromas Diagnosed in Sweden 1986–1993 in Relation to Clinical Data and to Presence of a Lobular Histological Pattern and Apoptosis-Like Bodies

Follow-up times for survivors range from 20 months to 6.5 years.

*NED, No evidence of disease; DOD, dead of disease.

[†]Cardiogenic shock at birth.

[‡]Skeletal involvement.

§Histologically verified.

Hepatic involvement.

[¶]Few scattered cells.

*Died from septic shock 12 months after diagnosis, while in autopsy-verified complete remission.

Figures 4, A and B, and 2B). Within the lobules, tumor cells displayed a gradient of increasing morphological maturation toward the center (Figures 2B and 4, C to E). Determinants of increasing maturation were increasing nuclear size with diminished basophilia and increasing cytoplasm.5-8,39 IGF2-positive cells were therefore the most mature of the lobule, typically displaying a more abundant cytoplasm and larger and brighter nuclei with punctate basophilia (Figure 4E). Usually the morphological differences between IGF2-positive and IGF2-negative tumor cells were subtle (eg, Figure 2B). The nuclear morphology of IGF2-expressing neuroblastoma cells was reminiscent of the normal fetal paraganglia and SIF cells (cf. Figures 3, C and D, and 4E). However, clear evidence of ganglionic differentiation with abundant neuropil formation, more prominent nuclear enlargement, and a distinct nucleolus was found in two *IGF2*-positive tumors with a lobular pattern (exemplified in Figure 4, F and G). Both tumors were diagnosed after infancy. The density of *IGF2*-expressing tumor cells varied considerably, both between specimens and between different areas within a specimen, depending on the extent of lobule formation. *IGF2*-negative tumors were less mature, usually with condensed nuclei (not shown) and lack of a lobular histological pattern (Table 1).

Adrenal Tumors

Three of the six infant tumors clinically assigned to an adrenal origin were stage 4S, with massive hepatic tumor growth causing death from respiratory problems in two of the children (Table 1). The other four infants remain alive despite distant tumor growth in three cases. Two of the 4S tumors expressed *IGF2* in paraganglion/SIF cell-like maturing cells within lobular pattern tumor areas, as described above. Intraadrenal tumor growth was histologically verified in both. The remaining infant tumors were morphologically immature, lacking the lobular pattern, and did not express *IGF2*, with the exception of one case (case 108, Table 1) where scattered cells were positive.

In contrast, most (6/9) children diagnosed with adrenal tumors after infancy died from tumor progression. Only two tumors of this older age group expressed *IGF2* (Table 1). One was diagnosed shortly after infancy (case 135), and the other was a prognostically favorable stage 2 tumor (case 127). Both of these tumors contained the lobular pattern characteristic of most extra-adrenal neuroblastomas (Table 1). One ganglioneuroblastoma, defined as a mixture of neuroblastic elements with varying degrees of ganglionic maturation and Schwann cells,^{7,39} was tested (case 155). This tumor neither expressed *IGF2* nor displayed a lobular arrangement of the maturing cells (Table 1).

IGF2 Expression in Ganglioneuroma

Ganglioneuroma is a benign tumor, histologically resembling the mature features of ganglioneuroblastoma.^{7,39} No *IGF2* expression was identified in two such tumors (Table 1).

Immunohistochemical Characterization of Neuroendocrine Differentiation and Proliferation in Neuroblastoma

Sections consecutive to those analyzed for IGF2 expression were used for immunohistochemical analyses. IGF2-expressing cells were readily identifiable in these sections by their increased size, central location in tumor lobules, and their nuclear morphology (Figure 2, B and C). Chromogranin immunoreactivity was principally specific to IGF2-expressing cells, although present with varying intensity (Figure 2C), whereas IGF2-negative tumor cells were also chromogranin-negative. Chromogranin-positivity was also seen in the central areas of tumor lobules associated with cellular debris (not shown). A more detailed inspection revealed that this part of the tumor lobule consists of apoptosis-like cellular remnants. Morphological evidences of apoptosis were present in a zone of cells with small, condensed, and occasionally fragmented nuclei adjacent to the zone of *IGF2*-expressing cells (Figures 2B and 4H). The DNA of these cells was fragmented as determined by the TUNEL terminal DNA nick end labeling technique (J. C. Hoehner, F. Hedborg, H. Jernberg-Wiklund, H. L. Olsen, S. Påhlman, Int J Cancer, in press). Invariably, these apoptosis-like bodies did not express *IGF2* (Figure 2B). The bulk of the cellular debris area consisted, however, of cell remnants devoid of nuclei (Figures 2B and 4H). Foci of apoptosis-like bodies were only observed in tumors with a lobular pattern, most of which expressed *IGF2* (10/12; Table 1).

The spatial distribution of proliferating tumor cells was determined immunohistochemically by detection of the cell cycle-specific Ki 67 protein. A particularly large proportion of tumor cells were positive in aggressive tumors. Ki 67 immunoreactivity was most prominent among tumor cells with fibrovascular stroma contact, whereas cellular zones with *IGF2* expression had a low fraction of Ki 67-immunoreactive cells (Figure 2B). Hence, *IGF2*-expressing cells do not appear to be proliferative to any great extent.

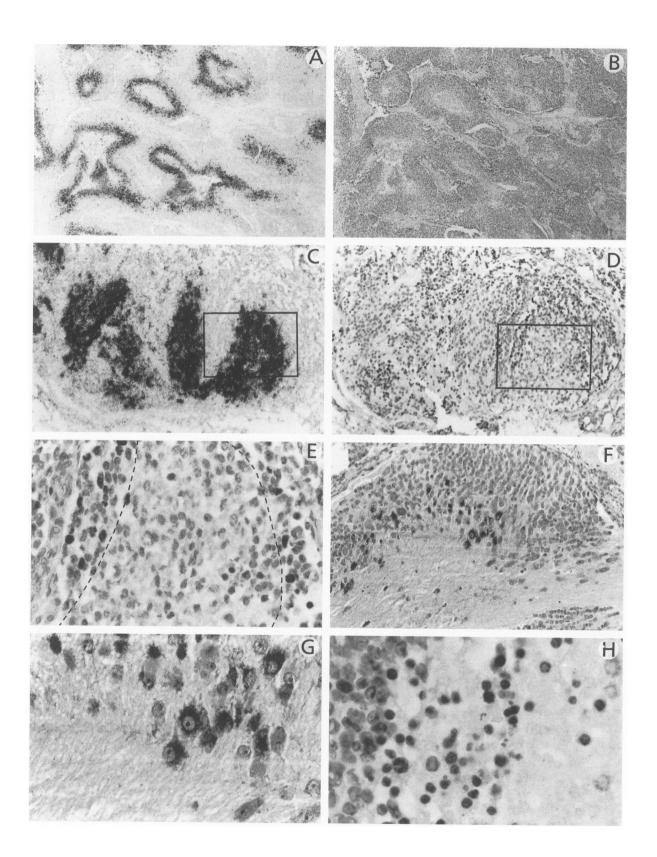
Correlation Between IGF2 Expression in Tumor Tissue and Clinical Signs of Endocrine Tumor Activity

Severe cardiovascular symptoms, a relatively rare occurrence with neuroblastoma,⁴⁰ were found in five cases, three of them diagnosed during infancy (cases 78, 118, and 157; Table 1) and the other two at 21 months and 3 years of age (cases 135 and 165). These tumors were particularly rich in *IGF2*expressing cells. Four of these children required treatment for hypertension. The fifth tumor was congenital; the child suffering unexpected asphyxia at birth (case 157; Table 1) died a few days later with a clinical picture of cardiogenic shock in combination with hepatic vascular thrombosis. The content of *IGF2*-expressing cells in this tumor was the highest found (Figure 4A).

Discussion

Specificity of IGF2 Expression in the Sympathetic Nervous System during Development

Whereas *IGF2* expression was absent in sympathetic neuronal cells at all stages of differentiation, paraganglia and SIF cells did express this gene at high levels. In contrast, *IGF2* expression in chromaffin cells of the



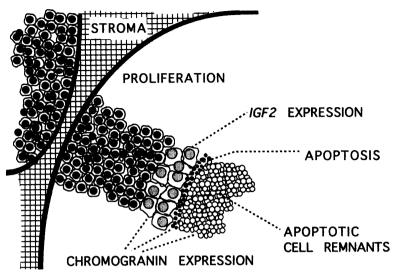


Figure 5. Schematic representation of functional zones in an IGF2-expressing neuroblastoma lobule (cf. Figure 2B).

adrenal medulla was negligible, as previously reported.⁴¹ Another type of adrenal medullary cell, primitive sympathetic cells, was present in small nests and showed no detectable expression. In the developing sympathetic nervous system high-level *IGF2* expression is therefore specific for extra-adrenal neuroendocrine cells.

Are There Two Separate Lineages in Neuroblastoma?

Observations such as spontaneous maturation of neuroblastoma into benign ganglioneuroma, ^{42–45} the neuronal morphology of ganglioneuroblastomas,^{7,39} neurite formation,^{46,47} and other neuronal characteristics of neuroblastoma and derived cell lines^{46–48} are fundamental to the traditional view that differentiation in neuroblastoma continues along a neuronal lineage. The results of this investigation reveal, however, that a subset of neuroblastomas express *IGF2* as a putative sign of neuroendocrine differentiation. This feature of differentiation in neuroblastoma provides a new perspective on its pathology. The two most consistent morphological signs linked to *IGF2* expression in neuroblastoma were a lobular pattern of cellular growth and moderately enlarged and pale nuclei with a punctate pattern of basophilia. When using the terminology suggested by Joshi et al^{7,39} such tumors would be classified as "poorly differentiated" and "differentiating" neuroblastomas. However, a greater emphasis was placed on ganglion cell-like nuclear features as determinants for differentiation in this classification system. We therefore believe that this and other systems for describing cellular maturation in neuroblastoma (reviewed in ref. 7) are confusing and would benefit considerably by introducing a distinction between neuronal and neuroendocrine phenotypes and that *IGF2* expression is a useful tool in this respect.

The prenatal abundance of paraganglia and SIF cells, contrasting with their postnatal decline, suggests an important developmental role of these cells. In view of the poorly developed adrenal medulla during fetal life, it is likely that the fetal sympathetic endocrine functions are exerted predominantly by the extra-adrenal chromaffin tissue. It has been shown that paraganglia are the major source of catecholamines during fetal asphyxia.^{17,18} If neoplasias were to develop from these prenatally active cell types, one might predict features such as a congenital or infantile tumor appearance in extra-adrenal locations, a high endocrine activity with a lack of

Figure 4. Cell type-specific IGF2 expression in neuroblastoma: (A, B) IGF2 is expressed in characteristic ringlike patterns in tumors with a lobular bistology. Consecutive sections of a congenital pararenal neuroblastoma (case 157; see also Figure 2B). (A) IGF2 expression (brightfield view) (B) Ki 67 immunobistochemistry; here used for illustration of the lobular bistological pattern. Magnification, $12 \times (C-E)$ IGF2 expression correlates to areas with morphological maturation. Consecutive sections from a prognostically favorable pelvic infant neuroblastoma (case 128; Table 1). (C) IGF2 expression (brightfield view; 58 \times). (D) Consecutive section stained with bematoxylin. (E) Higb-power magnification (180 \times) of boxed area in (D) showing nuclear morphology. Zone of IGF2-expressing cells is indicated (dashed lines). Note similarity in nuclear morphology compared to normal fetal SIF and paraganglia cells (cf. Figure 3, C and D). (F, G) Occasional IGF2-expressing tumors display ganglion cell-like features. A prognostically favorable pelvic neuroblastoma, diagnosed at 2.5 years (case 138) with a lobular pattern coexpressing chromogranin A+B(F) and IGF2 (not shown). Magnification, 58 \times . (G) Higb-power magnification (180 \times) of (F), showing nuclear morphology and neuropil. (H) Morphological evidence of apoptosis in the central area of an IGF2 positive tumor lobule. Nuclear condensation and fragmentation and cell remnants without nuclei are seen adjacent to a zone of IGF2-expressing tumor cells. Higb-power view (360 \times) of area indicated in Figure 2B1. All specimens are counterstained with behavior.

adrenalin in the profile of secreted catecholamines, a tendency for spontaneous postnatal regression, and histological features resembling paraganglia/SIF cells. Because prognostically favorable and stage 4S infant neuroblastomas meet all these criteria we suggest that these tumors, as well as extra-adrenal tumors of older children, constitute a subgroup of neuroblastomas that may be viewed as embryonal paraganglia/SIF-omas. However, these neuroendocrine neuroblastomas differ phenotypically from adult paragangliomas in that neuroblastomas express the neuronal c-src splice variant c-srcNI48 and bcl-2 (J. C. Hoehner, F. Hedbora, H. Wiklund-Jernbera, L. Olsen, Påhlman S; Int J Cancer, in press), while adult paragangliomas do not (ref. 48; J. C. Hoehner, unpublished data). Thus prognostically favorable infant and extra-adrenal neuroblastomas do show neuronal characteristics, suggesting a derivation from a progenitor cell with a mixed neuroendocrine and neuronal phenotype. This progenitor cell may have the capacity to mature either along a neuronal or a neuroendocrine lineage, as also discussed below.

Based on the bipotential properties of paraganglia and SIF cells, which both have a capacity for in vitro and in vivo transdifferentiation into a neuronal phenotype when exposed to nerve growth factor (NGF),⁴⁹⁻⁵¹ one could infer that low-malignant neuroblastomas, which are shown to express high transcript levels of c-trk, the high-affinity NGF receptor gene,^{52,53} may have the same capacity for transdifferentiation from a paraganglionic/SIF cell phenotype into tumors with a ganglionic morphology. This speculation is consistent with the clinical types of neuroblastoma reported to undergo spontaneous maturation into ganglioneuroma. Goldman et al43 reviewed eight cases of maturing neuroblastomas, all of which were extra-adrenally located and seven of which were diagnosed before 2 years of age. Spontaneous maturation into ganglioneuroma is also reported in 4S tumors (reviewed in refs. 43 and 44), here shown to be of the same neuroendocrine phenotype as other prognostically favorable infant tumors. Observations in ganglioneuroma, such as its typically extra-adrenal location,⁴⁵ the relatively late age at diagnosis,⁴⁵ and its association to the BWS^{33,35} also favor the hypothesis that this benign tumor represents a maturational remnant from infant tumors. Interestingly, the small number of IGF2-expressing tumors with a ganglion cell-like morphology found in this investigation were diagnosed after infancy, consistent with an agedependent conversion of phenotype. However, despite the ganglionic features of these tumors, they would rather be classified as "differentiating neuroblastomas" than "ganglioneuroblastomas" when applying the terminology suggested by Joshi et al,^{7,38} since no Schwann cell component was seen.

Do Some Neuroblastomas, Clinically Assigned to an Adrenal Origin, in Fact Originate from the Juxta-Adrenal Sympathetic Tissue?

It is likely that the abundant juxta-adrenal sympathetic tissue during development would be a significant contributor to the same types of neuroblastomas as found in other extra-adrenal parts of the sympathetic nervous system. Such a tumor origin may, however, be difficult to distinguish clinically from a true adrenal medullary origin. This may explain why suprarenal growth is not an infrequent finding in low-stage infant tumors. A true adrenal medullary growth for 4S tumors was histologically confirmed in this study despite the presence of the extra-adrenal chromaffin differentiation marker IGF2 expression. This apparent paradox would be reconciled by assuming that the adrenal siting of 4S tumors, instead of being the true site of origin, is the result of organ-specific metastatic spread, which is a prominent feature of this tumor form.

Clinical Implications

IGF2 expression is not a reliable predictor of a favorable outcome; however, it is associated with prognostically favorable tumors. It may rather provide improved biological characterization of an individual tumor, eg, in evaluation of endocrine activity. It is conceivable that the site-dependent phenotypical differences in aggressive neuroblastomas of older children, as indicated by their differences in *IGF2* expression, may have therapeutic implications. For example, targeted therapy with radiolabeled catecholamine metabolites could be more efficacious in extra-adrenal tumors because of the higher content of catecholamine-storing granular vesicles in tumors with a neuroendocrine phenotype.

The Tumor Lobule: a Clue to the Function of IGF2 in Neuroblastoma?

Analogous to the organoid "*Zellballen*" pattern of adult paragangliomas,¹³ most *IGF2*-expressing neuroblastomas were organized into lobules surrounded by thin fibrovascular septa. Distinct functional zones of proliferation, cell maturation, *IGF2* expression, and cell death could be defined in these lobules (Figure 5). Somewhat surprisingly, in view of the putative role

of IGF-2 as an autocrine growth factor in neuroblastoma cell lines,^{21–23} IGF2-expressing cells were nonproliferative. As IGF2-expressing cells were the most mature, a differentiation-promoting role of IGF-2 may be inferred instead, consistent with other cell line data.²² Alternatively, IGF2 expression could be the effect, rather than the cause, of cell maturation, as has been shown with retinoic acid-induced in vitro differentiation.54 Furthermore, as IGF2 expression defined a border to apoptosis-like cells, a role for this growth factor in the process of programmed cell death may be implicated. Nevertheless, the presence of proliferation in the periphery of tumor lobules followed by maturation and subsequent apoptotic cell death toward the center may be pertinent clues to the unique pattern of growth, maturation, and spontaneous regression that is associated with these tumors.

Acknowledgments

We are deeply grateful for valuable advice from Dr Per Uoquer, and for skillful technical assistance from Ulf Hörnberg, Kenneth Wester, Maj-Lis Book and Frank Bittkowski.

References

- Berthold F: Overview: biology of neuroblastoma. Neuroblastoma: Tumour Biology and Therapy. Edited by C Pochedly. Boca Raton, CRC Press, 1990, pp 1–27
- Evans A, D'Angio G, Propert K, Anderson J, Hann H-W: Prognostic factors in neuroblastoma. Cancer 1987, 59:1853–1859
- Brodeur G, Pritchard J, Berthold F, Carlsen N, Castel V, Castleberry R, DeBernardi B, Evans A, Favrot M, Hedborg F, Kaneko M, Kemshead J, Lampert F, Lee R, Look T, Pearson A, Philip T, Roald B, Sawada T, Seeger R, Tsuchida Y, Voute P: Revisions of the international criteria for neuroblastoma diagnosis, staging, and response to treatment. J Clin Oncol 1993, 11: 1466–1477
- Carlsen N, Christensen I, Schroeder H, Bro P, Erichsen G, Hamborg-Pedersen B, Jensen K, Nielsen O: Prognostic factors in neuroblastomas treated in Denmark from 1943–1980. Cancer 1986, 58:2726–2735
- Shimada H, Chatten J, Newton W, Sachs N, Hamoudi A, Chiba T, Marsden H, Misugi K: Histopathologic prognostic factors in neuroblastic tumors: Definition of subtypes of ganglioneuroblastoma and an age-linked classification of neuroblastomas. J Natl Cancer Inst 1984, 73:405–416
- Beckwith JB, Martin R: Observation on the histopathology of neuroblastomas. J Pediatr Surg 1968, 3:106– 110
- 7. Joshi VV, Cantor AB, Altshuler G, Larkin EW, Neill JSA,

Shuster JJ, Holbrook CT, Hayes FA, Castleberry RP: Recommendations for modification of terminology of neuroblastic tumors and prognostic significance of Shimada classification. Cancer 1992, 69:2183–2196

- Sandstedt B, Jereb B, Eklund G: Prognostic factors in neuroblastomas. Acta Path Microbiol Immunol Scand 1983, 91(A):365–371
- Hedborg F, Bjelfman C, Sparén P, Sandstedt B, Påhlman S: Biochemical evidences for mature phenotypes in morphologically poorly differentiated neuroblastomas with a favourable outcome. Eur J Cancer (in press)
- 10. Everson T, Cole W: Spontaneous Regression of Cancer. Philadelphia, WB Saunders Co, 1966, pp 11–87
- Matthay K, Sather H, Seeger R, Haase G, Hammond D: Excellent outcome of stage II neuroblastoma is independent of residual disease and radiation therapy. J Clin Oncol 1989, 7:236–244
- Beckwith JB, Perrin E: *In situ* neuroblastomas: a contribution to the natural history of neural crest tumours. Am J Pathol 1962, 43:1089–1104
- Tischler A: The adrenal medulla and extra-adrenal paraganglia. Functional Endocrine Pathology. Edited by K Kovacs and S Asa. St Louis, Missouri, Blackwell Scientific Publications, 1991, pp 509–545
- 14. Coupland R: The Natural History of the Chromaffin Cell. London, Longmans Green, 1965
- Coupland R: The development and fate of catecholamine secreting endocrine cells. Biogenic Amines in Development. Edited by H Parves and S Parves. Amsterdam, Elsevier/North-Holland Biomedical Press, 1980, pp 3–28
- Taxi J, Derer M, Domich A-M: Morphology and histophysiology of SIF cells in the autonomic ganglia. Edited by L-G Elfvin. Chichester, UK, John Wiley & Sons Ltd, 1983, pp 67–95
- Hervonen A, Korkala O: The effect of hypoxia on the catecholamine content of human fetal abdominal paraganglia and adrenal medulla. Acta Obstet Gynec Scand 1972, 51:17–24
- Brundin T: Studies on the preaortal paraganglia of newborn rabbits. Acta Physiol Scand 1966, 70 suppl 290:1–54
- Lempinen M: Extra-adrenal chromaffin tissue and the effect of cortical hormones on it. Acta Physiol Scand 1964, 62 suppl 231:1–87
- 20. Winkler H, Fishcer-Colbrie R: The chromogranins A and B: the first 25 years and future perspectives. Neuroscience 1992, 49:497–528
- El-Badry O, Romanus J, Helman L, Cooper M, Rechler M, Israel M: Autonomous growth of a human neuroblastoma cell line is mediated by insulin-like growth factor II. J Clin Invest 1989, 84:829–839
- Påhlman S, Meyerson G, Lindgren E, Schalling M, Johansson I: Insulin-like growth factor I shifts from promoting cell division to potentiating maturation during neuronal differentiation. Proc Natl Acad Sci USA 1991, 88:9994–9998

- Martin D, Feldman E: Regulation of insulin-like growth factor II expression and its role in autocrine growth of human neuroblastoma cells. J Cell Physiol 1993, 155: 290–300
- Recio-Pinto E, Rechler M, Ishii D: Effects of insulin, insulin-like growth factor II and nerve growth factor on neurite formation and survival in cultured sympathetic and sensory neurons. J Neurosci 1986, 6:1211–1219
- Barres B, Hart I, Coles H, Burne J, Voyvodic J, Richardson W, Raff M: Cell death and control of cell survival in the oligodendrocyte lineage. Cell 1992, 70: 31–46
- DeChiara T, Efstratiadis A, Robertson E: A growthdeficiency phenotype in heterozygous mice carrying an insulin-like growth factor II gene disrupted by targeting. Nature 1990, 345:78–80
- Ohlsson R, Nyström A, Pfeiffer-Ohlsson S, Hedborg F, Schofield P, Flam F, Ekström T: *IGF2* is parentally imprinted during human embryogenesis and in the Beckwith-Wiedemann syndrome. Nature Genet 1993, 4:94–97
- Junien C: Beckwith-Wiedemann syndrome, tumorigenesis and imprinting. Curr Opin Genet Dev 1992, 2:431–438
- Weksberg R, Shen D, Fei Y, Song Q, Squire J: Disruption of insulin-like growth factor 2 imprinting in Beckwith-Wiedemann syndrome. Nature Genet 1993, 5:143–149
- Hedborg F, Holmgren L, Sandstedt B, Ohlsson R: The cell type-specific *IGF2* expression during early human development correlates to the pattern of overgrowth and neoplasia in the Beckwith-Wiedemann syndrome. Am J Pathol 1994, 145:802–817
- Beckwith JB: Macroglossia, omphalocele, adrenal cytomegaly, gigantism, and hyperplastic visceromegaly. Birth Defects 1969, 5:188–196
- Wiedemann H-R: Tumours and hemihypertrophy associated with Wiedemann-Beckwith syndrome. Eur J Pediatr 1983, 141:129
- Wiedemann H-R: Complex malformatif familial avec hernie ombilical et macroglossie. Un syndrome noveau? J Genet Hum 1964, 13:223–232
- Emery L, Shields M, Shah N, Garbes A: Neuroblastoma Associated with Beckwith-Wiedemann Syndrome. Cancer 1983, 52:176–179
- Sirinelli D, Silberman B, Baudon J, Sinnassamy P, Gruner M, Montagne J: Beckwith-Wiedemann syndrome and neural crest tumors. A report of two cases. Pediatr Radiol 1989, 19:242–245
- Ping A, Reeve A, Law D, Young M, Boehnke M, Feinberg A: Genetic linkage of Beckwith-Wiedemann syndrome to 11p15. Am J Hum Genet 1989, 44:720–723
- Schneid H, Vazquez M, Seurin D, le Bouc Y: Loss of heterozygosity in non-tumoral tissue in two children with Beckwith-Wiedemann syndrome. Growth Regul 1991, 1:168–169
- Ohlsson R, Holmgren L, Glaser A, Szpecht A, Pfeifer-Ohlsson S: Insulin-like growth factor 2 and short-range

stimulatory loops in control of human placental growth. EMBO J 1989, 8:1993–1999

- 39. Joshi V, Silverman J, Altshuler G, Cantor A, Larkin E, Neill J, Norris H, Shuster J, Holbrook C, Hayes F, Smith I, Castleberry R: Systematization of primary histopathologic and fine-needle aspiration cytologic features and description of unusual histopathologic features of neuroblastic tumors: a report from the pediatric oncology group. Hum Pathol 1993, 24:493– 504
- Weinblatt M, Heisel M, Siegel S: Hypertension in children with neurogenic tumours. Pediatrics 1983, 71: 947–951
- El-Badry O, Helman L, Chatten J, Steinberg S, Evans A, Israel M: Insulin-like growth factor II-mediated proliferation of human neuroblastoma. J Clin Invest 1991, 87:648–657
- Cushing H, Wohlbach S: The transformation of a malignant paravertebral sympaticoblastoma into a benign ganglioneuroma. Am J Path 1927, 3:203–215
- 43. Goldman R, Winterling A, Winterling C: Maturation of tumors of the sympathetic nervous system. Report of long-term survival in 2 patients, one with disseminated osseous metastases, and a review of cases in the literature. Cancer 1965, 18:1510–1516
- Haas D, Ablin A, Miller C, Zoger S, Matthay K: Complete pathologic maturation and regression of stage IV-S neuroblastoma without treatment. Cancer 1988, 62:818–825
- 45. Hayes A, Green A, Rao B: Clinical manifestations of ganglioneuroma. Cancer 1989, 63:1211–1214
- Påhlman S, Odelstad L, Larsson E, Grotte G, Nilsson K: Phenotypic changes of human neuroblastoma cells in culture induced by 12-O-tetradecanoyl-phorbol-13acetate. 1981, 28:583–589
- Scott I, Åkerman K, Heikkilä J, Andersson L: Development of a neuronal phenotype in differentiating ganglion cell-derived human neuroblastoma cells. 1986, 128:285–292
- Örtoft E, Bjelfman C, Hedborg F, Grimelius L, Påhlman S: The expression profile of alternatively spliced neuronal c-*src* RNA distinguishes between human tumors of the sympathoadrenal lineage. Int J Cancer 1995, 60:38–44
- Anderson D, Axel R: A bipotential neuroendocrine precursor whose choice of cell fate is determined by NGF and glucocorticoids. Cell 1986, 47:1079– 1090
- Doupe J, Patterson H, Landis S: Small intensely fluorescent cells in culture: role of glucocorticoids and growth factors in their development and interconversions with other neural crest derivatives. J Neurosci 1985, 5:2143–2160
- Koistinaho J, Hatanpää K, Hervonen A: Human paraganglion cells differentiate into adrenergic neurons in culture. Exp Neurol 1990, 107:277–280
- 52. Nakagawara A, Arima-Nakagawara M, Scavarda N, Azar C, Cator A, Brodeur G: Association between high

levels of expression of the *trk* gene and favourable outcome in human neuroblastoma. N Eng J Med 1993, 328:847–854

53. Kogner P, Barbany G, Dominici C, Castello M, Raschellà G, Persson H: Coexpression of mRNA for *trk* protooncogene and low affinity nerve growth factor receptor in neuroblastoma with favourable prognosis. Cancer Res 1993, 53:2044–2050

54. Matsumoto K, Gaetano C, Daughaday W, Thiele C: Retinoic acid regulates insulin-like growth factor II expression in a neuroblastoma cell line. Endocrinology 1992, 130:3669–3676