### **SUPPLEMENTAL MATERIAL**

#### **Supplemental text**

#### **How to derive the relationship between** τ**o and** τ**o, obs ?**

If the pdf (probability density function) of the closed times is a single-exponential pdf represented by the equation:  $f(t)=(1/\tau_{c1})*\exp(-t/\tau_{c1})$ , the fraction of all closed events shorter than the filter dead time (t<sub>d</sub>) is  $P(t \le t_d) = 1-\exp(-t_d/\tau_{c1})$ . Since >95% of all closed events belongs to the first component, we can make the approximation  $P(t \le t_d) \approx 1$ -exp( $t_d/\tau_{c1}$ ) even for our 4-exponential closed-time distribution. This fraction of closed events will not be detected; we will only detect the fraction of events longer than the filter dead time, given by  $P(t>t_d) = \exp(-t_d/\tau_{c1})$ . Therefore, our apparent overall closing rate (koc<sub>obs</sub>) will be slower than the real overall closing rate koc<sub>real</sub>  $( =$  ko1+ko2). The relationship is  $koc<sub>obs</sub> = koc<sub>real</sub> * P(t>t<sub>d</sub>)$ , because koc is proportional to the number of closures and we can only detect a fraction  $P(\triangleright t_d)$  of all closures. Therefore, koc<sub>obs</sub> = koc<sub>real</sub>\*exp(-t<sub>d</sub>/ $\tau_{c1}$ ). Because  $\tau_{0.0bs} = 1/\text{koc}_{obs}$  and  $\tau_{0.0} = 1/\text{koc}_{real}$ , we can write  $\tau_{0.0} = \tau_{0.0bs} * \text{exp}(-t_d/\tau_{c1})$ . In our article τ<sub>o</sub> stands for τ<sub>o,real</sub>.

### **Figure legends**

### **Figure S1. Bar charts comparing various selected single-channel parameters for the SUR and TMD0 channels obtained at -40 mV.**

Statistical significance was calculated using an unpaired Student *t*-test and significant differences were found for the pairs of parameters marked by brackets ( $**$  for  $p<0.05$ ).

### **Figure S2. Bar charts comparing various selected single-channel parameters for the SUR and TMD0 channels obtained at -100 mV.**

Statistical significance was calculated using an unpaired Student *t*-test and significant differences were found for the pairs of parameters marked by brackets ( $**$  for  $p<0.05$ ).

#### **Figure S3. No detectable voltage dependence of ATP inhibition.**

A. Time course of a macroscopic current in response to different ATP concentrations at two membrane potentials. Current was recorded from a macropatch expressing SUR2A/Kir6.2∆26 in the inside-out configuraton. The same patch was exposed to a series of [ATP] *(bars)* both at -100 and at -40 mV. B. ATP dose response curves at -100 and -40 mV are superimposable. There is no difference in the IC50 or the Hill coefficients obtained at the two membrane potentials.

## **TABLE S1**



### **Number of SUR/Kir6.2**∆**26 recordings whose closed times were best fitted with 3-5 exponentials\***

 ${{^*{\sf n}}_{\sf i}}$  denotes the number of recordings that could be best fitted with i exponentials. The percentages of n<sub>i</sub> are in parentheses.

## **TABLE S2**



### **Single channel parameters for chimera/Kir6.2**∆**26 measured at -40 mV\***

 $^*$  τ<sub>ci</sub>, a<sub>ci</sub>, τ<sub>b</sub>, τ<sub>ib</sub>, τ<sub>o</sub> and N are defined in the Methods section;  $\gamma$  – single-channel conductance; Po – open probability; n – number of recordings used for the analyses; values are given in mean  $\pm$  SEM



**Rate constants for studied channels (+ Kir6.2**∆**26)**\*

\* rate constants obtained at -40 mV are bolded and italicized; rate constants obtained at -100 mV are in parentheses; values were given in mean  $\pm$  SEM

# **FIGURE S1**











### **FIGURE S2**

**0**

**20**



**0**

**0**

**2**

**0.000.01**

**FIGURE S3**



