

A study of the histamine H₂-receptor mediating relaxation of the parenchymal lung strip preparation of the guinea-pig

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- 1 The relaxation produced by several H₂-receptor agonists and forskolin was investigated on strips of guinea-pig lung parenchyma.
- 2 Dimaprit, 1 μM to 10 mM, 4-methyl histamine, 0.5 μM to 100 μM and impromidine, 10 nM to 1 μM, had no effect on the tone of the unstimulated strips of lung parenchyma but caused a dose-dependant relaxation of strips that were contracted by 2-pyridylethylamine (2-PEA), 15 μM. Forskolin, 10 nM to 4 μM, produced a dose-dependent relaxation of both the stimulated and unstimulated lung strips.
- 3 The muscarinic antagonist atropine, 1 μM, and the β₂-adrenoceptor antagonist propranolol, 10 μM, had no effect on the dose-response curve for dimaprit-induced relaxation of the lung strip.
- 4 The dose-response curve for dimaprit was shifted to the right in a dose-dependent manner by increasing concentrations of a variety of H₂-antagonists. Schild plots produced a straight line for all the H₂-antagonists with slopes not significantly different from unity. The equilibrium dissociation constants for the H₂-antagonists on the lung strip preparation were similar to those previously reported for inhibition of the chronotropic activity of histamine on guinea-pig right atria and inhibition of [³H]-tiotidine binding to homogenates of guinea-pig lung parenchyma.

Introduction

Histamine stimulates contraction of the smooth muscle of the guinea-pig ileum and the guinea-pig trachea and these effects are blocked by low concentrations of mepyramine, an H₁-receptor antagonist (Dews & Graham, 1946). However, there are some effects of histamine such as stimulation of acid secretion by the stomach, the positive chronotropic action on the heart and relaxation of the rat uterus, which cannot be antagonized by mepyramine. Ash & Schild (1955) suggested that these actions of histamine were mediated by a different receptor from those actions that are mepyramine-sensitive. The mepyramine-insensitive effects of histamine are antagonized by compounds such as metiamide (Black *et al.*, 1973), cimetidine (Brimblecombe *et al.*, 1975) and ranitidine (Bradshaw *et al.*, 1979), and the receptors interacting with these compounds have been defined as H₂-receptors (Black *et al.*, 1972).

In the lung, histamine contracts smooth muscle *in vitro* and *in vivo* by stimulation of H₁-receptors (Chand & Eyre, 1975). However, histamine stimulates H₂-receptors mediating relaxation of tracheobronchial

smooth muscle from a variety of different species such as guinea-pigs (Chand & Deroth, 1979), rhesus monkeys (Chand *et al.*, 1980) and man (Vincenc *et al.*, 1984). Also, antagonists acting at H₂-receptors enhance the response of guinea-pig bronchial smooth muscle to histamine (Eyre, 1977; Dunlop & Smith, 1977; Okpako *et al.*, 1978; Drazen *et al.*, 1980).

Dimaprit, a selective H₂-agonist (Parsons *et al.*, 1977) has been shown to cause reversal of pre-existing anaphylactic contractions of guinea-pig lung parenchymal strips (Chand, 1979) and there is also evidence that, in the guinea-pig, dimaprit has a bronchodilator activity *in vivo* (Drazen *et al.*, 1979; Bottomley *et al.*, 1984). Both these effects were blocked by the H₂-antagonist metiamide (Black *et al.*, 1973). Dimaprit is, therefore, a useful tool to use when studying the effect of histamine H₂-receptors in guinea-pig airways.

In disease states such as asthma, where bronchial hyperreactivity to histamine is one of the characteristic features, the site of action of immunologically released pharmacological mediators such as histamine is mainly in the terminal airways (Drazen & Austen, 1974)

rather than the proximal airways. We have, therefore, used the lung parenchymal strip (Lulich *et al.*, 1976) to study the effect of dimaprit on smooth muscle in the small airways of the lung and characterize the H₂-receptor in lung tissue. The lung strip is known to possess several different contractile elements, the most important being airway smooth muscle and vascular smooth muscle (Clayton *et al.*, 1981; Bertram *et al.*, 1983). Although we believe that 2-pyridylethylamine (2-PEA), a specific H₁-agonist (Durant *et al.*, 1975), and histamine contract only airway smooth muscle, the role of other contractile elements, particularly vascular smooth muscle, cannot be excluded.

Methods

Hartley guinea-pigs of either sex aged 4 to 6 months and weighing 400–600 g were killed by stunning and exsanguination by sectioning of the carotid arteries. The heart and lungs were removed together and placed in Tyrode solution maintained at 37°C. Strips of lung approximately 15 mm × 3 mm × 3 mm were cut from the periphery of each lobe essentially as described previously (Drazen & Schneider, 1978) and mounted under a tension of 0.5 g weight in 2 ml organ baths containing Tyrode solution maintained at 37°C. The strips were gassed continuously with a mixture of 95% O₂ and 5% CO₂. The strips were allowed to rest for 1 h and changes of bathing medium were made every 10 min. During this time period all the strips relaxed until a new steady resting level of tone was produced. All changes in tissue length were recorded isotonically by means of photoelectric transducers connected through a pre-amplifier to Linseis L600 pen recorders. The transducer was calibrated so that actual displacement in mm resulting from tissue contraction could be related to the magnitude of the pen movement on the recorder.

The composition of the Tyrode solution was (mM): NaCl 137, KCl 2.7, CaCl 1.8, MgCl 1.0, NaHCO₃ 11.9, NaH₂PO₄ 0.4 and glucose 5.6

Addition of 2-pyridylethylamine

Addition of 2-PEA, 100 μM, produced 70 ± 5% of the maximum response to 2-PEA on the lung strip. After the tissue had been incubated with histamine, 100 μM, which produced a maximum contraction, for 20 min and then washed thoroughly, addition of 2-PEA, 100 μM, produced 73 ± 8% of the maximum response to 2-PEA. In all experiments, the tissue was initially contracted with a maximally effective concentration of histamine, 100 μM, and was then washed several times until the resting level of tone was restored. As stated, this did not affect subsequent responses to 2-PEA and was done because it is accepted that 'priming' a tissue

with a maximally effective concentration of agonist, in this case histamine, stabilizes the tissue to subsequent doses of the same or a different agonist. Such pretreatment with an agonist can also reduce the variability sometimes seen when dose-response curves are constructed with single doses given in random order rather than being given as progressively increasing doses or by cumulative addition. Dose-response curves to 2-PEA were constructed by giving single doses in randomized order. When the response for a particular dose reached a maximum the bath fluid was replaced and the preparation was allowed to return passively to its resting tension, which took between 10 and 20 min depending on the size of the responses. The dose-response curve was repeated after allowing the tissue to rest for 1 h. All the contractile responses obtained have been expressed as a percentage of the maximum contraction to 2-PEA for that particular strip.

Cumulative addition of an H₂-agonist or forskolin

Doses of an H₂-agonist or forskolin were added in a non-random, cumulative order starting with the lowest dose, either to lung parenchymal strips which had been precontracted with 2-PEA, 100 μM, or to unstimulated strips. 2-PEA at a concentration of 100 μM gave a response that was 70% of the maximum response to 2-PEA. This level of response was chosen to avoid working at the top of the dose-response curve but it gave an adequate level of contraction for study of the effect of drugs inducing relaxation. Cumulative addition of H₂-agonist or forskolin was used because the tissue was pre-contracted with 2-PEA and it was considered undesirable to disturb the tissue or extend the duration of the experiment by repeated changes of bath fluid. The next dose was added when no further relaxation to the first dose was observed and a new steady level of tone had been achieved: this took 10 to 15 min. Cumulative addition of H₂-agonist or forskolin continued until there was no further relaxation of the lung strip. All relaxation responses produced have been expressed as a percentage reduction of the contraction produced by 2-PEA, 100 μM, on that particular strip. The exception was for the relaxation of resting tone produced by forskolin, where the responses have been expressed as a percentage of the maximum relaxation produced by forskolin on that particular strip where 100% maximum relaxation is equivalent to a lengthening of the lung strip of 1.2 ± 0.3 mm (mean ± s.e.mean).

Antagonist experiments

In these experiments lung strips were divided into pairs. A control, cumulative dose-response curve to dimaprit was established on one strip and on the other

strip a cumulative dose-response curve to dimaprit was established in the presence of the concentration of the antagonist being investigated, according to the experimental protocol. After this, the strips were washed thoroughly and allowed to rest for 2 h during which time the resting level of tone was restored. The procedure described above was reversed on the two strips and the results pooled. In the experiments where an H₂-antagonist was present, the lung strips were equilibrated with the H₂-antagonist for between 30 and 45 min before the dose-response curve to dimaprit was determined. From binding studies (Foreman *et al.*, 1985) it is clear that this time period is sufficient for the H₂-antagonist to reach equilibrium with any H₂-receptors in the lung strip. All relaxation responses produced have been expressed as a percentage of the maximum control relaxation to dimaprit on a particular strip.

Evaluation of data

All EC₅₀ values were estimated from dose-response curves fitted to the data points by a computerised, non-linear, least squares estimate (Marquardt, 1963).

In experiments involving the histamine H₂-antagonists, the dose-response curves in the absence and presence of the antagonist were constrained to be parallel. The magnitude of the antagonism was calculated using the dose-ratio of Gaddum *et al.* (1955)

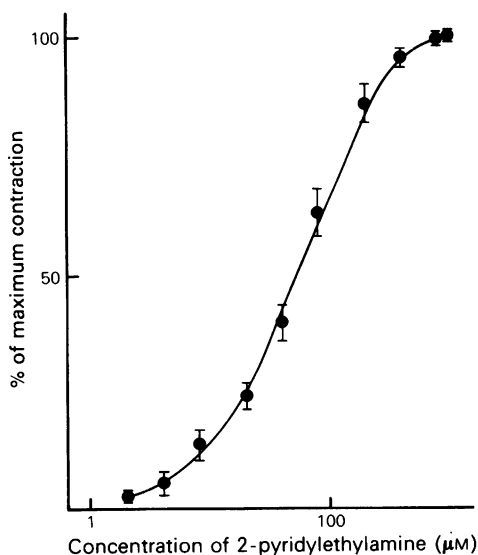


Figure 1 Dose-response curve showing the contraction of the guinea-pig lung strip induced by 2-pyridylethylamine (2-PEA). Responses have been expressed as a percentage of the maximum contraction to 2-PEA for a particular strip. Points are means of five determinations and the vertical bars represent s.e.mean.

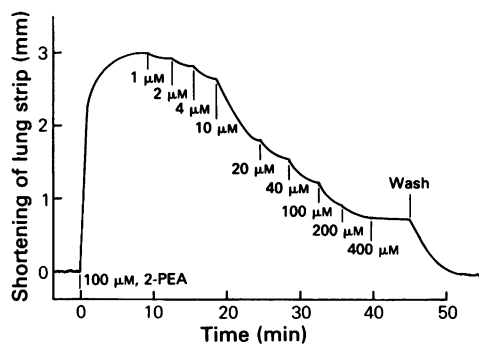


Figure 2 The effect of cumulative addition of dimaprit on the contraction produced by 2-pyridylethylamine (2-PEA), 100 μM, in a lung strip. A single experiment.

comparing the EC₅₀ values obtained in the presence and absence of antagonist. The dose-ratios were examined according to the method of Arunlakshana & Schild (1959) where least squares regression analysis was used to determine the relationship between log (dose-ratio - 1) and log of the molar concentration of antagonist.

Results

The effect of H₂-agonists on the unstimulated lung strip

Dimaprit, 1 μM to 1 mM, 4-methyl histamine, 0.5 μM to 100 μM or impromidine, 10 nM to 4 μM, had no effect on the level of tone in the unstimulated lung strip. Therefore, tone was induced in the lung strips by pre-contracting them to examine the relaxant effect of these H₂-agonists.

The effect of 2-pyridylethylamine on the lung strip

Carbachol, 0.1 μM to 100 μM and histamine, 1 μM to 1 mM were not used to induce tone in the lung strips because contractions produced by these two compounds were not sustained.

2-PEA, a specific H₁-agonist (Durant *et al.*, 1975), produced dose-related contractions of strips of guinea-pig lung parenchyma at concentrations between 1 μM and 1 mM. A dose-response curve for 2-PEA has been constructed from the results pooled from five experiments (Figure 1). The EC₅₀ for 2-PEA was found to be 50 ± 6 μM (mean ± s.e.mean) and the EC₅₀ for histamine was 6 ± 2 μM. The maximum response to 2-PEA occurred with a concentration of approximately 800 μM and was equivalent to 98.5% of the maximum response to histamine, which was achieved with a concentration of 100 μM. In a paired *t* test of the individual experimental results, the maximum response to 2-PEA was found not to be

significantly different from the maximum response to histamine ($P > 0.05$). A concentration of 2-PEA of $100 \mu\text{M}$ produced approximately 70% of the maximum response and this was used to induce tone in the lung strips prior to examining any relaxant effects of the H_2 -agonists. Contractions produced by 2-PEA, $100 \mu\text{M}$, were sustained for at least 1 h. Taking the contraction produced by 2-PEA, $100 \mu\text{M}$ at 5 min after addition as unity, the contraction recorded after 20, 40 and 60 min was 1.1, 0.9 and 1.0 times the 5 min value.

The effect of H_2 -agonists on the lung strip contracted by 2-pyridylethylamine

Cumulative addition of dimaprit to give concentrations within the range $1 \mu\text{M}$ to $400 \mu\text{M}$ produced a dose-dependent relaxation of the lung parenchymal strips which had been precontracted by 2-PEA, $100 \mu\text{M}$, (Figure 2). A cumulative concentration of dimaprit of greater than $400 \mu\text{M}$ produced no further relaxation of the lung strips. After washing, the tissue returned to its original resting tension. A dose-response curve has been constructed from the results pooled from five experiments (Figure 3). The EC_{50} for dimaprit was found to be $17.4 \pm 3.3 \mu\text{M}$. The maximum relaxation produced by cumulative addition of dimaprit to give a final concentration of $400 \mu\text{M}$ was a reduction of $89 \pm 2\%$ of the contraction produced by 2-PEA, $100 \mu\text{M}$ and the maximum relaxation produced by a single dose of dimaprit of $400 \mu\text{M}$ was a reduction of $82 \pm 12\%$ of the contraction produced by 2-PEA, $100 \mu\text{M}$. A non-paired *t* test of the individual experimental results showed no significant difference ($P > 0.05$) between the maximum relaxation produced by cumulative addition or single addition of dimaprit to give a final concentration of $400 \mu\text{M}$ in both cases.

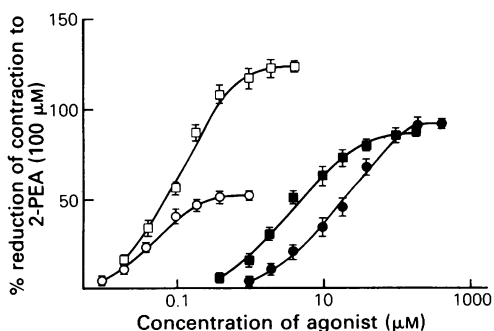


Figure 3 Dose-response curves showing the effect of dimaprit (●), 4-methyl histamine (■), impromidine (○) and forskolin (□) on the lung strip which had been contracted by 2-pyridylethylamine (2-PEA), $100 \mu\text{M}$. Responses have been expressed as a percentage reduction of the contraction produced by 2-PEA ($100 \mu\text{M}$) for a particular strip. Points are means of four to six determinations and the vertical bars represent s.e.mean.

4-Methyl histamine, $0.5 \mu\text{M}$ to $100 \mu\text{M}$ and impromidine, 10 nM to $1 \mu\text{M}$, also produced a dose-dependent relaxation of the lung strip contracted by 2-PEA, $100 \mu\text{M}$. Dose-response curves have been constructed for each of these compounds from the results pooled from four experiments (Figure 3). The mean EC_{50} values for 4-methyl histamine and impromidine were found to be $4 \pm 2 \mu\text{M}$ and $0.04 \pm 0.01 \mu\text{M}$ respectively. The maximum relaxation produced by 4-methyl histamine was equivalent to $85 \pm 2\%$ and that by impromidine $52 \pm 6\%$ reversal of the contraction produced by 2-PEA, $100 \mu\text{M}$. These maximum relaxations occurred with concentrations of $200 \mu\text{M}$ and $1 \mu\text{M}$ respectively.

The effect of forskolin on the lung strip

Forskolin does not combine with histamine H_2 -receptors but has a direct stimulatory effect on adenylate cyclase which is the enzyme thought to be activated when H_2 -receptors are stimulated. Cumulative addition of forskolin to give concentrations within the range 10 nM to $4 \mu\text{M}$ produced a dose-dependent relaxation of both unstimulated lung strips and lung strips which had been precontracted by 2-PEA, $100 \mu\text{M}$. In each case, dose-response curves have been constructed by pooling results from four experiments (Figures 3 and 4). The mean EC_{50} values for forskolin

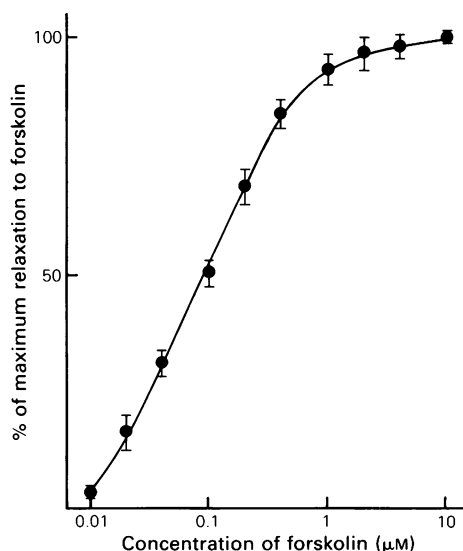


Figure 4 Dose-response curve showing the effect of forskolin on the resting level of tone in the guinea-pig lung strip. Responses have been expressed as a percentage of the maximum relaxation produced by forskolin on a particular strip. This maximum relaxation is equivalent to a mean (\pm s.e.mean) lengthening of the lung strip of $1.2 \pm 0.3 \text{ mm}$.

were found to be $0.11 \pm 0.01 \mu\text{M}$ for relaxation of resting tone and $0.16 \pm 0.04 \mu\text{M}$ for relaxation of lung strips pre-contracted by 2-PEA, $100 \mu\text{M}$.

In the experiments using the lung strips with resting tone, the maximum relaxation produced by forskolin was equivalent to a mean lengthening of the lung strip of $1.2 \pm 0.3 \text{ mm}$ and occurred with a concentration of $2.5 \mu\text{M}$. When precontracted lung strips were used the maximum relaxation produced by forskolin was a reduction of 130% of the contraction produced by 2-PEA, $100 \mu\text{M}$ and occurred with a concentration of forskolin of $1 \mu\text{M}$. The relaxation produced by forskolin on the 2-PEA-contracted strip (Figure 3) is an effect of forskolin on basal as well as stimulated contraction.

The effect of atropine and propranolol on the response to dimaprit

To establish that dimaprit was not causing relaxation of the lung strip by release of catecholamines we examined the effect of propranolol on the response to dimaprit and to determine whether released acetylcholine was affecting the level of tone in the lung strip and thereby modifying the dimaprit response, we examined the effect of atropine on the dimaprit dose-response curve.

Neither atropine, 0.1 to $10 \mu\text{M}$, nor propranolol, 1 to

$100 \mu\text{M}$, had any effect themselves on the level of tone in the unstimulated guinea-pig lung strip or on the lung strip which had been precontracted by 2-PEA, $100 \mu\text{M}$. The effects of atropine, $1 \mu\text{M}$ and propranolol, $10 \mu\text{M}$ have been studied on the cumulative dose-response curve to dimaprit in the lung strip. The mean EC_{50} values for dimaprit in the absence and presence of atropine, $1 \mu\text{M}$ were calculated as $15 \pm 2 \mu\text{M}$ ($n = 6$) and $18 \pm 3 \mu\text{M}$ ($n = 6$) and in the absence and presence of propranolol, $10 \mu\text{M}$ as $25 \pm 5 \mu\text{M}$ ($n = 6$) and $25 \pm 10 \mu\text{M}$ ($n = 6$) respectively. The differences between the EC_{50} values for dimaprit in the absence or in the presence of these drugs were not statistically significant ($P > 0.05$).

The effect of H₂-antagonists on the response to dimaprit

All of the H₂-antagonists examined, at concentrations up to $100 \mu\text{M}$, had no effect on the level of tone in the unstimulated lung strip. Using the experimental design described in the Methods, the effects of several H₂-antagonists on the dose-response curve for dimaprit were studied. Increasing concentrations of the H₂-antagonists caused a graded, parallel displacement to the right of the dimaprit dose-response curve, with no reduction in the maximum response to dimaprit. The effect of the H₂-antagonist burimamide is shown in Figure 5 as an example. As described in the

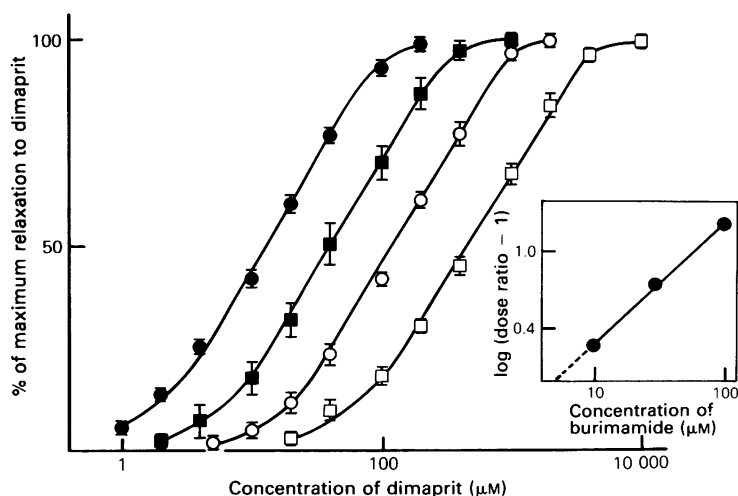


Figure 5 The antagonism of the response to dimaprit by burimamide. Dose-response curves are to dimaprit alone (●) and dimaprit in the presence of burimamide, $10 \mu\text{M}$ (■), $30 \mu\text{M}$ (○) and $100 \mu\text{M}$ (□). The curves have been constrained to be parallel. Responses have been expressed as a percentage of the maximum relaxation produced by dimaprit on a particular strip. Points are means of 65 to 72 determinations in the control curve and 4 to 6 determinations in the other curves. The vertical bars represent s.e.mean. The figure shows the control curve constructed by pooling results of all experiments. However the K_B values and Schild plots have been calculated from analysis of dose-response curves in the absence and presence of antagonist that were obtained in the same experiment. Inset: Schild plot constructed as described in Methods, uses dose-response curves for dimaprit in the presence and absence of burimamide obtained in individual experiments.

Table 1 The effect of H₂-antagonists on the dose-response curve for dimaprit

H ₂ -antagonist	Concentration of H ₂ -antagonist (μM)	n	Dose-ratio (mean with 95% confidence limits)	K _B (μM)	Slope of Schild plot
Tiotidine	0.01	4	3.8 (2.5– 5.0)	0.0037	1.00
	0.03	4	10.5 (7.8–13.2)		
	0.1	4	29.0 (24.5–33.5)		
YM11170	0.1	4	2.9 (2.1– 3.7)	0.5	1.05
	0.3	6	9.1 (6.8–11.4)		
	1	4	21.0 (17.5–26.3)		
Ranitidine	1	6	3.5 (2.0– 5.0)	0.35	0.90
	3	4	7.9 (5.8–10.0)		
	10	6	19.1 (15.2–23.0)		
Cimetidine	1	4	2.4 (1.5– 3.3)	0.71	0.98
	3	6	5.4 (3.3– 7.5)		
	10	6	15.1 (11.8–18.4)		
Metiamide	1	6	2.1 (1.8– 2.4)	0.91	1.15
	3	6	5.5 (3.4– 7.6)		
	10	4	18.2 (15.5–20.9)		
Burimamide	10	6	3.3 (2.4– 4.2)	5.1	1.10
	30	4	10.0 (6.9–13.1)		
	100	4	38.0 (31.8–44.2)		

Dose-ratios and equilibrium dissociation constants (K_B) were calculated as described in Methods. *n* indicates the number of experiments from which results were pooled.

Methods, dose-ratios have been calculated for each concentration of each H₂-antagonist used, and these are summarised in Table 1. Schild plots have been constructed for each H₂-antagonist on the basis of the dose-ratios shown in Table 1, and burimamide is again shown as an example in the inset to Figure 5. The equilibrium dissociation constants and accompanying slopes for the H₂-antagonists obtained from these Schild plots are also shown in Table 1. None of the slopes calculated from the Schild plots were significant-

tly different from one ($P > 0.05$).

The highest concentration of each H₂-antagonist used in the experiments described above has also been examined for its effect on the dose-response curve to 2-PEA. The results have been summarised in Table 2. In all cases the H₂-antagonists had no effect on the dose-response curve for 2-PEA, and the EC₅₀ values for 2-PEA in the absence and presence of each H₂-antagonist were not significantly different ($P > 0.05$).

Table 2 The effect of H₂-antagonists on the dose-response curve for 2-pyridylethylamine (2-PEA)

H ₂ -antagonist	Concentration (μM)	Parameters for 2-PEA			
		Absence of antagonist		Presence of antagonist	
		EC ₅₀ (μM)	Maximum response (%)	EC ₅₀ (μM)	Maximum response (%)
Tiotidine	0.1	55 ± 10	100	53 ± 7 ^{NS}	98 ± 4
YM11170	1	50 ± 8	100	48 ± 9 ^{NS}	103 ± 5
Ranitidine	10	67 ± 14	100	69 ± 17 ^{NS}	99 ± 3
Cimetidine	10	55 ± 6	100	52 ± 13 ^{NS}	97 ± 6
Metiamide	10	61 ± 12	100	64 ± 17 ^{NS}	98 ± 10
Burimamide	100	35 ± 10	100	32 ± 8 ^{NS}	105 ± 6

EC₅₀ values were calculated as described in Methods. The values given are the means ± s.e. mean from six experiments. NS = not significantly different ($P > 0.05$) from the EC₅₀ values obtained in the absence of antagonist.

Discussion

The effect of H₂-agonists were studied against contractions produced by 2-PEA because this H₁-agonist gave a sustained contraction of the tissue. It is not known why histamine and carbachol failed to produce sustained contractions but these compounds may be taken up or metabolised by the tissue or histamine may exert H₂-effects (relaxation) in addition to an H₁-mediated contraction.

There is now substantial evidence that H₂-antagonists potentiate the contractile action of histamine on the guinea-pig lung strip (Dunlop & Smith, 1977; Okpako *et al.*, 1978; Drazen *et al.*, 1980) but have no effect on contractions produced by the selective H₁-agonist, 2-PEA (Okpako *et al.*, 1978). It has been suggested that this is the result of histamine having an action on both H₁ and H₂-receptors whereas the effect of 2-PEA is specifically at the H₁-receptor. In this study we have shown that several H₂-antagonists had no effect on the dose-response curve for 2-PEA on the lung strip (Table 2) which suggests that 2-PEA has no agonist activity at histamine H₂-receptors. It is, in fact, unlikely that 2-PEA would be able to produce relaxation of the lung strip by an action on H₂-receptors since its H₁:H₂ selectivity ratio has been reported to be 30:1 (Durant *et al.*, 1975).

The EC₅₀ for 2-PEA of 50 μM calculated in this study is in good agreement with that of 40 μM previously reported for 2-PEA-induced contraction of the lung strip (Chand & Deroth, 1979). The maximum response to 2-PEA was approximately equal to the maximum response to histamine in this study and this finding is also in agreement with other work (Chand & Deroth, 1979; Drazen *et al.*, 1979).

In this study, relaxation of the guinea-pig lung parenchymal strip by each H₂-agonist could only be demonstrated in the presence of induced tone. This is in agreement with the work of Vincenc *et al.* (1984) who found that dimaprit could not produce relaxation of unstimulated human lung strips. This may suggest that there is little tone present in the lung strip in its resting state. However, it has previously been shown that β₂-adrenoceptor agonists are able to produce relaxation of guinea-pig lung strips which have not been precontracted (Goldie *et al.*, 1982; Carswell & Nahorski, 1983) and indeed, in this study, forskolin was able to produce a relaxation of unstimulated lung strips suggesting that the tissue has the ability to relax further when in a resting state. Although the low efficacy of impromidine (Durant *et al.*, 1978), which produced only a 50% reversal of the contraction to 2-PEA in this study, could explain why it had no measurable effect on resting tone, this explanation is unlikely because both dimaprit and 4-methyl histamine, which produced approximately 90% reversal of the contraction to 2-PEA, also had no measurable

effect on resting tone. Moreover it has previously been reported that dimaprit produces at least the same maximum response as histamine when acting at H₂-receptors (Parsons *et al.*, 1977), and indeed, Bottomley *et al.* (1985) have suggested that both dimaprit and 4-methyl histamine have a two to three times higher efficacy than histamine at H₂-receptors in the guinea-pig right atrium. Hence, it is unlikely that the efficacy of dimaprit and 4-methyl histamine is too low to show a response on the resting lung strip.

Another explanation for the lack of activity of the H₂-agonists on resting tone is that the H₂-receptors in the lung strip may only become activated when the contractile mechanism of the lung strip is stimulated either chemically or mechanically. This argument is supported by the work using forskolin which was able to produce a relaxation of the lung strips under resting tension but this drug activates adenylate cyclase directly without stimulating H₂-receptors. Forskolin produced 130% reversal of the contraction to 2-PEA on the lung strip compared to 90% reversal produced by dimaprit and 4-methyl histamine. This suggests that, in our study, there is some tone present in the unstimulated lung strip. However, it may not be sufficient to cause activation of the H₂-receptors. Previous workers have suggested that dimaprit is able to produce a relaxation of resting tone in the lung strip but in those studies the tissues were maintained at a much higher tension (Yen & Kreutner, 1979; Tomioka & Yamada, 1982). In our study, a much lower tension was applied to the lung strips and they were allowed to relax completely before addition of an H₂-agonist.

Forskolin gave EC₅₀ values of 0.11 μM and 0.16 μM for relaxation of lung strips under resting tension and lung strips contracted by 2-PEA respectively. Both these figures are in good agreement with EC₅₀ values of 0.2 μM and 0.4 μM reported respectively for relaxation of normal and sensitized guinea-pig lung strips (Burka & Saad, 1984).

The EC₅₀ values obtained in this study, for relaxation induced by dimaprit, 4-methyl histamine and impromidine were 17 μM, 2 μM and 0.04 μM respectively. Similar EC₅₀ values have been reported for the action of these H₂-agonists on H₂-receptors in the guinea-pig right atrium (Gajtkowski *et al.*, 1983). In the case of dimaprit and 4-methyl histamine, similar values were obtained for relaxation of lung strips precontracted with 2-PEA (Tomioka & Yamada, 1982).

It is unlikely that the action of dimaprit on the lung strip is due to release of catecholamines since propranolol did not affect the response to dimaprit. This result is in agreement with those of Parsons *et al.* (1977) where propranolol had no effect on the positive chronotropic effect of dimaprit in the guinea-pig right atrium.

The order of potency for the H₂-agonists and the

lack of effect of non-H₂-receptor antagonists suggests that dimaprit, 4-methyl histamine and impromidine are stimulating the histamine H₂-receptor to produce relaxation of the guinea-pig lung strip. This is substantiated by the parallel shift of the dose-response curve for dimaprit produced by several concentrations of a variety of H₂-receptor antagonists (Figure 4). There was also no reduction in the maximum response to dimaprit in the presence of the H₂-antagonists and this together with the fact that the slopes of the Schild plots for the H₂-antagonists were not significantly different from one is compatible with surmountable competitive antagonism between dimaprit and the H₂-antagonists at an H₂-receptor within the lung paren-

chymal strip. However, for a more precise investigation of the receptor type causing the relaxation of the lung strip produced by dimaprit it is necessary to compare the dissociation constants for the H₂-antagonists obtained in this study with their dissociation constants obtained in other pharmacological and biochemical studies and this has been done in the paper which follows.

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References

- ARUNLAKSHANA, O. & SCHILD, H.O. (1959). Some quantitative uses of drug antagonists. *Br. J. Pharmac. Chemother.*, **14**, 48–58.
- ASH, A.S.F. & SCHILD, H.O. (1966). Receptors mediating some actions of histamine. *Br. J. Pharmac. Chemother.*, **27**, 427–439.
- BERTRAM, J.F., GOLDIE, R.G., PAPADIMITRIOU, J.M. & PATERSEN, J.W. (1983). Correlations between pharmacological responses and structure of human lung parenchymal strips. *Br. J. Pharmac.*, **80**, 107–114.
- BLACK, J.W., DUNCAN, W.A.M., DURANT, C.J., GANELLIN, C.R. & PARSONS, M.E. (1972). Definition and antagonism of histamine H₂-receptors. *Nature*, **236**, 385–390.
- BLACK, J.W., DUNCAN, W.A.M., EMMET, J.C., GANELLIN, C.R., HESSELBO, T., PARSONS, M.E. & WYLLIE, J.H. (1973). Metiamide – an orally active histamine H₂-receptor antagonist. *Agents & Actions*, **3**, 133–138.
- BOTTOMLEY, K.M.K., LEWIS, S., RISING, T.J. & STEWARD, A. (1984). A new *in vivo* method for the measurement of repetitive anaphylactic responses in the guinea-pig. *Br. J. Pharmac.*, **81**, 195–199.
- BOTTOMLEY, K.M.K., RISING, T.J. & STEWARD, A. (1985). Relative efficacies of histamine H₂-receptor agonists in the guinea-pig right atrium. *Br. J. Pharmac.*, **84**, 47P.
- BRADSHAW, J., BRITAIN, R.T., CLITHEROW, J.W., DALY, M.J., JACK, D., PRICE, B.J. & STABLES, R. (1979). Ranitidine (AH 19065): a new, potent, selective histamine H₂-receptor antagonist. *Br. J. Pharmac.*, **66**, 464P.
- BRIMBLECOMBE, R.W., DUNCAN, W.A.M., DURANT, C.J., GANELLIN, C.R., PARSONS, M.E. & BLACK, J.W. (1975). The pharmacology of cimetidine, a new histamine H₂-receptor antagonist. *Br. J. Pharmac.*, **53**, 435–436P.
- BURKA, J.F. & SAAD, M.H. (1984). Bronchodilator-mediated relaxation of normal and ovalbumin-sensitised guinea-pig airways: lack of correlation with lung adenylate cyclase activation. *Br. J. Pharmac.*, **83**, 645–655.
- CARSWELL, H. & NAHORSKI, S.R. (1983). β -Adrenoceptor heterogeneity in guinea-pig airways: comparison of functional and receptor labelling studies. *Br. J. Pharmac.*, **79**, 965–971.
- CHAND, N. (1979). *In vitro* anaphylaxis in guinea-pig lung: evidence for the protective role of histamine H₂-receptors. *Eur. J. Pharmac.*, **55**, 337–339.
- CHAND, N. & DE ROTH, L. (1979). Dual histamine receptor mechanism in guinea-pig lung. *Pharmacology*, **19**, 185–190.
- CHAND, N., DHAWAN, B.N., SRIMAL, R.C., RAHMANI, N.H., SHUKLA, R.K. & ALTURA, B.M. (1980). Reactivity of trachea, bronchi and lung strips to histamine and carbachol in rhesus monkeys. *J. appl. Physiol.*, **19**, 729–734.
- CLAYTON, D.E., BUSSE, W.W. & BUCKNER, C.K. (1981). Contribution of vascular smooth muscle to contractile responses of guinea-pig isolated lung parenchymal strips. *Eur. J. Pharmac.*, **70**, 311–320.
- DEWS, P.B. & GRAHAM, J.D.P. (1946). The antihistamine substance 2786 R.P. *Br. J. Pharmac. Chemother.*, **1**, 278–286.
- DRAZEN, J.M. & AUSTEN, K.F. (1974). Effects of intravenous administration of slow-reacting substance of anaphylaxis, histamine, bradykinin and prostaglandin F_{2α} on pulmonary mechanics in the guinea-pig. *J. clin. Invest.*, **53**, 1679–1705.
- DRAZEN, J.M. & SCHNEIDER, M.W. (1978). Comparative responses of tracheal spirals and parenchymal strips to histamine and carbachol, *in vitro*. *J. clin. Invest.*, **61**, 1441–1447.
- DRAZEN, J.M., SCHNEIDER, M.W. & VENUGOPALAN. (1979). Bronchodilator activity of dimaprit in the guinea-pig *in vitro* and *in vivo*. *Eur. J. Pharmac.*, **55**, 233–239.
- DRAZEN, J.M., VENUGOPALAN, C.S. & SCHNEIDER, M.W. (1980). Alteration of histamine response by H₂-receptor antagonism in the guinea-pig. *J. appl. Physiol. Respir. Environ. Exercise Physiol.*, **48**, 613–618.
- DURANT, G.J., GANELLIN, C.R. & PARSONS, M.E. (1975). Chemical differentiation of histamine H₁- and H₂-receptor agonists. *J. med. Chem.*, **18**, 905–909.
- DURANT, G.J., DUNCAN, W.A.M., GANELLIN, C.R., PARSONS, M.E., BLAKEMORE, R.C. & RASMUSSEN, A.C. (1978). Impromidine (SK & F 92676) is a very potent and specific agonist for histamine H₂-receptors. *Nature*, **276**, 403–404.
- DUNLOP, L.S. & SMITH, A.P. (1977). The effect of histamine antagonists on antigen-induced contractions of sensitised human bronchus *in vitro*. *Br. J. Pharmac.*, **59**, 475P.
- EYRE, P. (1977). Pulmonary histamine H₁- and H₂-receptor studies. In *Asthma II: Physiology, Immunopharmacology and Treatment*. ed. Austen, K.F. & Lichtenstein, L. pp. 169–177. New York: Academic Press.

- FOREMAN, J.C., NORRIS, D.B., RISING, T.J. & WEBBER, S.E. (1985). The specific binding of [³H]-tiotidine to homogenates of guinea-pig lung parenchyma. *Br. J. Pharmac.*, **86**, 475–482.
- GADDUM, J.H., HAMED, K.A., HATHAWAY, D.E. & STEPHENS, F.F. (1955). Quantitative studies of antagonists for 5-hydroxytryptamine. *Q. J. exp. Physiol.*, **40**, 49–74.
- GAJTKOWSKI, G.A., NORRIS, D.B., RISING, T.J. & WOOD, T.P. (1983). Specific binding of [³H]tiotidine to histamine H₂-receptors in guinea-pig cerebral cortex. *Nature*, **304**, 65–67.
- GOLDIE, R.G., PATERSEN, J.W. & WALE, J.L. (1982). Pharmacological responses of human and porcine lung parenchyma, bronchus and pulmonary artery. *Br. J. Pharmac.*, **76**, 515–521.
- LULICH, K.M., MITCHELL, H.W. & SPARROW, M.P. (1976). The cat lung strip as an *in vitro* preparation of peripheral airways: a comparison of β -adrenoreceptor agonists, autocooids and anaphylactic challenge on the lung strip and trachea. *Br. J. Pharmac.*, **58**, 71–79.
- MARQUARDT, D.W. (1963). An algorithm for least squares estimation of non-linear parameters. *J. Soc. Indust. appl. Math.*, **11**, 431–441.
- OKPAKO, D.T., CHAND, N. & EYRE, P. (1978). The presence of inhibitor histamine H₂-receptors in guinea-pig tracheobronchial smooth muscle. *J. Pharm. Pharmac.*, **30**, 181–182.
- PARSONS, M.E., OWEN, D.A.A., GANELLIN, C.R. & DURANT, G.J. (1977). Dimaprit-[S[3-(N,N-dimethylamino)propyl]isothiourea] – a highly specific histamine H₂-receptor agonist. Part 1. Pharmacology. *Agents & Actions*, **7**, 31–37.
- TOMIOKA, K. & YAMADA, T. (1982). Effects of histamine H₂-receptor agonists and antagonists on isolated guinea-pig airway muscles. *Archs int. Pharmacodyn.*, **255**, 16–26.
- VINCENC, K., BLACK, J. & SHAW, J. (1984). Relaxation and contraction responses to histamine in the human lung parenchymal strip. *Eur. J. Pharmac.*, **98**, 201–210.
- YEN, S.S. & KREUTNER, W. (1979). Histamine H₂-receptors in guinea-pig peripheral airway smooth muscle. *Life Sci.*, **25**, 507–514.

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