

Prostaglandin D₂ interacts at thromboxane receptor-sites on guinea-pig platelets

S. Hamid-Bloomfield & B.J.R. Whittle

Department of Mediator Pharmacology, Wellcome Research Laboratories, Langley Court, Beckenham, Kent BR3 3BS

1 The anti-aggregatory prostanoid, prostaglandin D₂ (PGD₂) does not completely inhibit ADP-induced aggregation of guinea-pig platelets and thus produces a bell-shaped dose-inhibition curve. The nature of this bell-shaped curve has now been investigated in guinea-pig platelet-rich plasma.

2 Two selective thromboxane receptor antagonists, 13-aza-prostanoic acid (13-AZA; 16–64.4 μM) and BM 13.177 (5.9–29.8 μM), converted PGD₂ to a full inhibitor of aggregation in a dose-related manner.

3 The putative platelet PGD₂ receptor antagonist, N-0164 (75 μM) also converted PGD₂ to a full inhibitor of platelet aggregation. In contrast to 13-AZA and BM 13.177, higher concentrations of N-0164 (380 and 760 μM) caused a dose-related rightward shift of the PGD₂ dose-inhibition curve.

4 The thromboxane receptor antagonism of N-0164 was confirmed in studies in which the dose-aggregation curve to U-46619, a thromboxane mimetic, was competitively antagonized with a pA₂ value of 4.67 and a slope of 1.13, comparable to that of 13-AZA.

5 The results show that N-0164 acts as both a platelet PGD₂ and thromboxane-receptor antagonist in both human and guinea-pig platelet-rich plasma.

6 The results further indicate that PGD₂ can interact at thromboxane receptors in guinea-pig platelets.

Introduction

Prostaglandin D₂ (PGD₂) has potent anti-aggregatory activity on platelets of several species (Whittle *et al.*, 1978). As with the other prostanoid inhibitors of platelet aggregation, prostaglandin E₁ (PGE₁) and prostacyclin (PGI₂), PGD₂ elevates platelet adenosine 3':5'-cyclic monophosphate (cyclic AMP) levels which supports the concept of a similar intracellular mechanism of action for these prostanoids (Tateson *et al.*, 1977; Gorman *et al.*, 1977; Whittle *et al.*, 1978). However, the platelet receptors mediating the responses to PGD₂ appear distinct from those of PGE₁ and PGI₂. Thus, the experimental compound N-0164 in appropriate concentrations selectively antagonized the anti-aggregatory actions of PGD₂ with no effect on those of PGE₁ and PGI₂ on human platelets (MacIntyre & Gordon, 1977; Whittle *et al.*, 1978).

Our previous studies in guinea-pig platelet-rich plasma (PRP) have shown that PGD₂ produces a bell-shaped dose-anti-aggregatory relationship, with complete inhibition of platelet aggregation not being achieved (Hamid & Whittle, 1985). An earlier study has shown that the thromboxane receptor antagonist

EP045 partially blocks the contractile actions of PGD₂ on the guinea-pig trachea which thus could be indicative of an interaction of PGD₂ at thromboxane-sensitive contractile sites (Jones *et al.*, 1982). In the present study, we have therefore investigated whether the interaction of PGD₂ at the thromboxane receptor sites in guinea-pig platelets could explain the nature of the dose-response relationship using two selective thromboxane antagonists, 13-aza-prostanoic acid (13-AZA; Venton *et al.*, 1979) and BM 13.177 (Patscheke *et al.*, 1984). In addition, we have compared the activity of N-0164 as an antagonist of these PGD₂ responses in guinea-pig PRP.

A preliminary account of this work has been presented to the British Pharmacological Society (Hamid & Whittle, 1986).

Methods

The method was as described previously by Hamid & Whittle (1985). Male Halls guinea-pigs (350–450 g)

were anaesthetized with sodium pentobarbitone (60 mg kg⁻¹, i.p.) and 15 ml of blood was collected from the abdominal aorta into plastic tubes containing trisodium citrate (final concentration 0.315%). The blood was centrifuged (1000g) for 2 min to obtain platelet-rich plasma (PRP) which was removed and stored at room temperature. The platelet count in the PRP, determined using a Coulter Counter (model ZF), was $2.8-3.5 \times 10^8$ ml⁻¹.

Platelet aggregation was measured in a Payton dual channel aggregation module connected to a 'W + W' recorder 1200. Samples (0.5 ml) of PRP were incubated for 1 min at 37°C, stirred at 900 r.p.m. with or without the prostaglandin under investigation, prior to addition of submaximal doses of adenosine diphosphate (ADP, 2–4 μM) sufficient to just cause a non-reversible control aggregation.

Dose-inhibition curves were constructed for PGD₂, prostacyclin (PGI₂) and BW245C and the IC₅₀ was calculated as the concentration required to reduce the aggregation to 50% of its control amplitude.

To study the interaction of PGD₂ at thromboxane receptors, the PRP was preincubated (1 min at 37°C) with the two selective thromboxane antagonists (13-AZA and BM 13.177) before adding the appropriate concentration of PGD₂ for a further 1 min incubation, before the addition of ADP.

The activity of N-0164 as an antagonist of the PGD₂, BW245C and PGI₂ anti-aggregating responses was studied in a comparable fashion. In further studies, to investigate whether N-0164 was acting as a thromboxane receptor antagonist, guinea-pig PRP was preincubated (1 min) with N-0164 (19–760 μM) before the addition of the epoxy-methano endoperoxide analogue U-46619 (0.11–1.14 μM), a thromboxane mimetic (Coleman *et al.*, 1981). Likewise, the actions of N-0164 (38–760 μM) on U-46619 (0.11–2.8 μM)-induced aggregation of human PRP, prepared as described previously (Whittle *et al.*, 1978), were investigated.

Drugs

Prostacyclin as the sodium salt (Wellcome Foundation) was freshly dissolved in 1 M Tris buffer (pH 9.6 at 4°C) immediately prior to use. Prostaglandin D₂; 11α, 9α epoxy-methano PGH₂, U-46619 (from the Upjohn Company, Kalamazoo, U.S.A.) and the hydantoin prostaglandin 5-(6-carboxyhexyl)-1-(3-cyclohexyl-3-hydroxypropyl) hydantoin (BW245C, synthesized in the Dept Therapeutic Chemistry, Wellcome Research Laboratories) were stored in ethanol (10 mg ml⁻¹; 4°C) and diluted with 50 mM Tris buffer (pH 7.5 at 4°C) when required. Adenosine diphosphate (Sigma Chemical Co.) was dissolved in distilled water when required and kept on ice.

The thromboxane antagonists, 13-aza-prostanoic

acid (13-AZA) and 4-[2-(benzenesulphonamide)-ethyl]-phenoxyacetic acid (BM 13.177) were synthesized in the Dept Therapeutic Chemistry, Wellcome Research Laboratories by Dr W. Jackson. These compounds were stored in ethanol (10 mg ml⁻¹; 4°C) and diluted with 50 mM Tris buffer (pH 7.5 at 4°C) when required. Compound N-0164 (sodium *p*-benzyl-4-[1-oxo-2-(4-chlorobenzyl)-3-phenylpropyl]phenyl phosphonate, obtained from Nelson Research and Development Company, California) was freshly dissolved in distilled water (10 mg ml⁻¹ at 37°C).

Statistical analysis

Results are expressed as mean (± s.e.mean), where *n* is the number of values. The difference between groups was evaluated using Student's *t* test for unpaired data, where *P* < 0.05 was taken as significant. Compliance to simple competitive antagonism was tested by comparison of dose-ratios of agonists, obtained in the presence of antagonist, to the Schild equation where the abscissal intercept represents the empirical measure of potency, pA₂ (Arunlakshana & Schild, 1959). Linear line regression analysis with 95% confidence limits was carried out on these data. The slopes and asymptotes of the dose-response curves were analysed using the *F* test of equal curves.

Results

Effects of the thromboxane-receptor antagonists

Pre-incubation of guinea-pig PRP with PGD₂ produced a bell-shaped dose-anti-aggregatory relationship, with complete inhibition of platelet aggregation induced by ADP not being achieved (Figure 1), confirming previous findings (Hamid & Whittle, 1985). The effects of two selective thromboxane receptor antagonists 13-AZA and BM 13.177, on this bell-shaped dose-response relationship with PGD₂ were investigated. Both 13-AZA (16–64.4 μM) and BM 13.177 (5.9–29.8 μM) significantly converted, in a dose-related manner, the responses of PGD₂ to a full inhibition of aggregation (Figures 1 and 2). Higher concentrations of 13-AZA (129 μM) and BM 13.177 (60 μM) did not further shift the PGD₂ curves. BM 13.177 was more potent in converting PGD₂ to a full inhibitor of ADP-induced aggregation than 13-AZA.

Effects of N-0164

N-0164 at a low concentration (75 μM) converted the PGD₂ bell-shaped curve to a sigmoid dose-inhibition curve, exposing it as a full inhibitor of ADP-induced aggregation (Figure 3a). In contrast to 13-AZA and

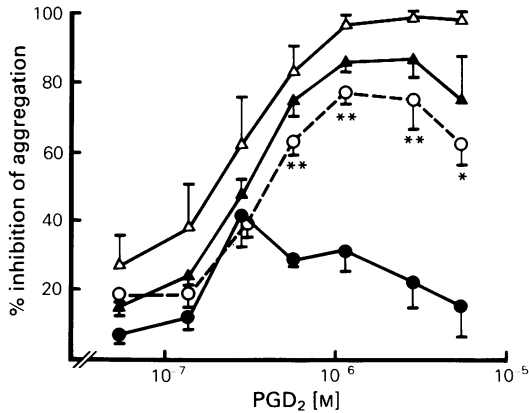


Figure 1 Effect of the thromboxane antagonist 13-azaprostanic acid (13-AZA) on the inhibition of ADP-induced aggregation of guinea-pig platelets by prostaglandin D₂ (PGD₂). Control PGD₂ curve (●, n = 3), PGD₂ in the presence of 16 μM 13-AZA (○, n = 3), PGD₂ + 32 μM 13-AZA (▲, n = 3) and PGD₂ + 64 μM 13-AZA (△, n = 3). Results, expressed as mean (with vertical lines indicating s.e.mean) % inhibition of platelet aggregation, where statistical differences from control PGD₂ curve are shown as *P < 0.05; **P < 0.01.

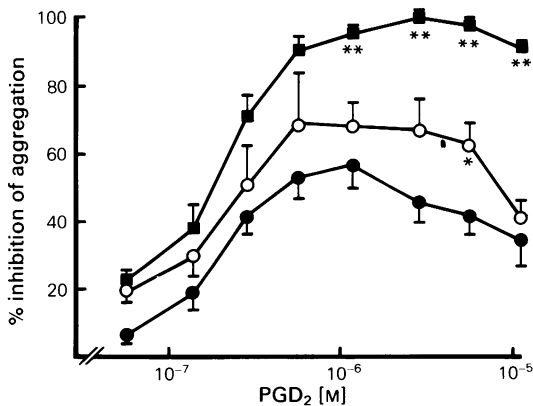


Figure 2 Effect of the thromboxane antagonist BM 13.177 on the inhibition of ADP-induced aggregation of guinea-pig platelets by prostaglandin D₂ (PGD₂). Control PGD₂ curve (●, n = 8), PGD₂ + 6 μM BM 13.177 (○, n = 4), PGD₂ + 30 μM BM 13.177 (■, n = 6). Results, expressed as mean (with vertical lines indicating s.e.mean) % inhibition of platelet aggregation. *P < 0.05; **P < 0.001.

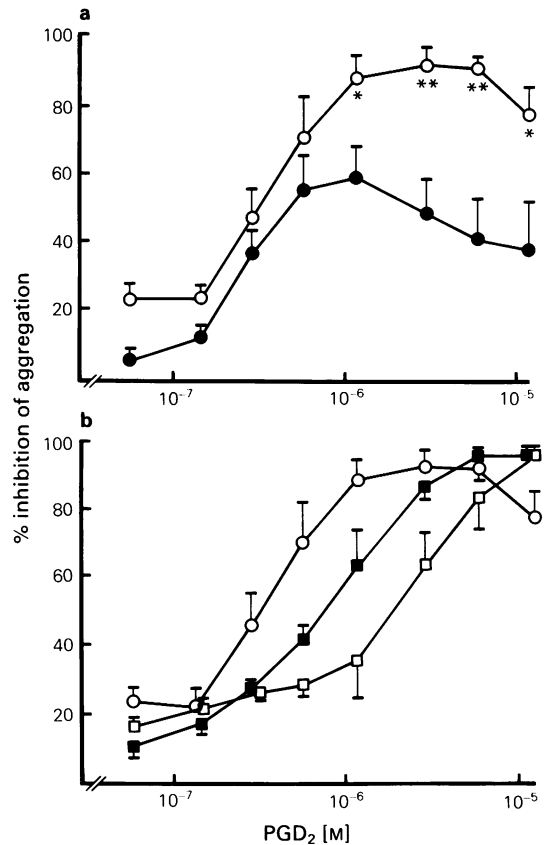


Figure 3 Effect of N-0164 on the inhibition of ADP-induced aggregation of guinea-pig platelets by prostaglandin D₂ (PGD₂). (a) Shows responses to PGD₂ (●, n = 8), PGD₂ + 75 μM N-0164 (○, n = 3), while (b) shows effect of PGD₂ + 75 μM N-0164 (○, n = 3), PGD₂ + 380 μM N-0164 (■, n = 4) and PGD₂ + 760 μM N-0164 (□, n = 4). Results shown are means (with vertical lines indicating s.e.mean). *P < 0.05; **P < 0.001.

BM 13.177, increased concentrations of N-0164 (380 and 760 μM) caused a dose-related rightward shift of PGD₂ dose-inhibitory response curves, with the IC₅₀ value (0.29 ± 0.02 μM, n = 4) being increased to 0.9 ± 0.18 and 2 ± 0.6 μM, respectively (P < 0.001; Figure 3b). These concentrations of N-0164 themselves had no consistent effect on the aggregation induced by ADP. Due to the complex nature of the PGD₂ dose-inhibition curve through its apparent interaction at thromboxane receptors, it was not possible to carry out a true Schild plot regression analysis to obtain a pA₂ value for N-0164 as an antagonist of PGD₂. However, a calculated pA₂ value of 3.6 for this action was derived. Furthermore, in the

Table 1 Estimates of N-0164 as a prostaglandin D₂ (PGD₂) antagonist and of N-0164 and 13-aza-prostanoic acid (13-AZA) as thromboxane receptor antagonists (using the epoxy-methano endoperoxide analogue U-46619 as the thromboxane agonist) in the guinea-pig platelet-rich plasma

Agonist	Antagonist	pA_2	Confidence limits (95%)	n	Slope	Confidence limits (95%)
PGD ₂	N-0164	3.6 (estimated)	—	(4)	—	—
PGD ₂ in the presence of BM 13.177	N-0164	3.64 (estimated)	—	(3)	—	—
U-46619	N-0164	4.67	4.87–4.52	(3)	1.13	0.93–1.32
U-46619	13-AZA	4.83	4.92–4.75	(3)	1.36	1.15–1.57

The results are expressed as pA_2 values and Schild-plot slope values (with 95% confidence limits) from (n) experiments.

presence of BM 13.177 (30 μ M) in a concentration which converted PGD₂ to a full inhibitor of aggregation, N-0164 (760 μ M) produced a rightward shift of the PGD₂ dose-inhibitory response curve with the IC₅₀ value (0.2 \pm 0.02 μ M, n = 6) being increased to 0.9 \pm 0.16 μ M (n = 4, P < 0.001). Under these conditions, the pA_2 value for N-0164 as a PGD₂ antagonist was calculated to be 3.64 (Table 1).

Analysis using the F test of equal curves showed that the dose-response curve to PGI₂ was not significantly different from that in the presence of N-0164 (760 μ M, P > 0.05); only the lower asymptote was found to be different from the control curve (Figure 4). In contrast, N-0164 (760 μ M) not only changed the lower asymptote of the BW245C dose-response curve, but also significantly changed the slope of the curve (P < 0.0001) and caused a rightward shift as shown in Figure 4. There was no significant difference in the

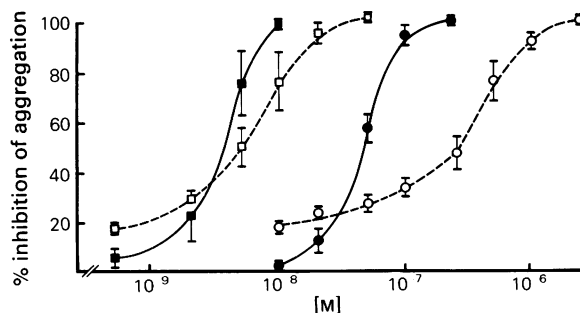


Figure 4 Effect of N-0164 on prostacyclin (PGI₂)- or BW245C-induced inhibition of guinea-pig platelet aggregation. Control PGI₂ (■, n = 4), in the presence of 760 μ M N-0164 (□, n = 4); control BW245C (●, n = 4) curve in the presence of N-0164 (760 μ M, ○, n = 4). Results shown are means (with vertical lines indicating s.e.mean).

IC₅₀ for PGI₂ in the presence of 760 μ M N-0164 (3.5 \pm 0.8 nM and 6.4 \pm 1.2 nM respectively), whereas the IC₅₀ value for BW245C was increased from 46 \pm 4 nM to 277 \pm 55 nM (P < 0.01).

Thromboxane-receptor antagonism by N-0164

To investigate further whether N-0164 was acting as a thromboxane receptor antagonist, its effects on aggregation induced by U-46619, a thromboxane mimetic (Coleman *et al.*, 1981), were evaluated. N-0164 (38–760 μ M) shifted the U-46619 dose-aggregation curve to the right in a concentration-related manner, with a pA_2 value of 4.67 in guinea-pig PRP (Table 1). This value was comparable to that obtained with the selective thromboxane receptor antagonist 13-AZA in guinea-pig platelets (Table 1). Likewise, in three experiments with human PRP, N-0164 induced a rightward shift of the dose-aggregation curve to U-46619, with a pA_2 of 4.09 (4.28–3.96) and a slope of 1.05 (0.76–1.34).

Discussion

We have previously shown that PGD₂ produced a bell-shaped dose-inhibition curve in guinea-pig PRP, with complete inhibition of ADP-induced platelet aggregation not being achieved (Hamid & Whittle, 1985). We proposed several possibilities to explain this bell-shaped curve, one of them being that at high concentrations PGD₂ might be interacting with other stimulatory and pro-aggregatory receptors on guinea-pig platelets, such as those for thromboxane A₂. In an earlier study, Jones *et al.* (1982) suggested that PGD₂ might be interacting at thromboxane-sensitive contractile sites since EP045, a thromboxane receptor antagonist, partially blocked the contractile actions of PGD₂ on the guinea-pig trachea. It has further been

suggested that the constriction of a helical strip of the dog cerebral artery by PGD₂ could be mediated via PGF_{2α} and/or thromboxane receptors (Narumiya & Toda, 1985).

Using two selective thromboxane antagonists of divergent structures, 13-AZA and BM 13.177, the results of the present study support the suggestion that at high concentrations PGD₂ can interact at platelet thromboxane receptors. Both, 13-AZA and BM 13.177, converted PGD₂ to a full inhibitor of ADP-induced aggregation of guinea-pig platelets in a dose-related manner, BM 13.177 being the more potent. Hence it appears that the secondary phase of the bell-shaped curve to PGD₂ at high concentrations reflects stimulation of thromboxane receptors. In the human PRP, however, PGD₂ acts as a full potent inhibitor of ADP-induced platelet aggregation and probably would require much higher concentrations to stimulate the thromboxane receptor sites. Although the hydantoin BW245C is considered to interact with platelet PGD₂ receptors (Whittle *et al.*, 1983; Town *et al.*, 1983; Hamid & Whittle, 1985) 13-AZA did not affect its full dose-inhibition curve, indicating that this compound does not interact at platelet thromboxane receptors.

Previous studies on human platelets have shown that N-0164 acts as a selective antagonist of the anti-aggregatory actions of PGD₂ (MacIntyre & Gordon, 1977; Whittle *et al.*, 1978). In the present study in guinea-pig PRP, N-0164 at low concentrations (75 μM), like the thromboxane antagonists 13-AZA and BM 13.177, converted the PGD₂ bell-shaped curve to a sigmoid dose-inhibition curve, exposing it as a full inhibitor of aggregation. In contrast to the thromboxane receptor antagonists, however, increased concentrations of N-0164 (380 and 760 μM) caused a dose-related rightward shift of the PGD₂ dose-inhibition curves, indicating antagonism at the PGD₂ receptor site. The apparent pA₂ value of N-0164 as a PGD₂ antagonist was very similar when calculated in the absence and presence of BM 13.177. Furthermore, although N-0164 did not significantly shift the dose-inhibition curve for PGI₂, it did cause a rightward shift

of the curve for BW245C. These findings are consistent with the previous studies that PGD₂ acts at platelet receptor sites distinct from those of prostacyclin, and that BW245C interacts with PGD₂ (but not thromboxane)-sensitive sites (Whittle *et al.*, 1978, 1983; Seigl *et al.*, 1979; Schafer *et al.*, 1979; Miller & Gorman, 1979).

Since low concentrations of N-0164 converted the PGD₂ bell-shaped curve to that of a full inhibitor of aggregation in a manner similar to those of the two thromboxane antagonists, it was of interest to determine whether N-0164 could act as an antagonist against platelet aggregation induced by U-46619, a thromboxane mimetic (Coleman *et al.*, 1981). N-0164 acted as a thromboxane antagonist with a pA₂ value comparable to that of the thromboxane antagonist 13-AZA in both guinea-pig and human platelets. Although N-0164 has been found to be a PGD₂ antagonist in the platelet (MacIntyre & Gordon, 1977; Whittle *et al.*, 1978), the present study suggests that it is more potent as a thromboxane receptor antagonist in guinea-pig platelets.

These results thus show that there is a fine balance and interaction between PGD₂ and thromboxane receptor-sites. In guinea-pig platelets, PGD₂ appears to have at least two actions, those at PGD₂ receptor-sites and those at thromboxane receptor sites. The close relationship between platelet PGD₂ receptors and thromboxane receptors is also reflected in the finding that N-0164, the putative PGD₂ antagonist, acts as a thromboxane antagonist. This PGD₂-thromboxane interaction has also been observed in recent studies with the experimental compound AH 6809, which antagonized both PGD₂- and U-46619-induced responses in human PRP with pA₂ values of 5.4 and 4.4, respectively (Keery & Lumley, 1985). Clearly, however, not all thromboxane antagonists non-specifically antagonize PGD₂ receptors (Bennett & Sanger, 1982; Armstrong *et al.*, 1985) as confirmed in the present study. The full characterization of both *in vitro* and *in vivo* responses to PGD₂ in various tissues will necessarily require the use of both specific thromboxane- and PGD₂-receptor antagonists.

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