

Table S3 Plasmids used in this study

Plasmid	Characteristic	Primers for construction	Reference or Source
Plasmids used for the <i>B. subtilis</i> transformation			
pCH11	pUB110 <i>ori-ts</i> pBR322 <i>ori cat</i> (harboring nonsense mutation) (Km ^r Amp ^r)		(1)
pCA191	pUC19 derivative harboring <i>cat</i> (Cm ^r Amp ^r)		(2)
pCA191 <i>glpD</i> -FL	pCA191 derivative harboring full length of <i>glpD</i> (-288 to 1668) (Cm ^r)	<i>glpD</i> -N-F, <i>glpD</i> -C-R	This study
pCA191 <i>glpD</i> -N	pCA191 derivative harboring N-terminal fragment of <i>glpD</i> (-288 to 942) (Cm ^r)	<i>glpD</i> -N-F, <i>glpD</i> -N-R	This study
pCA191 <i>glpD</i> -C	pCA191 derivative harboring C-terminal fragment of <i>glpD</i> (820 to 1668) (Cm ^r)	<i>glpD</i> -C-F, <i>glpD</i> -C-R	This study
pMutinNC	Plasmid vector to construct knockout or conditional-null mutants of the <i>B. subtilis</i> genes (Em ^r Amp ^r)		(3)
pMutinNC <i>glpD</i>	pMutNC derivative harboring internal fragment of <i>glpD</i> (23 to 182) (Em ^r)	pMut- <i>glpD</i> -F, pMut- <i>glpD</i> -R	This study
pAPNC213	Plasmid vector to integrate a target gene fused with <i>Pspac</i> into the <i>B. subtilis aprE</i> locus (Spec ^r Amp ^r)		(3)
pDG780	pBluescript KS ⁺ harboring <i>kan</i> (Amp ^r Km ^r)		(4)
pAPNCK	pANNC213 derivative harboring <i>kan</i> (Km ^r Amp ^r)		This study
pAPNCK <i>plsX</i>	pAPNCK derivative harboring <i>plsX</i> (-22 to 1011) (Km ^r)	pAP- <i>plsX</i> -F, pAP- <i>plsX</i> -R	This study
Plasmids used for protein expression in <i>E. coli</i>			
pSTV28	pACYC184 derived cloning vector (Cm ^r)		Takara
pSTV29	pACYC184 derived cloning vector (Cm ^r)		Takara
pSTV29 <i>plsB</i>	pSTV29 derivative harboring <i>plsB</i> (-229 to 2428) (Cm ^r)	PLSB-F, PLSB-R	This study
pSTV28 <i>EcplsX</i>	pSTV29 derivative harboring <i>E.coli plsX</i> (-81 to 1045) (Cm ^r)	ECPLSX-F, ECPLSX-R	This study
pSTV29 <i>ygiH</i>	pSTV29 derivative harboring <i>ygiH</i> (-177 to 625) (Cm ^r)	YGIH-F, YGIH-R	This study
pSTV28 <i>BsplsX</i>	pSTV29 derivative harboring <i>B.subtilis plsX</i> (-22 to 1011) (Cm ^r)	BSPLSX-F, BSPLSX-R	This study
pSTV29 <i>yneS</i>	pSTV29 derivative harboring <i>yneS</i> (-129 to 584) (Cm ^r)	YNES-F, YNES-R	This study
pSTV28 <i>plsB-p</i>	pSTV28 derivative harboring <i>plsB</i> (-90 to 2424) (Cm ^r)	PLSB-p-F, PLSB-p-R	This study
pMW118	pSC101 derived cloning vector (Amp ^r)		Nippon Gene
pMW118 <i>ygiH</i>	pMW118 derivative harboring <i>ygiH</i> (-177 to 625) (Amp ^r)		This study
pMW118 <i>yneS</i>	pMW118 derivative harboring <i>yneS</i> (-129 to 584) (Amp ^r)		This study
pSTV29 <i>plsB26</i>	pSTV29 derivative harboring <i>plsB26</i> (-169 to 2488) (Cm ^r)	PLSB-F, PLSB-R	This study
pUC18	pMB1 derived cloning vector (Amp ^r)		Takara
pUC18S	pUC18 derivative harboring <i>spec</i> (Spec ^r)	Spec-F-AatII, Spec-R-Eam1105I	This study
pUC18 <i>StesA</i>	pUC18S derivative harboring <i>tesA</i> (-82 to 638) (Spec ^r)	TesA-FEcoRI, TesA-RBamHI	This study
Plasmids used for construction of IAA-inducible strain in <i>E. coli</i>			
pMC1403	pBR322 derived <i>lacZ</i> fusion vector (Amp ^r)		(5)
pMC1403Pw	pMC1403 derivative harboring the <i>trp</i> promoter region (<i>trpL</i> -105 to 120) (Amp ^r)	P _{trp} -FMunI, P _{trp} -REcoRI	This study
pMC1403Pw- <i>plsB</i>	pMC1403Pw derivative harboring <i>plsB</i> (-20 to 2424) (Amp ^r)	PLSB-F2, PLSB-R2	This study
pMC1403Pw- <i>EcplsX</i>	pMC1403Pw derivative harboring <i>E.coli plsX</i> (-81 to 1045) (Amp ^r)	ECPLSX-F2, EC-PLSX-R2	This study
pMC1403Pw- <i>ygiH</i>	pMC1403Pw derivative harboring <i>ygiH</i> (-16 to 625) (Amp ^r)	YGIH-F2, YGIH-R2	This study

References

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