Supporting materials

The analysis of ¹⁵N relaxation of free L11

The dipole-dipole coupling and chemical shift anisotropy are dominant relaxation mechanisms for an amide ¹⁵N spin at high field. Therefore, rate constants for spin-lattice relaxation (R_1), spin-spin relaxation ($R_{1\rho}$), and dipolar cross-relaxation ¹⁵N-[¹H] NOE are given by^{22,23,38,68}:

$$R_1 = \frac{d^2}{4} [J(\omega_H - \omega_N) + 3J(\omega_N) + 6J(\omega_H + \omega_N)] + c^2 J(\omega_N)$$
(1)

$$R_{2} = \frac{d^{2}}{8} (4J(0) + J(\omega_{H} - \omega_{N}) + 3J(\omega_{N}) + 6J(\omega_{H}) + 6J(\omega_{H} + \omega_{N})) + \frac{c^{2}}{6} (4J(0) + 3J(\omega_{N})) + R_{ex} (2)$$

¹⁵ N - {¹H}NOE = 1 +
$$\frac{d^2}{4R_1} \frac{\gamma_H}{\gamma_N} (6J(\omega_H + \omega_N) - J(\omega_H - \omega_N)),$$
 (3)

in which $d = (\mu_0 h \gamma_N \gamma_H / 8\pi^2) < r_{NH}^{-3} >$, $c = \omega_N \Delta \sigma_N / \sqrt{3}$, μ_0 is the permeability in vaccum; h is Planck's constant; γ_H and γ_N are the magnetic gy-ratios of ¹H and ¹⁵N, respectively; r_{NH} =1.02 Å is the nitrogen-proton bond length; ω_N and ω_H are the Larmor frequencies of ¹H and ¹⁵N, respectively; $\Delta \sigma_N$ =-160 ppm is the chemical shift anisotropy for ¹⁵N; and R_{ex} is the chemical exchange rate. Since we have used the 1 ms delay between the refocusing pulses in CPMG sequence, the significant dephasing of the signal may be possible only if the exchange rate exceeds approximately 1000 s⁻¹. Using the results from the rotational diffusion tensor calculation (D:=1.97 10⁻⁷ s⁻¹, D_f=1.27 10⁻⁷ s⁻¹, and $\theta = 0.63^\circ$; Table 2), we have analyzed the data of ¹⁵N relaxation using the spectral density function given by:²²

$$J(\omega) = \frac{2}{5} \left(\sum_{k=1}^{3} A_k \left(\frac{S^2 \tau_k}{1 + (\omega \tau_k)^2} \right) + \frac{(1 - S^2) \tau}{1 + (\omega \tau)^2} \right),$$
(4)

where $A_1=(3/2 \cos^2\theta-1/2)^2$; $A_2=3 \sin^2\theta \cos^2\theta$; and $A_3=3/4 \sin^4\theta$. θ is the angle the nitrogen-proton bond vector makes with the unique diffusion tensor axis, $\tau_1=(6 D_f)^{-1}$; $\tau_2=(D_1 + 5 D_f)^{-1}$; $\tau_3=(4 D_1 + 2 D_f)^{-1}$; $1/\tau = 1/\tau_c + 1/\tau_e \tau_c = (D_1 + 2 D_f)/3 \tau_e$ is the effective correlation time of the internal motion; and S is the generalized order parameter, which contains information about the spatial extent of nitrogen-proton bond motion. Since a maximum of three free parameters (two [S², τ_e] from equation (4) and one [R_{ex}] from equation (2)) are required to fit experimental data, we have used the model using S², τ_e and R_{ex} to fit the experimental data.

Figure S1. The structure of L11 based on the NOE and dihedral angle constraints. The L11-NTD (a) and the L11-CTD (b) are superimposed for the best fit using residues 5–66 and 75–140, excluding the RBL, respectively.

Figure S2. Relaxation parameters for free L11. The (a) R_1 , (b) R_2 , and (c) ¹⁵N-[¹H] NOE were measured at pH 6.5 and 30 °C using Bruker Avance 600. The error was estimated by duplicated the measurement separated by several days. The concentrations of ¹⁵N,²H-labeled L11 were 0.7 mM.

Figure S3. Model-free parameters for free L11. (a) Generalized order parameters (S^2); (b) the effective internal correlation time of the internal motion (τ_e); and (c) the chemical exchange contributions (R_{ex}) were determined from ¹⁵N relaxation measurements.

Figure S4. Comparison of the relative orientation between the L11-NTD and -CTD. (a) The solution structure of L11 is shown in ribbon drawing. (b) The L11-CTD and (c) the L11-NTD structures placed onto the rotational diffusion tensor frame were obtained by using R_2/R_1 ratios in each domain separately for the calculation of the rotational diffusion tensor.

Figure S5. Comparison of the solution structure of L11 (blue) and the polypeptide chain of L11-rRNA complex in the crystal form (gold). (a) The L11-NTD and (b) the L11-CTD are superimposed for the best fit using residues 8–66 and 75–139, excluding the RBL.

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Fig. S1



Fig. S2



Fig. S3





Fig. S4



Fig. S5