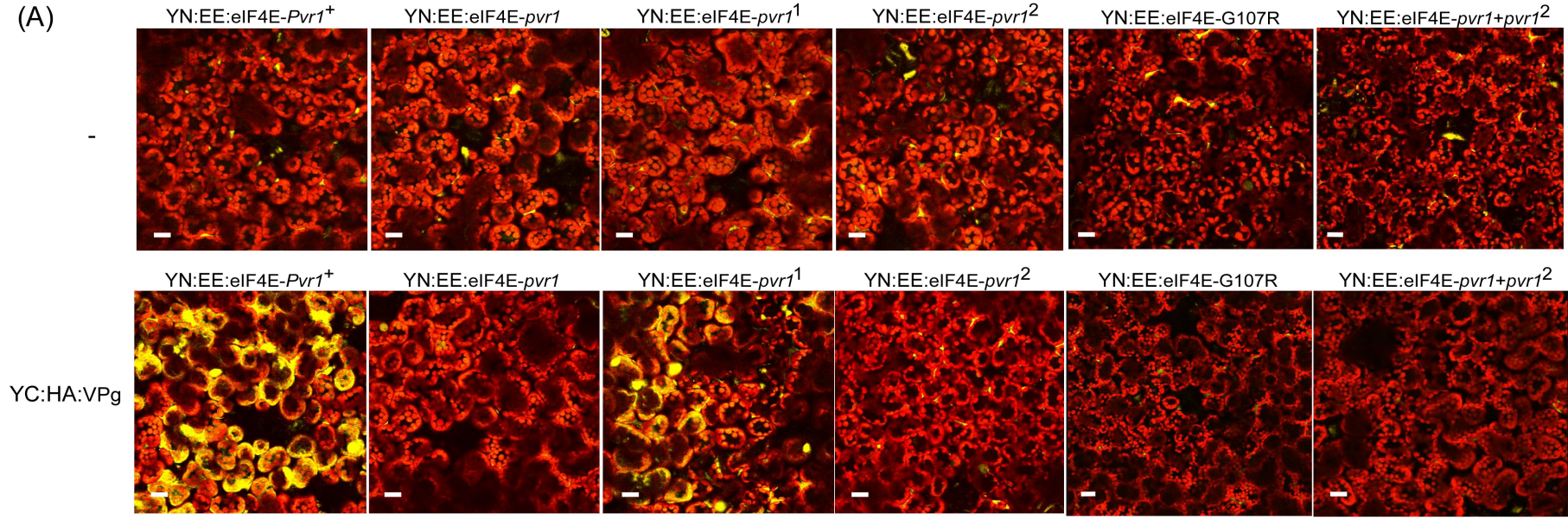
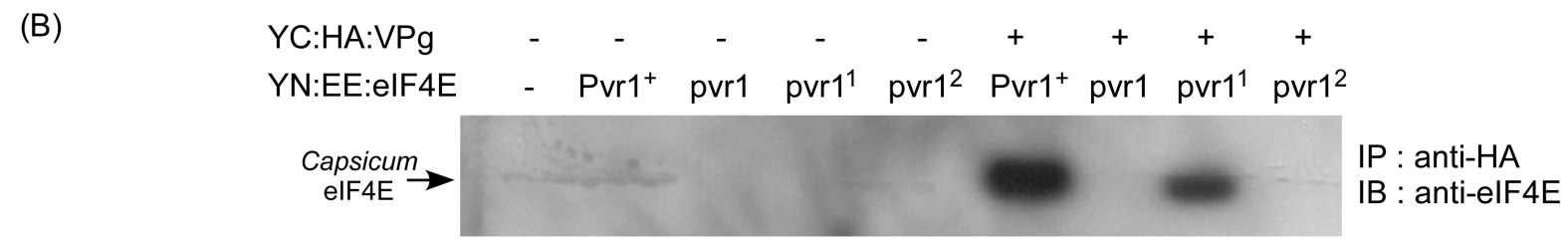


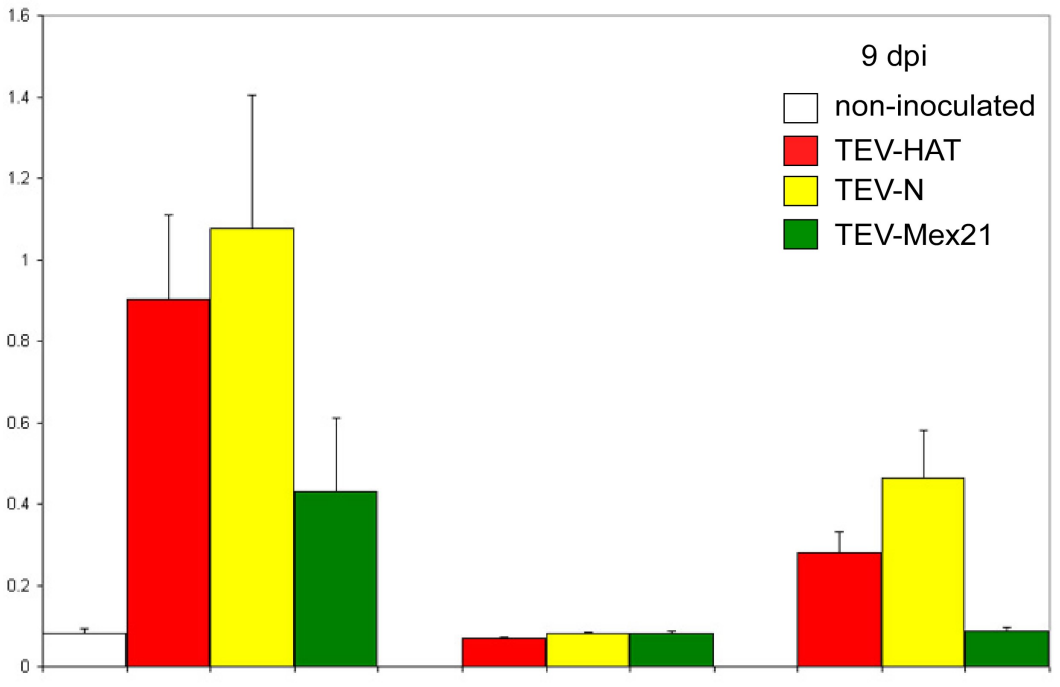


Supplemental Figure1

Confocal microscope images from bimolecular fluorescence complementation assay (BiFC) using Agrobacterium-mediated transient expression assay in *N. benthamiana*.

(A) eIF4E proteins fused with YN; YN:EE:eIF4E-*Pvr1*$^{+}$, YN:EE:eIF4E-*pvr1* YN:EE:eIF4E-*pvr1*^{1} YN:EE:eIF4E-*pvr1*^{2}, YN:EE:eIF4E-G107R, YN:EE:eIF4E-*pvr1*^{1}+*pvr1*^{2}, were transiently expressed in *N. bethamiana* leaf tissue with and without VPg protein fused with YC; YN:HA:VPg (TEV-HAT). Yellow fluorescent signal generated by the protein-protein interaction were detected in mesophyll cells 60 hours post-infiltration. Chloroplast autofluorescence is shown in red. Scale bar = 10 micron.

(B) Immunoblot image from coimmunoprecipitation assays. Total protein extracts were pulled down with anti-HA agarose beads and immunoblotted with an antibody for *Capsicum*-eIF4E.



Supplemental Figure 2

Accumulation of TEV coat protein determined by ELISA for transgenic tomatoes containing empty vector, eIF4E-*pvr1*, or eIF4E-*pvr1*^{1}.

ELISA was performed for T2 plants containing empty vector, eIF4E-*pvr1*, or eIF4E-*pvr1*^{1}, 9 days after inoculating with TEV-HAT, TEV-N and TEV-Mex21. Accumulation of TEV coat protein determined by ELISA of tissue sampled from upper un-inoculated leaves at 9 dpi.

Supplemental Table 1. Primer sequences used for site-directed mutagenesis.

Primer name	Sequence (5'→ 3')
T51Af	caa aga aat agc a**g**c aaa gca tcc at
T51Ar	at gga tgc ttt gct g**c**t att tct ttg
P66Tf	ctg gtt tga taa t**a**c agt ggc gaa atc
P66Tr	gat ttc gcc act g**t**a tta tca aac cag
G107Rf	gca agt tag ttg tg**a** gag cag act tac attg
G107Rr	caa tgt aag tct gct c**t**c aca act aac ttgc
V67Ef	ctg gtt tga taa tcc ag**a** ggc gaa atc gaaac
V67Er	gtt tcg att tcg cc**t** ctg gat tat caa accag
L79Rf	ggg tag ctc gc**g** tcg caa cgt cta cac
L79Rr	gtg tag acg ttg cga **c**gc gag cta ccc
D109Nf	gtt agt tgt ggg agc a**a**a ctt aca ttg
D109Nr	caa tgt aac t**t**t gct ccc aca act aac
G107R_D109Nf	gtt agt tgt g**a**g agc a**a**a ctt aca ttg
G107R_D109Nr	caa tgt aac t**t**t gct c**t**c aca act aac
P66T_L79Rf	ggt ttg ata at**a** cag **a**gg cga aat cg
P66T_L79Rr	cga ttt cgc c**t**c tgt **t**tt atc aaa cc

Supplemental Table 2. Primers sequences used for various plasmid construction.

Primer name	Primer sequence (5' → 3')	Cloning vector
VPgMex-xr	ccctcgagctattcaaacatcaactcct	pEG202/pJG4-5
VPgNW-xr	ccctcgagctattcaaacgtcaactcct	pEG202/pJG4-5
eIF4E-sf	ccgagctcatggcaacagctgaaatgg	pSY735/pSY736
eIF4E-br	tccggatccctatacggtgtaacg	pSY735/pSY736
VPgHAT-sf	gagctcatggggaagaagaatcagaa	pSY735/pSY736
VPgMex-sf	gagctcatggggaagaagaatcag aa	pSY735/pSY736
VPgMex-bsr	ggatccactattcaaacatcaactcct	pSY735/pSY736
VPgNNW-bsr	ggatccactattcaaacgtcaactcct	pSY735/pSY736
VPgHAT-bsr	ggatccactattcaaacgtcaagtcct	pSY735/pSY736
VPgHAT-BamHIeraserF	cattaggtttgtggacccattgacaggtcac	pSY735/pSY736
VPgHAT-BamHIeraserR	gtgacctgtcaatgggtccacaaacctaatg	pSY735/pSY736
VPgHAT-HindIIIeraserF	cagaagcacaagctgaagatgagagaggcg	pCAMBIA
VPgHAT-HindIIIeraserR	cgcctctctcatcttcagcttgtgcttctg	pCAMBIA
eIF4E_SmaI F	tcccccgggatggcaacagctgaaatgg	pBI121
eIF4E_SacIR	tccgagctcctatacggtgtaacg	pBI121