

References for Supplementary Material.

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Supplementary Table 1. Class I Mutations Identified In Study Cohort.

ID	Mutation		Mutation Effect	Exon	Domain	Rate	Ref.
	cDNA	Protein					
<u>PKD1 gene</u>							
Frameshift:							
JHU111	559delTTTAA	N116fsX	Stop Codon at 117	3	LRR2	1/164	N
JHU568	1124insCT	W305 fsX	Stop Codon at 334	5	PKDR1	1/164	N
JHU582	2291ins1	P694 fsX	Stop Codon at 713	10		1/164	N
JHU585	2297ins1	A696 fsX	Stop Codon at 713	10		1/164	N
JHU15	5225delAG	R1672 fsX	Stop Codon at 1721	15	PKDR11	1/164	[1-3]
JHU508	5365insT	V1718 fsX	Stop Codon at 1770	15	PKDR12	1/164	N
JHU613	6666insG	D2152 fsX	Stop Codon at 2174	15	REJ	1/164	N
JHU611	6881insA	P2224 fsX	Stop Codon at 2261	15	REJ	1/164	N
JHU577	8713delT	F2834 fsX	Stop Codon at 2874	23		1/164	N
JHU600	9134ins1	P2975 fsX	Stop Codon at 3068	24		1/164	N
JHU579	9536ins5	I3109 fsX	Stop Codon at 3317	26		2/164	N
JHU609	9536ins5	I3109 fsX	Stop Codon at 3317	26		2/164	N
JHU599	10239delT	L3343 fsX	Stop Codon at 3395	30	TM3	1/164	N
JHU104	11587delG	G3792 fsX	Stop Codon at 3824	40		1/164	[4]
Nonsense:							
JHU605	483 C>A	S91X	Stop Codon at 91	2	LRR1	1/164	N
JHU567	4517 C>T	R1436X	Stop Codon at 1436	15	PKDR8	1/164	N
JHU108	7006 C>A	Y2265X	Stop Codon at 2265	15	REJ	1/164	N
JHU563	7499 C>T	R2430X	Stop Codon at 2430	18	REJ	1/164	[5, 6]
JHU593	7877 C>T	Q2556X	Stop Codon at 2556	19	REJ	1/164	[7]
JHU083	8267 C>T	Q2686X	Stop Codon at 2686	22	REJ	1/164	N
JHU574	8639 G>T	E2810X	Stop Codon at 2810	23	REJ	1/164	N
JHU620	12243 G>A	W4011X	Stop Codon at 4011	44	TM9	1/164	N

Splicing:

JHU572		IVS4+1G>A.	Loss of donor site	4		1/164	N.
JHU580		IVS19+1G>T.	Loss of donor site	19	REJ	1/164	N.

PKD2 gene**Frameshift:**

JHU586	2226insA	720fsX	Stop Codon at 730	11		2/164	N
JHU116	2226insA	720fsX	Stop Codon at 730	11		2/164	N
JHU591	2422delAG	786fsX	Stop Codon at 793	12	CC	1/164	N

Nonsense:

JHU578	982 C>T	R306X	Stop Codon at 360	4	TM1	2/164	[8]
JHU583	982 C>T	R306X	Stop Codon at 360	4	TM1	2/164	[8]
JHU607	2224 C>T	R742X	Stop Codon at 742	11		1/164	[9]
JHU594	2680C>T	R872X	Stop Codon at 872	14		3/164	[10]
JHU566	2680C>T	R872X	Stop Codon at 872	14		3/164	[10]
JHU608	2680C>T	R872X	Stop Codon at 872	14		3/164	[10]

Splicing:

JHU562		IVS7-1G>A	Loss of acceptor site	7		1/164	N
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Rate=the number of times that the variant was observed in the study population (N=82, 164 chromosomes). Leucine Rich Repeat (LRR), Polycystic Kidney Disease Repeat # (PKD-R), Repector for Egg Jelly domain (REJ), Transmembrane (TM), Coiled Coil (CC), Novel change, identified in this study (N).

Supplementary Table 2. Analysis Of Splice Site Mutations.

ID	Mutation	Intron	SSPNN			ASSA					PKDx Class.	Rate	Ref.
			Initial	Final	Mutation	Initial	Final	Fold	% Binding	Mutation			
			Score	Score	Effect	(Ri)	(Ri)	Change	(Ri/Rf)	Effect			
<u>PKD1 gene</u>													
JHU572	IVS4+1G>A	4	0.97	N/A	Site abolished	9.5	-3.4	-699.5	N/A	Site abolished	Class I	1/164	N
JHU411	IVS10-4G>A	10	0.62	0.98	Strengthened	12.9	13	1.1	109.6	Strengthened	Polymorphism	1/164	N
JHU580	IVS19+1G>T	19	1	N/A	Site abolished	10.7	2.9	-223.6	N/A	Site abolished	Class I	1/164	N
JHU617	IVS20-16C>G	16	0.71	0.71	No change	3.4	3.4	1	100	No change	Polymorphism	1/164	N
JHU573	IVS24+5G>C	24	0.91	N/A	Site abolished	6.9	3	-15.5	6.4	Leaky site	Class II	2/164	N
JHU595	IVS24+5G>C	24	0.91	N/A	Site abolished	6.9	3	-15.5	6.4	Leaky site	Class II	2/164	N
JHU590	IVS24+28G>T	37	N/A	0.63	Site created	-1.1	7.1	138.4	13839.8	Site created	Class II	1/164	N
JHU569	IVS26+76C>A	26	0.69	0.69	No change	5.6	5.8	1.2	115.9	Strengthened	Polymorphism	1/164	N
JHU601	IVS37-4C>T	37	0.99	0.99	No change	4.9	4.9	0	100	No change	Polymorphism	1/164	N
JHU604	IVS37-10C>A	37	0.89	N/A	Site abolished	4.9	2.9	-4.1	24.5	Leaky site	Class II	1/164	[4]
<u>PKD2 gene</u>													
JHU559	IVS6-4C>T	6	0.92	0.92	No change	7.8	8.1	1.2	121.4	Strengthened	Polymorphism	1/164	N
JHU562	IVS7-1G>A	7	1	N/A	Site abolished	6.3	-1.3	-77.5	N/A	Site abolished	Class I	1/164	N
JHU105	IVS8+5G>A	8	0.83	N/A	Site abolished	7.8	4.3	-11.5	8.7	Leaky site	Class II	1/164	N

Intronic changes were analyzed using the splice site prediction programs SSPNN and ASSA (see text). Class I variants directly affect consensus splice donor/acceptor sites. As expected both programs predict that these changes will abolish splicing. Class II mutations are intronic changes that do not involve canonical splice sites but which are nonetheless predicted by both programs to alter the native splicing pattern. Initial (Ri): initial information content measured at the base before a mutation is made. Final (Rf): final information content measured at the base after a mutation is made. Fold change indicates the change in binding affinity of the two sites. Percent (%) Binding (Final/Initial), indicates the change of binding energy calculated as a percentage. If there is no change at the splice site then the fold change is 1.0. A positive fold change indicates that a new site is created while a negative change signifies that the site is abolished. (N), novel, variant identified in this study. Rate=the number of times that the variant was observed in the study population (N=82, 164 chromosomes).

Supplementary Table 3. Class II Mutations: In Frame Deletions/Insertions.

ID	Mutation		Exon Domain	Conservation				Rate	Ref	
	cDNA	Protein		Fr	Mm	Hs	LC			
<u>PKD1 gene</u>										
JHU115	514-551delCAA	N101del	3	LRR2	N	N	N	FC	1/164	N
JHU107	1848-1851delTGG	V546del	8		V	V	V	FC	1/164	N
JHU560	8892-8898del CCAACTCCG	ANS2894del	23		AGA	VGS	ANS	HC	1/164	N
JHU592	9905-9909delAAG	K3232del	28	PLAT	I	K	K	HC	1/164	N
JHU571	10070-10074delCTC	L3287del	29	TM2	L	L	L	FC	1/164	N
JHU112	12597-12600delTGG	V4129del	45		V	V	V	FC	1/164	N
<u>PKD2 gene</u>										
JHU596	374-378insTGG	103insV	1	Poly- Glu	-	V	V	HC	1/164	N
JHU416	1879-1882delTTC	F605del	8	TM5	-	F	F	HC	1/164	N

Leucline Rich Repeat-2 (LRR2), PLAT (Polycystin/Lipoxygenase/α-toxin), Poly-Glu (Poly-Glutamic acid region) TM2 (Transmembrane domain 2), *Fr* (*Fugu rubripes*), *Mm* (*Mus musculus*), *Hs* (*Homo Sapiens*), LC (Level of amino acid Conservation), FC (Fully Conserved amino acid) were those that were identical in all three species compared, while amino acids with similar properties (i.e. belonging to the same amino acid class) were deemed to be “highly” conserved (HC) residues. NC (amino acid residues that were Not Conserved), N (Novel change, identified in this study), Rate=the number of times that the variant was observed in the study population (N=82, 164 chromosomes).

Supplementary Table 4. Analysis of Class II and Class III Sequencing Results

ID	Amino acidic change											Total # of		
	Gene	cDNA	Protein	Location	Domain	M&K Matrix	Conservation			Rate	Family history	Ref	variants.	
							Fr	Mm	LC				PKD1	PKD2
<u>JHU612</u>	<i>PKD1</i>	1023C>A	A271D	Exon 5	WSC	Path.H.	A	A	FC	1/164	No	N	4	0
	<i>PKD1</i>	485A>G	A92T	Exon 2		Equal	F	A	HC	1/164		N		
<u>JHU602</u>	<i>PKD1</i>	1470A>G	Y420C	Exon 6	C-LECT ^a	Path.H.	F	Y	HC	1/164	No	N	25	1
	<i>PKD1</i>	4262C>T	R1351W	Exon 15	PKDR7 ^a	Path.H.	R	R	FC	1/164		N		
	<i>PKD1</i>	8855T>A	W2882R	Exon 23		Path.H.	G	Q	NC	1/164		N		
	<i>PKD1</i>	9109G>C	E2966D	Exon 24		Poly.H.	G	E	HC	1/164		[6, 11-13]		
<u>JHU103</u>	<i>PKD1</i>	1794A>G	Y528C	Exon 7	C-LECT ^a	Path.H.	Y	Y	FC	1/164	Yes	N	28	1
	<i>PKD1</i>	6036G>A	R1942H	Exon 15	PKDR14	Equal	R	R	FC	1/164		N		
<u>JHU001</u>	<i>PKD1</i>	2042C>T	R611W	Exon 9		Path.H.	R	R	FC	1/164	Yes	N	6	0
	<i>PKD1</i>	8651G>A	G2814R	Exon 23	REJ	Path.H.	A	G	HC	6/164		[2, 14]		
<u>JHU411</u>	<i>PKD1</i>	3351C>T	S1047L	Exon 13	PKDR4 ^a	Path.H.	M	S	HC	1/164	Yes	N	60	1
	<i>PKD1</i>	6756A>G	Q2182R	Exon 15	REJ	Path.H.	G	Q	HC	2/164		[2, 14]		
	<i>PKD2</i>	634G>A	A190T	Exon 1		Equal	-	A	HC	3/164 ^b		N		
<u>JHU100</u>	<i>PKD1</i>	5793C>T	T1861I	Exon 15	PKDR13 ^a	Path.H.	T	S	HC	1/164	Yes	N	8	1
	<i>PKD1</i>	6707C>T	R2166C	Exon 15	REJ	Path.H.	P	R	HC	1/164		N		
	<i>PKD1</i>	4229C>T	R1340W	Exon 15	PKDR6 ^a	Path.H.	H	H	FC	3/164		[2]		
<u>JHU564</u>	<i>PKD1</i>	10187G>C	G3326R	Exon 30	TM3	Path.H.	G	G	FC	1/164	Yes	N	4	1
	<i>PKD1</i>	7116C>G	A2302G	Exon 15	REJ	Equal	S	A	HC	1/164		N		
	<i>PKD1</i>	10311A>G	I3367V	Exon 31		Poly. H.	I	V	HC	1/164		N		
<u>JHU588</u>	<i>PKD1</i>	7554T>C	L2448P	Exon 18	REJ	Path.H.	L	L	FC	1/164	Yes	N	39	0
	<i>PKD1</i>	4229C>T	R1340W	Exon 15	PKDR6 ^a	Path.H.	H	H	HC	3/164		[2]		
<u>JHU603</u>	<i>PKD1</i>	7757C>T	R2516C	Exon 19	REJ	Path.H.	R	R	FC	1/164	Yes	N	4	0
<u>JHU569</u>	<i>PKD1</i>	8067T>C	L2619P	Exon 20	REJ	Path.H.	L	L	FC	1/164	No	N	22	2
	<i>PKD1</i>	8411C>A	P2734T	Exon 23	REJ	Equal	P	P	FC	1/164		[6]		
	<i>PKD1</i>	8415A>T	Q2735L	Exon 23	REJ	Equal	S	Q	HC	1/164		[6]		
<u>JHU597</u>	<i>PKD1</i>	8138C>T	R2643C	Exon 21	REJ	Path.H.	R	R	FC	1/164	No	N	3	1
<u>JHU101</u>	<i>PKD1</i>	8509C>T	R2767C	Exon 23	REJ	Path.H.	R	R	FC	1/164	Yes	N	4	2
<u>JHU109</u>	<i>PKD1</i>	8522G>A	E2771K	Exon 23	REJ	Path.H.	E	E	FC	1/164	Yes	[3, 6, 11]	3	0
<u>JHU589</u>	<i>PKD1</i>	8769T>C	F2853S	Exon 23		Path.H.	F	F	FC	1/164	Yes	[11, 15]	22	2
<u>JHU 576</u>	<i>PKD1</i>	10096C>A	N3295K	Exon 29	TM2	Path.H.	N	N	FC	1/164	Yes	N	3	0
<u>JHU114</u>	<i>PKD1</i>	12658C>T	R4149C	Exon 46		Path.H.	R	R	FC	1/164	Yes	N	22	1
	<i>PKD1</i>	4229C>T	R1340W	Exon 15	PKDR6 ^a	Path.H.	R	R	HC	3/164		[2]		

<u>JHU601B</u>	<u>PKD1</u>	<u>9258A>G</u>	<u>Q3016R</u>	<u>Exon 25</u>	<u>GPS</u>	<u>Path.H.</u>	<u>Q</u>	<u>Q</u>	<u>FC</u>	<u>1/164</u>	<u>Yes</u>	<u>[6, 11, 13, 15]</u>	43	1
	<i>PKD1</i>	2427A>G	Q739R	Exon 11		Path.H.	R	Q	HC	11/164 ^e		[16]		
<i>JHU565</i>	<i>PKD1</i>	7476C>A	T2422K	Exon 18	REJ	Equal	T	T	FC	1/164	Yes	N	24	0
	<i>PKD1</i>	3527C>G	L1106V	Exon 15	PKDR4	Poly.H.	S	V	NC	1/164		N		
	<i>PKD1</i>	3713C>T	P1168S	Exon 15	PKDR5	Equal	-	P	HC	2/164 ^d		[2]		
<i>JHU570</i>	<i>PKD1</i>	1947C>A	P579Q	Exon 9		Equal	P	P	FC	1/164	Yes	N	5	1
	<i>PKD1</i>	2427A>G	Q739R	Exon 11		Path.H.	R	Q	HC	11/164 ^e		[16]		
<i>JHU575</i>	<i>PKD1</i>	3312A>G	N1034S	Exon 13	PKDR4	Poly.H.	G	S	NC	1/164	No	N	17	1
<i>JHU178</i>	<i>PKD1</i>	3713C>T	P1168S	Exon 15	PKDR5	Equal	-	P	HC	2/164 ^d	Yes	[2]	2	1
	<i>PKD2</i>	634G>A	A190T	Exon 1		Equal	-	A	HC	3/164 ^b		N		
<i>JHU610</i>	<i>PKD2</i>	634G>A	A190T	Exon 1		Equal	-	A	HC	3/164 ^b	No	N	40	2
<i>JHU617</i>	<i>PKD1</i>	4391C>G	L1394V	Exon 15	PKDR8 ^a	Poly.H.	V	L	HC	1/164	Yes	N	5	1
	<i>PKD1</i>	11040T>A	L3730Q	Exon 39		Equal	F	L	HC	1/164		N		
<i>JHU587</i>	<i>PKD1</i>	840G>T	C210F	Exon 5		Equal	C	C	FC	1/164	No	N	6	2
	<i>PKD1</i>	7197G>A	R2329Q	Exon 16	REJ	Equal	E	R	HC	1/164		N		
	<i>PKD1</i>	2427A>G	Q739R	Exon 11		Path.H.	R	Q	HC	11/164 ^e		[16]		
<i>JHU559</i>	<i>PKD1</i>	351G>C	C47S	Exon 1	LRR-N	Poly.H.	W	C	HC	1/164	Yes	N	24	3
	<i>PKD2</i>	2464A>C	M800L	Exon 13		Poly.H.	-	M	HC	1/164		[17]		
<i>JHU606</i>	<i>PKD1</i>	6809C>T	R2200C	Exon 15	REJ	Path.H.	R	R	FC	4/164 ^d	No	[3]	5	2
<i>JHU584</i>	<i>PKD1</i>	6809C>T	R2200C	Exon 15	REJ	Path.H.	R	R	FC	4/164 ^d	No	[3]	20	1
<i>JHU106</i>	<i>PKD1</i>	8651G>A	G2814R	Exon 23	REJ	Path.H.	A	G	HC	6/164	Yes	[2, 6]	4	1
<i>JHU614</i>	<i>PKD1</i>	4757G>A	A1516T	Exon 15	PKDR9	Equal	T	I	NC	2/164	No	N	10	0
	<i>PKD1</i>	1973A>C	E586D	Exon 9		Poly.H.	A	E	HC	1/164		N		
	<i>PKD1</i>	2427A>G	Q739R	Exon 11		Path.H.	R	Q	HC	11/164 ^e		[16]		
<i>JHU102</i>											Yes		21	0
<i>JHU598</i>											No		19	0
<i>JHU616</i>											Yes		17	0
<i>JHU110</i>											Yes		3	0
<i>JHU113</i>											No		2	1
<i>JHU615</i>											No		0	1

Only study participants with at least one predicted pathogenic missense change are underlined. The amino acid substitutions that meet the criteria for pathogenicity are shadowed in grey. An amino acid substitution was deemed to be pathogenic if it i) occurred at a fully or highly conserved amino acid residue, if it ii) was also predicted to have a higher pathogenic potential using the matrix of Miller and Kumar and if iii) it was absent in normals. Key: Disrupts consensus sequence of the domain (see Figure 2). Found in 3.2 % (^b), 6.4 % (^c), 1.7%

(^d) and 1.4% (^d) of the normal population. Cell-wall integrity and Stress-response Component (WSC), C-type Lectin (C-LECT), Polycystic Kidney Disease Repeat # (PKD-R#), Receptor for Egg Jelly domain (REJ), Transmembrane # (TM#), GPCR proteolytic site (GPS), Leucine Rich Repeat (LRR), Novel change. LC (Level of amino acid Conservation), FC (Fully Conserved amino acid) were those that were identical in all three species compared, while amino acids with similar properties (i.e. belonging to the same amino acid class) were deemed to be “highly” conserved (HC) residues. NC (amino acid residues that were Not Conserved). Rate=the number of times that the variant was observed in the study population (N=82, 164 chromosomes).

Supplementary Table 5. Missense Variants In Participants With Truncating Mutations, In frame Deletions/Insertions And Predicted Splicing Mutations.

<i>Pedigree</i>	<i>Mutations</i>		<i>Amino acid changes</i>		<i># of Changes</i>	
	<i>disease associated</i>		<i>highly pathogenic</i>		<i>per patient</i>	
	<i>PKD1</i>	<i>PKD2</i>	<i>PKD1</i>	<i>PKD2</i>	<i>PKD1</i>	<i>PKD2</i>
JHU605	S91X				4	0
JHU 567	R1436X				24	0
JHU108	Y2265X		Q739R ^a		5	1
JHU563	R2430X		Q739R ^a	R807Q	19	1
JHU593	Q2556X				3	2
JHU083	Q2686X				5	0
JHU574	E2810X		G2814R		4	0
JHU620	W4011X				22	0
JHU568	W305 fsX		W305C ^b		4	1
JHU582	P694 fsX				1	0
JHU585	A696 fsX				6	2
JHU508	V1718 fsX				5	2
JHU613	D2152 fsX		E624K		21	0
JHU611	P2224 fsX				29	1
JHU600	P2975 fsX		Q739R ^a		25	2
JHU609	I3109 fsX				41	1
JHU579	I3109 fsX		G2814R		23	2
JHU577	F2834fsX		Q739R ^a		4	1

JHU111	N116 fsX	R2200C ^a	7	1
		S1619F		
JHU15	R1672 fsX		5	1
JHU599	L3343 fsX	<i>R1312Q</i>	20	1
JHU104	G3792 fsX		4	2
JHU115	N101del		20	0
JHU107	V546del	R1142W	25	1
JHU560	ANS2894del		21	1
JHU592	K3232del		3	1
JHU571	L3287del		10	0
JHU112	V4129del	S4053F	19	2
JHU580	IVS19+1G>T.	G2814R	5	1
JHU572	IVS4+1G>A	S349P	17	1
JHU573	IVS24+5G>C	R2200C ^a	5	0
JHU604	IVS37-10C>A	Q739R ^a	2	0
	IVS24+28G>			
JHU590	T		3	1
JHU595	IVS24+5G>C		16	2
JHU578		R306X	3	3
JHU583		R306X	5	1
JHU607		R742X	21	1
JHU594		R872X	22	3
JHU566		R872X	1	3

JHU608	R872X	G2814R	4	2
JHU596	103insV	Q2182R	35	1
JHU416	F605del		2	3
JHU591	786fsX		4	2
JHU562	IVS7-1G>A		3	2
	IVS8+5G>			
JHU105	A	T1773I ^b	3	2
JHU116	720 fsX	Q739R ^a	3	2
JHU586	720 fsX		22	1

Missense variants listed in the table are predicted to be pathogenic according to the Matrix of Miller and Kumar (see text). Some of these variants are found in normal controls (a) or disrupt a consensus sequence for a *PKDI* domain (b).

Supplementary Table 6. Polymorphisms Identified By Sequencing of PKD1 and PKD2

<i>ID#</i>	<i>Designation</i>	<i>cDNA</i>	<i>Location</i>	<i>Domain</i>	<i>Frequency</i>	<i>Ref.</i>
<i>Change (s)</i>						
<i>PKD1 Polymorphisms.</i>						
-	T263S(H)*	1004C>T	Exon 5		2/164	N
-	P572S(H)*	1925C>T	Exon 8		4/164	[2]
-	M1092T(H)*	3486T>C	Exon 14	PKD R4	30/164	[2]
-	W1399R(H)*	4406T>G	Exon 15	PKD R8	22/164	[1, 2, 18]
-	V1943I(H)*	6038G>A	Exon 15	PKD R14	5/164	[2]
-	E2548Q(H)*	7853G>C	Exon 19	REJ	4/164	[1]
-	H2638R(H)*	8124A>G	Exon 21	REJ	32/164	[1]
-	P2674S(H)*	8231C>T	Exon 21	REJ	2/164	[2, 6]
-	F3066L (H)*	9407T>C	Exon 25		38/164	[6, 19]
-	V3408L(H)*	10433G>C	Exon 33		5/164	N
-	A3511V(H)*	10743C>T	Exon 35		13/164	[2, 6]
-	I4044V(H)*	12341A>G	Exon 44	TM10	42/164	[2, 6, 12, 19, 20]
-	A4058V(H)*	12386C>T	Exon 45		12/164	[2, 12]
1	*	104C>T	Exon 1	5'UTR	1/164	N
2	*	145C>T	Exon 1	5'UTR	2/164	N
3	*	160C>T	Exon 1	5'UTR	1/164	N
4	*	210C>T	Exon 1	5'UTR	1/164	N
5	L72L	425C>T	Exon 1	LRR1	2/164	N
6	G109G	538A>T	Exon 3	LRR2	1/164	N
7	S196S	799C>T	Exon 5		2/164	N
8	A341A	1234C>T	Exon 5	PKD R1	5/164	[6]

9	L373L(H)	1330T>C	Exon 5		36/164	[2, 4, 6]
10	G441G	1534G>A	Exon 6	C-LECT	1/164	N
11	H570H	1921C>T	Exon 8		1/164	[2, 6]
12	IVS9+2del7*		Intron 9		12/164	N
13	IVS9+2T>A**		Intron 9		1/164	N
14	IVS9+28del7 (H)*		Intron 9		4/164	[2]
15	ISV9-44G>C*		Intron 9		1/164	[2]
16	IVS9-4A>G*		Intron 9		42/164	[2]
17	IVS10-4 G>A ^{S1047L}		Intron 10		1/164	N
18	P738P(H)	2425C>G	Exon 11		4/164	N
19	A745A	2448C>G	Exon 11		1/164	N
20	A898A	2905A>C	Exon 11	PKD R2	4/164	[2, 14]
21	P900P	2911G>A	Exon 11	PKD R2	10/164	[2, 18]
22	D910D	2941C>T	Exon 11	PKD R2	10/164	[2, 18]
23	IVS11-5C>T*		Intron 11		2/164	[2]
24	IVS11+23C>T(H)*		Intron 11		4/164	N
25	IVS12-15C>T*		Intron 12		5/164	N
26	G1021G(H)	3274T>C	Exon 13	PKD R4	30/164	[2, 18]
27	L1037L	3392A>G	Exon 13	PKD R4	15/164	[14]
28	E1061E	3394G>A	Exon 14	PKD R4	1/164	N
29	P1076P	3439G>A	Exon 14	PKD R4	1/164	N
30	A1124A	3583C>T	Exon 15	PKD R4	25/164	[2, 14]
31	S1125S	3586C>T	Exon 15	PKD R5	25/164	[2, 14]
32	F1163F	3700C>T	Exon 15	PKD R5	1/164	N
33	T1171T	3724C>G	Exon 15	PKD R5	1/164	N

34	D1310D	4141C>T	Exon 15	PKD R7	1/164	N
35	L1357L	4282G>T	Exon 15	PKD R7	1/164	N
36	S1373S	4330C>T	Exon 15	PKD R7	1/164	N
37	S1452S	4567T>C	Exon 15	PKD R8	1/164	N
38	P1511P	4744G>A	Exon 15	PKD R9	1/164	N
39	A1555A(H)	4876A>C	Exon 15		42/164	[1, 14, 18]
40	T1558T	4885G>A	Exon 15		9/164	[5]
41	S1603S	5020C>T	Exon 15		1/164	N
42	T1724T(H)	5383C>T	Exon 15	PKD R12	40/164	[2, 14, 16]
43	A1818A(H)	5665G>A	Exon 15	PKD R13	5/164	[2, 14]
44	G1860G	5791C>A	Exon 15	PKD R13	1/164	N
45	A1894A	5893C>T	Exon 15	PKD R14	1/164	[2, 14]
46	L1921L	5974G>A	Exon 15	PKD R14	2/164	[2, 14]
47	V2026V	6289C>T	Exon 15	PKD R15	1/164	N
48	R2121R	6574C>T	Exon 15	PKD R16	1/164	N
49	T2180T	6751C>T	Exon 15	REJ	1/164	N
50	A2202A	6817G>A	Exon 15	REJ	1/164	N
51	V2257V	6982G>A	Exon 15	REJ	1/164	N
52	G2309G	7138C>T	Exon 16	REJ	4/164	[2, 14]
53	IVS16+10 G>A*		Intron 16	REJ	1/164	N
54	R2359R	7289G>C	Exon 17	REJ	3/164	N
55	L2389L(H)	7376T>C	Exon 17	REJ	46/164	[1, 2, 5, 14]
56	G2425G	7486C>T	Exon 18	REJ	1/164	N
57	L2481L(H)	7652C>T	Exon 18	REJ	39/164	[1, 2]
58	IVS19+24 C>A*		Intron 19	REJ	2/164	N

59	L2570L(H)	7919T>C	Exon 20	REJ	31/164	[1, 14]
60	ISV20-16C>G ^{C47S, L3720Q}		Intron20	REJ	2/164	N
61	T2708M	8334C>T	Exon 22	REJ	1/164	[2, 6]
62	IVS22+8G>A(H)*		Intron 22	REJ	1/164	[1, 2]
63	S2729S	8398G>A	Exon 23	REJ	2/164	N
64	A2749A	8458G>A	Exon 23	REJ	1/164	N
65	S2766S	8509C>T	Exon 23	REJ	1/164	[21]
66	D2789D	8578C>T	Exon 23	REJ	2/164	N
67	S2813S	8650C>T	Exon 23	REJ	2/164	[2, 6, 11]
68	S2893S	8890C>G	Exon 23		2/164	[6]
69	A2971A(H)	9124T>C	Exon 24		2/164	N
70	IVS24-20G>A(H)*		Intron 24		3/164	N
71	IVS24-17A>G(H)*		Intron 24		6/164	N
72	IVS24+17A>G*		Intron 24		32/164	N
73	S3007S	9232C>T	Exon 25		1/164	N
74	V3065V(H)	9406G>C	Exon 25		38/164	[11]
75	V3090V	9481C>T	Exon 26	TM1	3/164	N
76	P3110P(H)	9543T>C	Exon 26		37/164	[15]
77	IVS26+76C>A ^{L2617P}		Intron26		1/164	N
78	IVS27-13T>C(H)*		Intron27		15/164	[6]
79	T3223T	9880G>A	Exon 28	PLAT	2/164	[2, 6, 15]
80	S3265S	10006C>T	Exon 29		1/164	N
81	IVS29-4C>T*		Intron29		1/164	N
82	A3455A	10576C>T	Exon 34		1/164	N
83	L3589L	10976C>T	Exon 36	TM5	5/164	[22]

84	IVS37-4C>T ^{Q3016R}		Intron 37		1/164	N
85	IVS38+11G>A*		Intron 38		4/164	N
86	R3752R	11385C>A	Exon 39	PC	1/164	N
87	L3753L	11465G>C	Exon 39	PC	1/164	N
88	IVS39-25del72bp*		Intron 39		1/164	[6, 23]
89	IVS41+3insGGG*		Intron 41		2/164	[2]
90	IVS41-11C>T*R2516C	C>T	Intron 41		2/164	N
91	S3893S(H)	11890C>T	Exon 42		3/164	[2]
92	IVS43+42C>A*		Intron 43		6/164	N
93	R3971R	12124C>T	Exon 43		3/164	N
94	L4025L	12286C>T	Exon 44		1/164	N
95	L4035L	12316C>T	Exon 44	TM10	1/164	N
96	IVS44+22delG*		Intron44		4/164	N
97	L4089L	12478C>G	Exon 45	TM11	1/164	N
98	A4091A(H)	12484A>G	Exon 45	TM11	43/164	[2, 6, 19, 23]
99	L4136L(H)	12617C>T	Exon 45		13/164	[2, 20]
100	V4152V	12667C>T	Exon 46		2/164	N
101	P4161P	12696C>A	Exon 46		1/164	N
102	S4189S	12778C>T	Exon 46		1/164	N
103	P4209P(H)	12838T>C	Exon 46		40/164	[2, 6]
104	L4221L	12874C>T	Exon 46	CC	1/164	N
105	A4255A	12978C>T	Exon 46		1/164	N
106		13135G>A	3'UTR		2/164	[2]

PKD2 Polymorphisms.

-	R28P(H)*	149C>T	Exon 1		50/164	[2, 12, 24]
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107	R60R	246G>A	Exon 1	1/164	N
108	G140G(H)	486G>A	Exon 1	22/164	N
109	IVS6-4C>T ^{C47S}		Intron 6	1/164	N
110	L539L	1683G>C	Exon 7	1/164	N

Polymorphisms were defined as sequence variants that were 1) not predicted to alter an amino acid, 2) intronic changes associated with truncating mutations, 3) polymorphisms reported by others, 4) missense changes associated with truncating mutations and 5) changes in the 3' UTR of unknown significance. Some polymorphisms were found in homozygosity (H). Polycystic Kidney Disease Repeat # (PKD-R#), Receptor for Egg Jelly domain (REJ), Transmembrane # (TM#), 5' Untranslated region (5'UTR), Leucine Rich Repeat# (LRR#), C-type Lectin (C-LECT), Cell-wall integrity and Stress-response Component (WSC), GPCR proteolytic site (GPS), Coiled Coil (CC), PM (Polycystin Motif), Novel change (N). Polymorphisms observed in patients with Class I change in PKD1 (*) or PKD2 (**), Class II missense amino acid substitutions (^{superscript}) are indicated. Rate=the number of times that the variant was observed in the study population (N=82, 164 chromosomes).