## A Protein Antigen of *Mycobacterium leprae* Is Related to a Family of Small Heat Shock Proteins

AUDUN H. NERLAND,<sup>1</sup><sup>†\*</sup> ABU SALIM MUSTAFA,<sup>1</sup><sup>‡</sup> DOUGLAS SWEETSER,<sup>2</sup> TORE GODAL,<sup>1</sup> AND RICHARD A. YOUNG<sup>2</sup>

Laboratory for Immunology, Institute for Cancer Research, The Norwegian Radium Hospital, Montebello, N-0310, Oslo 3, Norway,<sup>1</sup> and Whitehead Institute for Biomedical Research, Cambridge, Massachusetts 02142, and Department of Biology, Massachusetts Institute of Technology, Cambridge, Massachusetts 02139<sup>2</sup>

Received 29 March 1988/Accepted 22 August 1988

The gene encoding an immunologically important 18-kilodalton protein antigen of *Mycobacterium leprae* has been sequenced, and the amino acid sequence of the antigen has been deduced. The 18-kilodalton antigen is strikingly similar in size and sequence to a family of eucaryotic heat shock proteins.

Six protein antigens have thus far been implicated in the immune response to Mycobacterium leprae (2, 3). These proteins have estimated molecular masses of 70, 65, 36, 28, 18, and 12 kilodaltons (kDa). Most of these proteins have

clones were isolated from healthy vaccinated volunteers (9). The identity and cellular function of the 18-kDa protein is not known.

To obtain clues to the nature of the 18-kDa antigen, the

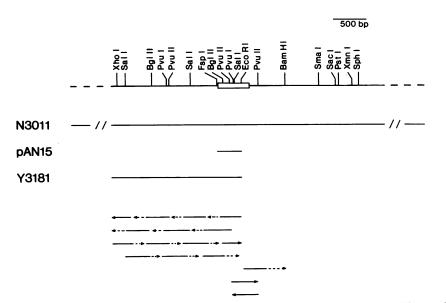


FIG. 1. Restriction maps of DNA encoding the *M. leprae* 18-kDa antigen and the DNA sequence strategy. The restriction map of *M. leprae* DNA containing the coding sequence for the 18-kDa antigen (boxed) is shown. The extents of insert DNA at clones N3011, Y3181, and pAN15, as well as the sequenced fragments, are indicated below the restriction map. bp, Base pairs.

been shown to stimulate human T cells (6, 8a, 9, 10, 12) and are therefore likely to be involved in cell-mediated immunity, an important determinant in protection against leprosy. The 18-kDa protein was the first of these antigens demonstrated to stimulate M. leprae-specific T-cell clones; these gene that encodes it was isolated and sequenced. A portion of the gene encoding the 18-kDa protein was isolated previously by probing a  $\lambda$ gtl1 recombinant DNA library with monoclonal antibodies directed against the *M. leprae* protein (17). This recombinant clone, Y3181, expressed a portion of the 18-kDa protein as a fusion protein with  $\beta$ -galactosidase. To obtain the sequence of the entire region encoding the 18-kDa protein, we sequenced the insert DNA from Y3181 as well as a portion of the insert DNA from a  $\lambda$ EMBL clone containing the entire gene encoding the 18-kDa protein (Fig. 1). The  $\lambda$ EMBL clone N3011 was isolated from a  $\lambda$ EMBL library of *M. leprae* DNA by probing with a plasmid containing a portion of the insert DNA of clone Y3181 (pAN15),

<sup>\*</sup> Corresponding author.

<sup>†</sup> Present address: Center for Biotechnology, University of Bergen, P.b. 3152, Aarstad, 5001 Bergen, Norway.

<sup>‡</sup> Present address: Whitehead Institute for Biomedical Research, Cambridge, MA 02142.

<sup>§</sup> Present address: World Health Organization, 1211 Geneva 27, Switzerland.

			10			20			30	)			40			50			
AGC	алт	стс	стс	AGC	tgt	тса	GAC	λGλ	<b>7</b> 77	стт	GTC	тат	слс	yyc	TTG	САТ	CVY	тат	
60	)			70			80			90	)		:	100			110		
* ATC	GAC	CAG	TGC	* TAT	ATC	ллл	тст	ATG	TAG	* TCA	GGA	аса	GCT	* АТА	TAG	тта	* TAG	TTT	
	120	, ,			130			140			150	,			160			170	
GTC	*		GAT		*	GCG	AGG	*	CCA	CAC			ATG		*	GAC	ccc	*	
													Met						
	180			190 *				200			210			22			20 *		
						GCC Ala													
230		240			0 2			50			260			270			280		
ATG	ccc	ATG Met	GAC	GCT	TGG	CGT Arg	GAG	GGC	GAA Glu	GAA Glu	TTC Phe	GTC Val	GTC Val	GAG Glu	TTC Phe	GAC	CTT Leu	CCT Pro	
	290			30	-	,		310			320			33				340	
~~~~			~~~	*		CTG		*	<b>CA</b> C	100		œc		*		200	CTC	*	
						Leu													
	350			360			370				380			390					
GCC	GAG	ccc	CCA	GGC	GTC	GAC Asp	ccc	GAT	CGG	GAA	ATG	CTT	GCT	GCC	GAG	CGG	CCA	OGC	
400	GIU	πy	410	-	vai	42		nap		430			440			45		,	
*			*				-						*				-		
GGT Gly	GTG Val	TTC Phe	λλT Asn	Arg	Gln	CTG Leu	GTT Val	Leu	GGC	GAA	λAC λsn	Leu	GAC Asp	Thr	GAA	Arg	Ile	Leu	
460			470 4				80 490				500 *			510					
GCT	TCC	TAC	CAA	GAA	GGT	GTC Val	CTG	AAG	TTG	TCG	ATA	CCA		GCC	GAN	AGG	GCT	LVS	
AIG		1y1 520	GIN	GIU	530		DBU	54				550		100	560			570	
		+			*		~~~								*			*	
Pro	Arg	Lys	Ile	Ser	Val	Asp	Arg	GGC	λsr	λsr	Gly	His	Glr	Thr	Ile	Asn	Lys	ACC	
	580				590			600			610			620			,		
						GCC Ala			NC1	GT	GTI	TGC	GCI	VC)	AGO	: TĀA	•		

FIG. 2. Sequence of the gene encoding the M. leprae 18-kDa antigen. Sequences were determined in both directions by the chain termination method (13). The deduced amino acid sequence is shown below the DNA sequence.

using standard procedures (7). Clone N3011 contained 17 kb of inserted DNA that included all of the DNA contained within the Y3181 insert DNA (Fig. 1).

Because Y3181 produces a fusion protein containing portions of the 18-kDa protein, the location and orientation of the gene encoding this protein could be estimated in the

60

overlapping restriction maps of clones Y3181 and N3011 (Fig. 1). DNA fragments isolated from clones Y3181 and N3011 were inserted into M13 vectors (8) and subjected to sequence analysis (13). The entire 1.7-kb *Eco*RI insert DNA of Y3181, a 0.4-kb *Pvu*II fragment of N3011 that overlaps the right end of the Y3181 insert DNA, and the 0.6-kb *Eco*RI-*Bam*HI fragment of N3011 that abuts the right end of the Y3181 insert DNA were sequenced (Fig. 1).

Figure 2 shows the DNA sequence obtained for the gene encoding the 18-kDa protein and the amino acid sequence predicted for the 18-kDa protein. The sequence predicts a protein of 148 amino acids with a molecular mass of 16,607 daltons, in good agreement with the molecular weight estimated on sodium dodecyl sulfate-polyacrylamide gels. For the sake of consistency, we will continue to refer to this protein by its apparent molecular mass of 18 kDa.

The amino acid sequence deduced for this antigen was compared with that of proteins in the National Biomedical Research Foundation protein sequence data base. The M. leprae antigen exhibited striking sequence similarity to a family of 17-kDa soybean heat shock proteins (11) (Fig. 3). A comparison of the M. leprae protein sequence with that of the soybean 17.5E protein revealed 31% amino acid sequence identity in 127 overlapping amino acids. A substantial fraction of the nonidentical amino acids were conserved amino acid replacements. Thus, the 18-kDa M. leprae protein antigen is significantly similar in sequence and size to the 17-kDa soybean heat shock proteins, which are representative of a class of small plant stress proteins (11). Moreover, these small plant heat shock proteins exhibit sequence similarities with a class of stress proteins present in animals, known as the small heat shock proteins (5, 11). We postulate that the 18-kDa M. leprae protein antigen is a stress protein.

It would be useful to determine directly whether specific stresses would induce increased synthesis of the *M. leprae* 18-kDa protein. However, *M. leprae* has not yet been successfully cultivated in vitro, which precludes direct analysis of the stress response in this organism.

Stress proteins have been identified as targets of the immune response for a variety of pathogens, including

<u>M</u> . <u>leprae</u> Soybean	MLMRTDPFRELDLRRASVSTSARPAVMPMDAWREG-EEFVV MSLIPGFFGGRRSNVFDPFSLDMWDPFKDFHVPTSSVSAENSAFVSTRVDWKETPEAHVF *** ** ***** - ** *-*-* -*
	120
<u>M</u> . <u>leprae</u> Soybean	EFDLPGIKADSLDIDIER-NVVTVRAERPGVDPDREMLAAERPRGVFNRQLVLGENLD KADIPGLKKEEVKVEIEDDRVLQISGERNVEKEDKNDTWHRVERSSGKFTRRFRLPENAK * -**- ****** *-* *** * -* * ***** *
<u>M</u> . <u>leprae</u> Soybean	TERILASYQEGVLKLSIPVAERAKPRKISVDRGNNGHQTINKTAHEIIDA VNEVKASMENGVLTVTVPKEEVKKPDVLAIEISG **** ******- * ****

FIG. 3. Comparison of the amino acid sequences of the *M. leprae* 18-kDa antigen and the soybean 17-kDa heat shock protein 17.5E (11). The sequence of the 18-kDa *M. leprae* antigen (top line) is aligned with that of the 17-kDa soybean heat shock protein (bottom line). Symbols: -, identical residues; \*, conserved amino acid replacements.

mycobacteria (14, 16), *Coxiella* spp. (15), plasmodia (1), schistosomes (4), and filaria (13a). This fact suggests that the stress response may be a natural response of the infectious pathogen to the hostile environment of the host. The observation that stress proteins are common immune targets of pathogens may reflect the fact that the stress response and the proteins induced by stress are highly conserved through procaryotes and eucaryotes.

This research was supported by grants from the National Institutes of Health (Public Health Service AI23545), the World Health Organization Programme for Vaccine Development, and the World Health Organization/World Bank/United Nations Development Program Special Program for Research and Training in Tropical Diseases. R. A. Y. is a Burroughs Wellcome scholar in molecular parasitology.

## LITERATURE CITED

- Bianco, A. E., J. M. Favaloro, T. R. Burkot, J. G. Culvenor, P. E. Crewther, G. V. Brown, R. F. Anders, R. L. Coppel, and J. Kemp. 1986. A repetitive antigen of *Plasmodium falciparum* that ic homologous to heat shock protein 70 of *Drosophila melanogaster*. Proc. Natl. Acad. Sci. USA 83:8713-8717.
- Britton, W. J., R. J. Garsia, L. Hellqvist, J. D. Watson, and A. Basten. 1986. The characterization and immunoreactivity of a 70 Kd protein common to *Mycobacterium leprae* and *Mycobacterium bovis* (BCG). Lepr. Rev. 57(Suppl. 2):67–75.
- 3. Engers, H. D., B. R. Bloom, and T. Godal. 1984. Monoclonal antibodies against mycobacterial antigens. Immunol. Today 6: 342–348.
- Hedstrom, R., J. Culpepper, R. A. Harrison, N. Agabian, and G. Newport. 1987. A major immunogen in *Schistosoma mansoni* infections is homologous to the heat shock protein HSP70. J. Exp. Med. 165:1430-1435.
- 5. Ingolia, T. D., and E. A. Craig. 1982. Four small *Drosophila* heat shock proteins are related to each other and to mammalian  $\alpha$ -crystallin. Proc. Natl. Acad. Sci. USA 79:2360–2364.
- Lamb, J. R., J. Ivanyi, A. D. M. Rees, J. R. Rothbard, K. Howland, R. A. Young, and D. B. Young. 1987. Mapping of T cell epitopes using recombinant antigens and synthetic peptides. EMBO J. 6:1245–1249.
- 7. Maniatis, T., E. F. Fritsch, and J. Sambrook. 1982. Molecular cloning: a laboratory manual. Cold Spring Harbor Laboratory,

NOTES

5921

Cold Spring Harbor, N. Y.

- Messing, J. 1983. New M13 vectors for cloning. Methods Enzymol. 101:20-78.
- 8a.Mustafa, A. S. 1988. Identification of T cell activating recombinant antigens shared between three candidate anti-leprosy vaccines, killed *M. leprae*, *Mycobacterium bovis* BCG and *Mycobacterium w.* Int. J. Leprosy 56:265–273.
- Mustafa, A. S., H. K. Gill, A. H. Nerland, W. J. Britton, V. Mehra, B. R. Bloom, R. A. Young, and T. Godal. 1986. Human T cell clones recognize a major *M. leprae* protein antigen expressed in *E. coli*. Nature (London) 319:63-66.
- Mustafa, A. S., F. Oftung, H. K. Gill, and I. Natvig. 1986. Characteristics of human T-cell clones from BCG and killed M. *leprae* vaccinated subjects and tuberculosis patients. Recognition of recombinant mycobacterial antigens. Lepr. Rev. 57(Suppl, 2):123-130.
- Nagao, R. T., E. Czarnecka, W. B. Gurley, F. Schoffl, and J. L. Key. 1985. Genes for low-molecular-weight heat shock proteins of soybeans: sequence analysis of a multigene family. Mol. Cell. Biol. 5:3417–3428.
- Offenhoff, T. H. M., P. R. Klaster, J. Ivanyi, B. G. Elferink, M. Y. L. DeWit, and R. R. P. DeVries. 1986. Mycobacterium leprae specific protein antigens defined by cloned human helper T cells. Nature (London) 319:66-68.
- Sanger, F., S. Nicklen, and A. R. Coulson. 1977. DNA sequencing with chain termination inhibitors. Proc. Natl. Acad. Sci. USA 74:5463-5467.
- 13a.Selkirk, M. E., P. J. Rutherford, D. A. Danham, F. Partono, and R. M. Maizels. 1987. Cloned antigen genes of *Brugia filarial* parasite. Biochem. Soc. Symp. 53:91–102.
- 14. Shinnick, T. M., M. H. Vodkin, and J. C. Williams. 1988. The Mycobacterium tuberculosis 65-kilodalton antigen is a heat shock protein which corresponds to common antigen and to the Escherichia coli GroEL protein. Infect. Immun. 56:446–451.
- 15. Vodkin, M. H., and J. C. Williams. 1988. A heat shock operon in *Coxiella burnetii* produces a major antigen homologous to a protein both in mycobacteria and *Escherichia coli*. J. Bacteriol. 170:1227–1234.
- Young, D., R. Lathigra, R. Hendrix, D. Sweetser, and R. A. Young. 1988. Stress proteins are common targets in leprosy and tuberculosis. Proc. Natl. Acad. Sci. USA 85:4267-4270.
- 17. Young, R. A., V. Mehra, D. Sweetser, T. M. Buchanan, J. Clark-Curtiss, R. W. Davis, and B. R. Bloom. 1985. Genes for the major protein antigens of the leprosy parasite *Mycobacterium leprae*. Nature (London) **316**:450-452.