

Table S1: Mutants isolated from the qualitative screen. Ten mutants were isolated and identified as having less dead cells than WT on solid media. The mutant name represents the position of the mutant in the ordered library. Tn10 insertion locations were determined by arbitrary PCR and sequencing (see materials and methods). The functions of the genes affected by the Tn10 insertion are from the SWISSPROT database. ND= not determined.

Mutant name	Tn10 insertion (location)	Function of gene affected by Tn10 insertion	Ratio percentage of dead cells WT/mutant In LB 30°C	Ratio percentage of dead cells WT/mutant in MM 30°C
44D3	<i>rssB</i> 27'79	negative regulator of σ^S	3.3 (+/- 0.03)	3.7 (+/- 0.04)
46E3	<i>znuC</i> , <i>znuB</i> 41'84	high affinity ABC transport system for Zn	ND	ND
47B3	<i>yiaT</i> 80'81	potential outer membrane protein (similar to <i>mipA</i> family)	ND	ND
47E4	<i>b2881</i> 65'08	unknown function	1.64 (+/- 0.04)	1.90 (+/- 0.03)
48H10	<i>yhaB</i> 70'39	unknown function	1.12 (+/- 0.02)	1.95 (+/- 0.14)
59H8	<i>yieC</i> 84'03	potential outer membrane protein (similar to <i>lamB</i> family)	0.92 (+/- 0.02)	1.49 (+/- 0.07)
65D2	<i>yjdK</i> 93'77	unknown function	2.21 (+/- 0.03)	1.88 (+/- 0.15)
64D10	<i>yfdV</i> 53.61	potential inner membrane protein	1.08 (+/- 0.08)	1.27 (+/ 0.15)
66G11	<i>ykgG</i> 6'	unknown function	1.32 (+/- 0.06)	1.3 (+/- 0.3)
70B5	<i>fliA</i> 43'09	σ^F (motility and flagellar synthesis)	2.5 (+/- 0.04)	6.5 (+/- 0.02)

Table S2: Strains used in this study. All strains are derived from *E. coli* K-12 (MG1655).

Strain name	Relevant genotype	Source/ reference
NEC283	$\Delta fliA$ FRT	This work
NEC284	$\Delta flhD$ FRT	This work
NEC285	$\Delta flgM$ FRT	This work
NEC286	$\Delta fliC$ FRT	This work
NEC287	$\Delta flgJ$ FRT	Allele from Keio collection
NEC288	$\Delta motA$ FRT	Allele from Keio collection
NEC289	<i>motA448</i> <i>zea</i> ::Tn10	(Samuel and Berg, 1996)
NEC290	$\Delta rssB$ FRT	This work
NEC291	<i>rpoS</i> ::Tn10	(Bjedov <i>et al.</i> , 2003)
NEC292	$\Delta rssB$ <i>rpoS</i> ::Tn10	This work
NEC293	$\Delta fliA$ FRT $\Delta rssB$ FRT	This work
NEC294	<i>rpsL</i> (<i>StrepR</i>)	(Giraud <i>et al.</i> , 2001)
NEC295	$\Delta fliA$ <i>rpsL</i>	This work
NEC296	$\Delta rssB$ <i>rpsL</i>	This work
NEC297	<i>gyrA</i> (<i>nalR</i>)	(Giraud <i>et al.</i> , 2001)
NEC298	$\Delta fliA$ FRT <i>gyrA</i>	This work
NEC299	$\Delta rssB$ FRT <i>gyrA</i>	This work
NEC300	<i>ypdA</i> :: <i>Patet</i> -GFP ⁺⁺	This work
NEC301	$\Delta fliA$ <i>ypdA</i> :: <i>Patet</i> -GFP ⁺⁺	This work
NEC302	$\Delta rssB$ <i>ypdA</i> :: <i>Patet</i> -GFP ⁺⁺	This work
NEC303	<i>Patet</i> -mRFP	This work
NEC304	$\Delta fliA$ <i>ypdA</i> :: <i>Patet</i> -mRFP	This work
NEC305	$\Delta rssB$ <i>ypdA</i> :: <i>Patet</i> -mRFP	This work
NEC306	<i>PibpAB</i> -mRFP ; <i>intC</i> :: <i>P2rrnB</i> -CFP ⁺⁺ ; <i>PyiaG</i> -YFP ⁺⁺	This work
NEC307	$\Delta rssB$ <i>PibpAB</i> -mRFP ; <i>intC</i> :: <i>P2rrnB</i> -CFP ⁺⁺ ; <i>PyiaG</i> -YFP ⁺⁺	This work
NEC308	$\Delta fliA$ <i>PibpAB</i> -mRFP ; <i>intC</i> - <i>P2rrnB</i> ::CFP ⁺⁺ ; <i>PyiaG</i> -YFP ⁺⁺	This work

Table S3: Primers used in this study. Underlined portions of the primers for deletion correspond to the sequence for Cm cassette amplification.

Primer name	Sequences (5'-3')
<i>rssB</i> delfor	atgacgcagccattggtcggaaaacagattctcattggt <u>ggtgtaggctggagctgcttc</u>
<i>rssB</i> delback	tcattctgcagacaacatcaagcgcagtcgaccaccgggt <u>catatgaatatcctccttag</u>
<i>rssB</i> checkfor	gcagtcgtaacccaatttc
<i>rssB</i> checkback	ggcaacatcctggttctta
<i>fliA</i> delfor	actcctggtagtcaaagttaaagtgcggcatttactgacg <u>gtgtaggctggagctgcttc</u>
<i>fliA</i> delback	taatcatgccgataactcatataacgcagggctgtttatc <u>catatgaatatcctccttag</u>
<i>fliA</i> checkfor	aacaactccggctacatctt
<i>fliA</i> checkback	gcttatcaggcctacaagt
<i>flgM</i> delfor	cagtactctgcaagtcttgctgctgcttgatcagcgcacgcgc <u>gtgtaggctggagctgcttc</u>
<i>flgM</i> delback	gagtattgatcgcacttcgctctgaagcctgtaagcaccgttcaaccgc <u>catatgaatatcctccttag</u>
<i>flgM</i> checkfor	tgaacctggcgctttattggtc
<i>flgM</i> checkback	gcgctaaggctcatctggtat
<i>flhD</i> delfor	caggcccttttctgcgagcgccttctcaggctgattaacatc <u>gtgtaggctggagctgcttc</u>
<i>flhD</i> delback	gtgggaataatgcatactccgagttgctgaaacac <u>catatgaatatcctccttag</u>
<i>flhD</i> checkfor	caaacgccggaagtgagtc
<i>flhD</i> checkback	caaccggtagcaccaga
<i>fliC</i> up	atggcacaagtcattaataccaacagcctctcgctgatca <u>tgtgtaggctggagctgcttc</u>
<i>fliC</i> dwn	ttaaccctgcagcagagacagaacctgctgcggtacctgg <u>catatgaatatcctccttag</u>
<i>fliC</i> checkfor	cctgacccgactcccagc
<i>fliC</i> checkback	tcggacgattagtgggtg
ARB1	ggccacgcgtcgactagttacNNNNNNNNNNgat
ARB2	ggccacgcgtcgacttagttac
ARB6	ggccacgcgtcgactagttacNNNNNNNNNNacgcc
Tn10ext	tggaaggaacgtcaattccc
Tn10int	ccattgctgttgacaaaggg

References

- Bjedov, I., Tenaillon, O., Gerard, B., Souza, V., Denamur, E., Radman, M., Taddei, F., and Matic, I. (2003) Stress-induced mutagenesis in bacteria. *Science* **300**: 1404-1409.
- Giraud, A., Matic, I., Tenaillon, O., Clara, A., Radman, M., Fons, M., and Taddei, F. (2001) Costs and benefits of high mutation rates: adaptive evolution of bacteria in the mouse gut. *Science* **291**: 2606-2608.
- Samuel, A.D., and Berg, H.C. (1996) Torque-generating units of the bacterial flagellar motor step independently. *Biophys J* **71**: 918-923.