

To investigate the extent to which the changes in inactivation are coupled to changes in channel activation, we examined the kinetics of channel activation and inactivation at +20 mV.

METHODS

The time course of inactivation was fitted by a simple fitting procedure in pClamp 6.0 (Axon Instruments, Inc.). From the time point where the rate of current decay by visual inspection was judged to be maximum, the current ( $I_{Na}(t)$ ) was fitted by the expression:

$$I_{Na}(t) = I_1(\exp(-t/\tau_1)) + I_2(\exp(-t/\tau_2)) + I_3,$$

where  $\tau_1$  and  $\tau_2$  are the inactivation time constants.  $I_1$  and  $I_2$  are the corresponding amplitudes, and  $I_3$  signifies noninactivating current and noise. (The time course of inactivation was best fit by a sum of two exponential distributions; but the fit of the steady state inactivation was not improved when fitted as a three-state process [not depicted]). Following Sarkar, S.N., A. Adhikari, and S.K. Sikdar. 1995. *J. Physiol.* 488:633–645, the time course of current activation ( $I'_{Na}(t)$ ) was determined by transforming the measured currents as:

$$I'_{Na}(t) = I_{Na}(t)/I_1(\exp(-t/\tau_1)) + I_2(\exp(-t/\tau_2) + I_3),$$

using the values for  $I_1$ ,  $\tau_1$ ,  $I_2$ ,  $\tau_2$ , and  $I_3$  obtained from the fit to the inactivation time course. Subsequently,  $I'_{Na}(t)$  was fitted with the expression:

$$I'_{Na}(t) = 1 - \exp(-(t - k)/\tau_a)^3,$$

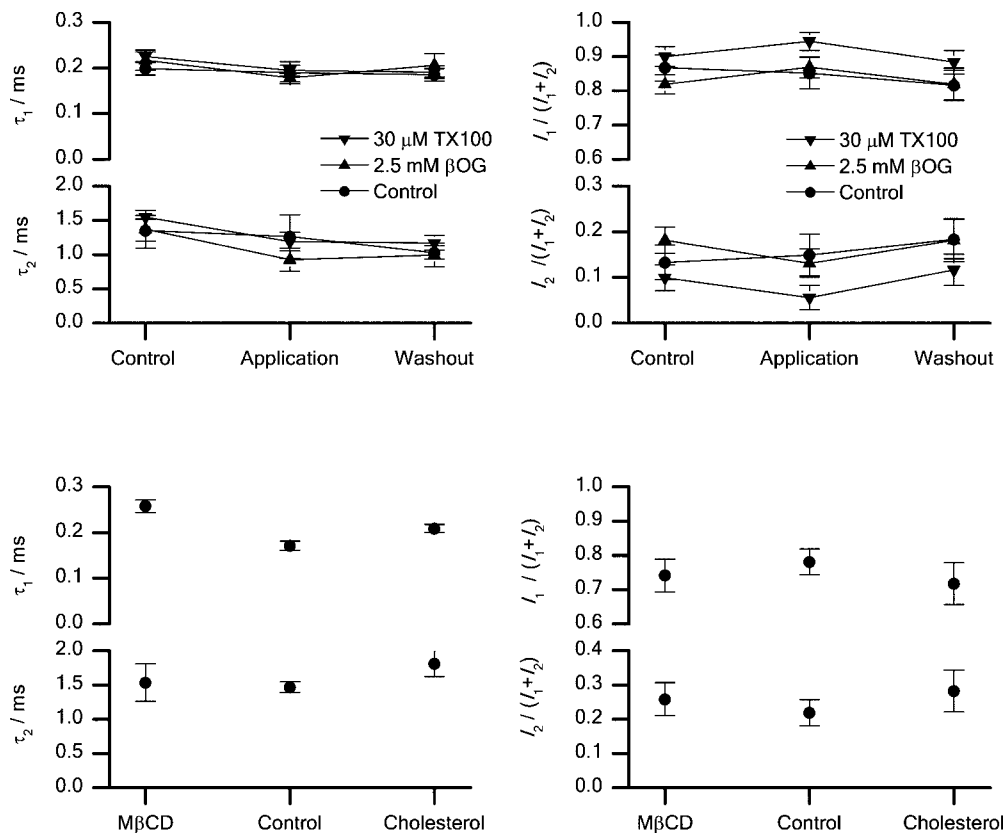


FIGURE S1. The effects of TX100,  $\beta$ OG, and cholesterol concentration on the kinetic parameters of the time course of inactivation at +20 mV. (Top left) Effects of 30  $\mu$ M TX100 ( $\nabla$ ) or 2.5 mM  $\beta$ OG ( $\blacktriangle$ ) on  $\tau_1$  and  $\tau_2$ . Control cells ( $\bullet$ ). (Top right) Effects of 30  $\mu$ M TX100 ( $\nabla$ ) or 2.5 mM  $\beta$ OG ( $\blacktriangle$ ) on  $I_1/(I_1 + I_2)$  and  $I_2/(I_1 + I_2)$ . Control cells ( $\bullet$ ). Effects of cholesterol content on  $\tau_1$  and  $\tau_2$  (bottom left) and  $I_1/(I_1 + I_2)$  and  $I_2/(I_1 + I_2)$  (bottom right). Cholesterol depletion significantly altered  $\tau_2$  ( $P < 0.05$ ). Mean  $\pm$  SEM,  $n = 5, 6, 6$  (TX100,  $\beta$ OG, timed controls); 9, 8, 6 (cholesterol-enriched, cholesterol-depleted, timed controls for cholesterol experiments).

where  $k$  is a phenomenological delay and  $\tau_a$  is the activation time constant. (Initial analysis showed that an exponent of three gave a better fit than two or four [not depicted].) Neither  $k$  nor  $\tau_a$  were significantly altered by changing the filter and sample frequency from 10 and 40 kHz ( $n = 3$ ) to 50 and 200 kHz, respectively ( $n = 3$ ) ( $P > 0.4$  and  $P > 0.4$  [not depicted]).

## RESULTS

### *Effects of $\beta$ OG, TX100, and Cholesterol on the Kinetics of Inactivation*

In control cells, the time course of inactivation was best described by a double-exponential decay with a major fast component and a minor slow component:  $\tau_1 = 0.20 \pm 0.01$  ms,  $I_1/(I_1 + I_2) = 0.87 \pm 0.04$ ;  $\tau_2 = 1.4 \pm 0.16$  ms,  $I_2/(I_1 + I_2) = 0.13 \pm 0.04$  ms. The top panels in Fig. S1 show the effects of 30  $\mu$ M TX100 or 2.5 mM  $\beta$ OG on  $\tau_1$ ,  $\tau_2$ ,  $I_1/(I_1 + I_2)$  and  $I_2/(I_1 + I_2)$ .

Neither TX100 nor  $\beta$ OG altered these parameters. The bottom panels in Fig. S1 show the effects of cholesterol on the inactivation kinetics; increasing the cell cholesterol content altered neither  $\tau_1$  nor  $\tau_2$  nor their relative contributions to the current. Decreasing the cholesterol content increased  $\tau_1$  from  $0.17 \pm 0.01$  ms to  $0.26 \pm 0.01$  ms with no change in the relative contribution of the fast component.

### *Effects of $\beta$ OG, TX100, and Cholesterol on the Kinetics of Activation*

None of the experimental maneuvers altered the activation time constant ( $\tau_a$ ) significantly, but the time course of activation is so fast that small changes in  $\tau_a$  may have been masked by the time constant of the voltage clamp. Fig. S2 A shows the (lack of) effects of 30  $\mu$ M TX100 and 2.5 mM  $\beta$ OG on ( $\tau_a$ ); Fig. S2 B shows the corresponding (lack of) effects of changes in cholesterol content.

Whereas neither the application of the micelle-compounds or cholesterol enrichment altered the phenomenological delay ( $k$ ) relative to its value in timed control experiments, cholesterol-depletion decreased  $k$  from  $0.17 \pm 0.02$  ms in the timed control experiments to  $0.10 \pm 0.01$  ms ( $P < 0.05$ ). We do not understand why.

In conclusion, none of the experimental maneuvers, except for the effect of cholesterol-depletion on  $\tau_1$ , have major effects on the kinetics of channel activation and inactivation at +20 mV. That said, we are not able to discern modest changes (decreases) in  $\tau_a$ , which would tend to be obscured by the time course of the membrane charging. (The cholesterol-enrichment-induced changes in  $V_{act}$  certainly indicates that channel activation is effected by the maneuver.)

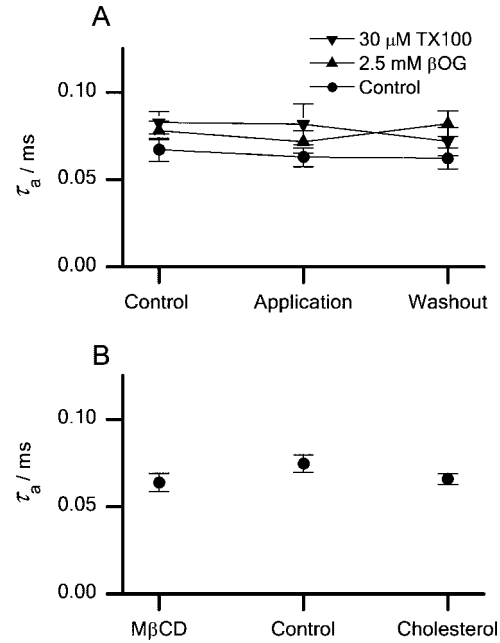


FIGURE S2. (A) Effects of 30  $\mu$ M TX100 ( $\blacktriangledown$ ) or 2.5 mM  $\beta$ OG ( $\blacktriangle$ ) on the time constant of activation,  $\tau_a$ . Control cells ( $\bullet$ ). (B) Effects of cholesterol content on  $\tau_a$ . Every 5 s the cells were depolarized to +20 mV from a holding potential of  $-80$  mV. Neither the addition of micelle nor manipulation of cholesterol content changed  $\tau_a$  relative to timed control experiments ( $P > 0.05$ ). Separate control groups for experiments with micelle-forming compounds and experiments with cholesterol were done at similar times from membrane rupture as the experimental group. Experimental conditions as in Fig. S1.