

Text S1: Steady state equations and relevant parameters for cAMP-PKA, MAPK and TOR pathways.

Steady state equations for upstream regulation of cAMP and MAPK pathways by Mep2.

(1) Fractional activation of Mep2 by ammonium sulphate (**Figure S1, Module1**)

$$fMep2 = \left(\frac{Am^{nH8}}{K^{nH8} + Am^{nH8}} \right) \times \left(\frac{K_9^{nH9}}{K_9^{nH9} + Am^{nH9}} \right)$$

(2) Ras2 activation module involving *cdc25p* and *Ira1* (**Figure S1, Module 2**)

$$\frac{k1 \times Ras2GTP \times Ira1}{Km1} = \frac{k2 \times Ras2GDP \times Cdc25}{Km2}$$

Activated Mep2 function to destabilize Ira1 by inactivating Gpb1/2, thereby reduces the active Ira1 concentration. Here, we assume that the total concentration of Ira1 is affected with activation of Mep2, which is given by $Ira1t = fMep2 \times Ira1max$.

(3) *Gpa2* regulation by Mep2 (**Figure S1, Module 3**)

$$k3 \times G\alpha\beta\gamma \times RL = k4 \times G\alpha GTP \times Rgs2$$

$$k4 \times G\alpha GTP \times Rgs2 = k5 \times G\alpha GDP \times G\beta\gamma$$

These equations represent the regulation of GPCR by binding of ligand to the receptor. Here, $G\alpha\beta\gamma$ represents *Gpa2-Gpb1/2* complex, $G\alpha GTP$ represents *Gpa2GTP*, $G\beta\gamma$ represents *Gpb1/2* and $G\alpha GDP$ represents *Gpa2GDP*. RL represents the receptor-ligand complex. Here, RL represents the Mep2 activated by ammonium sulphate.

Steady state equations relevant to cAMP and MAPK pathways (Figure S1, Module 4) were taken from our previous work [1].

<i>Reaction description</i>	<i>Equation</i>
<i>Flo8 activation by Tpk2</i>	$\frac{k11}{Km11}[Flo8][Tpk2] = \frac{k22}{Km22}[Flo8p][E2]$

Pde1p activation

$$\frac{k13}{Km13}[Pde1][Tpk2] = \frac{k14}{Km14}[Pde1p][E14]$$

cAMP rate balance

$$k16[Adc_a] \\ = \frac{k14}{Km14}[Pde2][cAMP] + \frac{k15}{Km15}[Pde1p][cAMP]$$

Cdc25 activation

$$\frac{k24}{Km24}[Cdc25][Kss1p] = \frac{k25}{Km25}[Cdc25p][E25]$$

Dig1/2 inactivation

$$\frac{k26}{Km26}[Dig1/2][Kss1p] = \frac{k27}{Km27}[Dig1/2p][E27]$$

Ste12Tec1 activation

$$\frac{k28}{Km28}[Ste12Tec1][Kss1p] = \frac{k29}{Km29}[Ste12pTec1p][E29]$$

Ste12Tec1_Dig1/2_kss1 complex phosphorylation by Ste7pp (MAPK activation by MAPKK)

$$\frac{k31}{Km31}[Ste12Tec1_Dig1/2_Kss1][Ste7pp] \\ = \frac{k32}{Km32}[Ste12Tec1_Dig1/2_Kss1p][E32]$$

Ste7 (MAPKK) activation by Ste20 (MAPKKK)

$$\frac{k34}{Km34}[Ste11p_Ste50][Ste7p] = \frac{k35}{Km35}[Ste7pp][E35]$$

Ste7p (MPKK) activation by Ste20 (MAPKKK)

$$\frac{k36}{Km36}[Ste11p_Ste50][Ste7] = \frac{k37}{Km37}[Ste7p][E37]$$

Ste11 (MAPKKK) activation by Ste20 complex (MAPKKKK)

$$\frac{k38}{Km38}[Ste20_Cdc42GTP_Bmh1/2][Ste11_Ste50] \\ = \frac{k39}{Km39}[Ste11p_Ste50][E39]$$

Cdc42 activation by Cdc24p

$$\frac{k43}{Km43}[Cdc42GDP][Cdc24p] \\ = \frac{k44}{Km44}[Cdc42GTP][Rgal]$$

- Nomenclature: If a protein is ‘A’ and another protein is ‘B’ then ‘A-B’ represents the complex between ‘A’ and ‘B’. Also, ‘Ap’ represents the activated protein (phosphorylated). Ei represents a phosphatase.
- Relevant parameters (**Rate constants, Michaelis Menten constant, total concentrations and disassociation constants**) for the module cAMP and MAPK (Module 4) were taken from Sengupta et al 2007 [1].

Steady state equations for TOR mediated control of G1 cyclins and Msn2/4 translocation (Figure S2)

(1) *Fractional activation of TOR by ammonium sulfate*

$$fTOR = \left(\frac{Am^{nH4}}{K_4^{nH4} + Am^{nH4}} \right)$$

(2) *Translational control of Cln3 by TOR*

$$C \ln 3 = C \ln 3_{\max} \left(\frac{TOR^{nH14}}{K_{14}^{nH14} + TOR^{nH14}} \right)$$

(3) *Inactivation of Phosphatase Pph21/22 by TOR*

$$Pph21/22 = Pph21/22_{\max} \left(\frac{K_{13}^{nH13}}{K_{13}^{nH13} + TOR^{nH13}} \right)$$

(4) *Control of nuclear translocation of Msn2/4 by Pph21/22 and Tpk*

$$Msn2/4_{cyc} \times K_{imp} \times \underbrace{\left(\frac{K_{10}^{nH10}}{Tpk^{nH10} + K_{10}^{nH10}} \right)}_{term1} = Msn2/4_{nuc} \times K_{exp} \times \underbrace{\left(\frac{K_{11}^{nH11}}{K_{11}^{nH11} + Pph21/22^{nH11}} \right)}_{term2} \times \left(\frac{Tpk^{nH12}}{K_{12}^{nH12} + Tpk^{nH12}} \right)$$

(5) *Sbf activation and inactivation by Cln3 and Clb2, respectively*

$$Va = kasbf \times C \ln 3$$

$$Vi = k_{isbf} + ki \times Clb2$$

$$kasbf \times C \ln 3 - (k_{isbf} + ki \times Clb2) \times Sbf = 0$$

(6) *Sbf mediated synthesis and degradation of G1 cyclin Cln1/2*

$$ks \times Sbf - k_{deg} \times C \ln 1 = 0$$

<i>Species</i>	<i>Total molar balances</i>
<i>Ras2t</i>	= <i>Ras2GTP + Ras2GDP + Ras2GDP_Cdc25p + Ras2GTP_Iral</i>
<i>Irat</i>	= <i>Iral + (Ras2GTP)(Iral) /Km1</i>
<i>Gat</i>	= <i>Gαβγ + GαGTP + GαGDP + Adc_GαGTP</i>
<i>Gβγt</i>	= <i>Gβγ + Gαβγ</i>
<i>Msn2t</i>	= <i>Msn2nuc + Msn2cyc</i>
<i>Adct</i>	= <i>Adc + Adc_GαGTP + Adc_Ras2GTP</i>
<i>Flo8t</i>	= <i>Flo8 + Flo8p</i>
<i>E2t</i>	= <i>E2 + Flo8p_E2</i>
<i>Pde1t</i>	= <i>Pde1 + Pde1p-Camp + Pde1p + Pde1p_E12 + Pde1_C</i>
<i>R2C2t</i>	= <i>2.R2C2 + 2 R2(Camp)4</i>
<i>Pde2t</i>	= <i>Pde2 = Pde2t/(1+ [Camp]/Km(2))</i>
<i>Ct</i>	= <i>2 R2C2 + C + Pde1_C</i>
<i>Ste12tec_t</i>	= <i>Kss1_Dig12_Ste12Tec1 + Kss1p_Dig12_Ste12Tec1 + Ste12Tec1 + Ste12Tec1p + Ste12Tec1_Kss1p + Ste12pTec1p_E29</i>
<i>Kss1t</i>	= <i>Kss1p_Dig12_Ste12Tec1 + Kss1p + Kss1_Dig12_Ste12Tec1 + Kss1 + Ste12Tec1_Kss1p + Kss1p_Dig12_Ste12Tec1_E32 + Kss1_Dig12_Ste12Tec1_Ste7p + Kss1p_Dig12 + Cdc25_Kss + Kssfus3</i>
<i>Dig12t</i>	= <i>Dig12p + Dig12 + Kss1_Dig12_Ste12Tec1 + Kss1p_Dig12_Ste12Tec1 + Kss1p_Dig12_Ste12Tec1_E32 + Kss1_Dig12_Ste12Tec1_Ste7pp + E27_Dig12p + Dig12_Kss1p</i>
<i>Cdc25t</i>	= <i>Cdc25p + Cdc25 + Cdc25_Kss + Cdc25_Fus3 + Cdc25_Kss1p + Cdc25p_E10</i>

$Fus3t$	$=$	$Fus3 + Cdc25_Fus3 + Kssfus3$
$E35t$	$=$	$E35 + (Ste7pp)(E35)/Km(2)$
$E37t$	$=$	$E37 + (Ste7p)(E37)/Km(4))$
$E39t$	$=$	$E39 + (Ste7p)(E39)/Km(6))$
$Ste7t$	$=$	$Ste7 + Ste7pp + Ste7p$ (<i>intermediate complexes are neglected</i>)
$Ste11ste50t$	$=$	$Ste11ste50 + Ste11pste50$
$Ste20t$	$=$	$Ste20_Hl7p + Ste20 + Cdc42p_Ste20_Bmh + Cdc42p_Ste20_Bmh$
$Hl7pt$	$=$	$Hl7p + Ste20_Hl7p$
$Rgalpt$	$=$	$Rgalp + Cdc42pGTP_Rgalp$
$Cdc42pt$	$=$	$Cdc42p_Gtp + Cdc42pGtp_Ste20 + Cdc42p_Ste20_Bmh + Cdc42p_Gdp + Cdc24p_Cdc42pGdp + Rgalp_Cdc42pGtp$

Relevant parameters (Rate constant, Michaelis Menten constant, total concentration and disassociation constant) for TOR pathway and Modules 1, 2 and 3 of Figure S1.

Component Concentrations (nM)

<i>Species</i>	<i>Conc. (nM)</i>	<i>Reference</i>	<i>Species</i>	<i>Conc.(nM)</i>	<i>Reference</i>
Pph21/22	160*	[4]	Adc	40	[2]
Msn2	2	[4]	Ira	30	[1]
Tor1	15	[4]	Sbf	50	[3]
Cln3	50	[3]	Rgs2	50	[2]
Cln1/2	50	[3]	G β α	75	[4]
Clb2	50	[3]	Mep2	50	[4]
G α	75	[4]	Ras2	200	[2]

Unknown concentrations were calculated from molecules numbers obtained from Yeast GFP fusion localization database (<http://yeastgfp.ucsf.edu>) [4]. Molecule numbers were converted into nM by considering a cell volume of 100fL

* Only 10% of Pph21/22 is actually involved in the TOR signaling [5].

Rate constants

<i>Nomenclature</i>	<i>Value</i> (min^{-1})	<i>Reference</i>	<i>Nomenclature</i>	<i>Value</i> (min^{-1})	<i>Reference</i>
k1	36	[7]	kasbf	0.38	[3]
k2	12	[7]	ki	8	[3]
k3	0.5	[2]	kisbf	0.8	[3]
k4	4	[2]	ks	0.15	[3]
k5	2.5	[2]	kdeg	0.12	[3]
kimp	1	[assumed]	kexp	1	[assumed]

<i>Dissociation constants</i>			<i>Michaelis Menten constants</i>		
<i>Kd</i>	<i>Value</i> (nM)	<i>Reference</i>	<i>Km</i>	<i>Value</i> (nM)	<i>Reference</i>
kd1	10	[6]	Km1	250	[7]
kd2	2	[2]	Km2	160	[7]

Hills Coefficient

<i>nH</i>	<i>value</i>	<i>Reference</i>	<i>nH</i>	<i>value</i>	<i>Reference</i>
nH1	4	[1]	nH9	2	[calculated]
nH2	0.8	[1]	nH10	2	[assumed]

nH3	3 - 4	[calculated]	nH11	3 - 4	[calculated]
nH4	0.3	[calculated]	nH12	2	[assumed]
nH5	2- 3	[calculated]	nH13	0.9 - 1.2	[calculated]
nH6	4	[1]	nH14	2	[calculated]
nH7	0.8	[1]			
nH8	1- 2	[calculated]			

K0.5	Value (nM)	Reference	K0.5	Value(nM)	Reference
K1	67	[1]	K9	1000*	[calculated]
K2	10	[1]	K10	10	[assumed]
K3	2.5	[calculated]	K11	3	[calculated]
K4	15-30*	[calculated]	K12	10	[assumed]
K5	5	[calculated]	K13	2	[calculated]
K6	14	[1]	K14	7.5	[calculated]
K7	2	[1]			
K	2-16 *	[calculated]			

* Units in μM based on the extracellular ammonium sulphate concentration

References

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