

Supplemental data

Experimental procedures

Oligonucleotides and plasmids

The open reading frames (ORF) of SART3, DDX30, DHX36, DDX47, RHA, HuR, hnRNP-C, RBM4, Matrin3, UPF1/RENT1, hnRNP-F, PABP-C1, ZBP1 and ZBP3 were cloned from HeLa Marathon cDNA library (Clontech Inc.). All cDNAs were PCR-amplified using primers that introduce Not1 restriction sites at the 5' end and BamH1 or EcoR1 restriction sites at the 3' end of the ORF. Restricted cDNAs were subsequently ligated into pIRES-VP5 (Meister et al., 2004). For expression of recombinant proteins N-terminal fragments of Ago1, Ago2 or full length TRBP were cloned into pGEX-6P1 (Amersham) using EcoR1 and Xho1 restriction sites. The construction of the human FLAG/HA-Ago1 and FLAG/HA-Ago2 was reported earlier (Meister et al., 2004). miRNA reporter plasmids which express the firefly luciferase orf fused to a regulatory 3'UTR sequence and renilla luciferase as a transfection control were generated from pMIR-REPORT (Ambion) as follows: The renilla luciferase gene including a SV40 promoter and a poly(A) site was PCR-cloned from pRL-SV40 (Promega) into the SspI site of the pMIR-REPORT plasmid. Furthermore, the CMV promoter of the firefly luciferase was replaced by a HSV-TK promoter using PCR amplification from pRL-TK (Promega). A 3'UTR fragment from the KRAS mRNA was PCR-amplified from a published construct (Johnson et al., 2005) and inserted into the SacI and NaeI restriction sites of the reporter plasmid. Alternatively, the HMGA2 3'UTR sequence was PCR amplified from HEK 293 cDNA using oligonucleotides 5' CGCTGAGCTCTACTAATAGTTTGTTGATCTG and 5' CGCTGCCGGCGACCAAACCTTTATTACTCATT and cloned into the described reporter construct via the SacI and NaeI restriction sites. The SERBP1, DNAJB11 and Raver2 reporter constructs have been reported before (Beitzinger et al., 2007). Further, the following DNA oligonucleotides containing a sequence perfectly complementary to miR-21 were annealed, digested with SacI and NaeI and inserted into the SacI and NaeI restriction sites of the

reporter plasmid: 5'
CGCTGAGCTCATCGCCACCTTGTTTAAGCCTCAACATCAGTCTGATAAGCTAATT
AGACCTACGCACTCCAGGCCGGCTCGC, 5'
GCGAGCCGGCCTGGAGTGCGTAGGTCTAATTAGCTTATCAGACTGATGTTGAGGC
TTAAACAAGGTGGCGATGAGCTCAGCG. The construct carrying a mutated miR-21
binding site was cloned using the following DNA oligonucleotides: 5'
CGCTGAGCTCATCGCCACCTTGTTTAAGCCTCAACATCAGCACCATTCTATAATTA
GACCTACGCACTCCAGGCCGGCTCGC, 5'
GCGAGCCGGCCTGGAGTGCGTAGGTCTAATTATAGAATGGTGCTGATGTTGAGGC
TTAAACAAGGTGGCGATGAGCTCAGCG.

The following oligonucleotides were used for detection of KRAS mRNA by qRT-PCR:

ODN1, 5' TTTTAGGACTCTTCTTCCATATTA; ODN2, 5'
TGGGGCATGTGGAAGGTAGGGAGG. For qPCR validation of siRNA knock downs, the
following primers were used: 5' CTTGAGGTGGGATGTGTGTG, 5'
GCAGGAGAGGAAAGGAAAGG (RBM4); 5' AAGTGATGGAGGGTGCTGAC, 5'
TGCGTCCGTAATTGAAGTTG (YB1); 5' AGTTGTTGTCCCTCGTGACC, 5'
AGCCTTCTGTTGTTGGTGCT (ZBP3); 5' CACCTCAAAGCGAGCACATA, 5'
CAATAGCAGTGACCCAGGT (FMRp); 5' TGGTATCGTGGAAGGACTCATGAC,
5' ATGCCAGTGAGCTTCCCGTTCAGC (GAPDH).

The following siRNAs were used for knock down experiments: RBM4#1:

UUACGGCUUUGUGCACAUAUT, UAUGUGCACAAAGCCGUAAUT; RBM4#2:
GGAGCUUCGAGCCAAGUUUUT, AAACUUGGCUCGAAGCUCCT; YB1#1:
AACCUUCGUUGCGAUGACCUT, GGUCAUCGCAACGAAGGUUUT; YB1#2:
GCAGACCGUAACCAUUAUAUT, UAUA AUGGUUACGGUCUGCUT; YB1#3:
AGAAGGUCAUCGCAACGAAUT, UUCGUUGCGAUGACCUUCUUT; ZBP3#1:
UCCAGAACGCACUAUUAUAUT, UGUAAUAGUGCGUUCUGGAUT; FMRp#1:

GGCAGCUUGCCUCGAGAUUUT, AAUCUCGAGGCAAGCUGCCUT; FMRp#2:
CCUCCUGUAGGUUAUAAUAUT, UAUUAUAACCUACAGGAGGUT; FMRp#3:
GAACGUCUAAGAUCUGUUAUT, UAACAGAUCUUAGACGUUCUT; FMRp#4:
ACAGGUACUUUGUCUAAGAUT, UCUUAGACAAAGUACCUGUUT; ctrl.:
UUGUCUUGCAUUCGACUAAUT, UUAGUCGAAUGCAAGACAAUT. The siRNAs
against TNRC6B and Ago1-4 were described before (Meister et al., 2004).

The following 2'OMe oligoribonucleotides were used: let-7a 2'OMe: 5'-
AACTATACAACCTACTACCTCAT; BART5 2'OMe: 5'-
CTATCAAGGAAACAAAACCACTGT. BART5 is a miRNA derived from the Epstein-Barr
virus and used as a control for unspecific effects caused by the 2'-OMe oligoribonucleotides.

Tissue culture and transfection

HEK 293 cells were grown in Dulbecco's modified Eagle's medium (DMEM) (PAA) supplemented with 10% heat inactivated fetal bovine serum (Biochrom AG), 100 unit/ml penicillin and 100 μ g/ml streptomycin (Gibco) at 37°C in 5% CO₂ atmosphere. Cells were transfected using calcium phosphate. Approximately 3h prior to transfection cells were plated at 30% confluency. Per 15 cm dish 15 μ g plasmid DNA was diluted in 1082 μ l H₂O and 153 μ l 2M CaCl₂. 1250 μ l 2 \times HEPES-buffered saline (274 mM NaCl, 1.5 mM Na₂HPO₄, 54.6 mM HEPES-KOH pH 7.1) were added drop-wise under gentle agitation. The transfection solution was then sprinkled onto the cells and extracts were generated 48 h after transfection. For luciferase assays, HeLa cells were transfected as described below.

Mass spectrometry (MS) and protein identification

After Coomassie Blue staining gel lanes with corresponding glycerol gradient fractions, were cut into pieces of similar size (23 splices), and subjected to in-gel trypsinization (Shevchenko et al., 1996) The extracted peptides were analyzed by liquid chromatography-coupled tandem

MS[LC-MS/MS] on a Q-ToF Ultima mass spectrometer (Waters). MSMS spectra of doubly and triply charged precursors were acquired for max. 3.3 sec (0.1 sec interscan time). The raw data were processed and transformed into a peaklist using MassLynx software 4.0 (Waters) with the following settings: i) Smoothing: smooth window (channels) 3.00, number of smooths 3 using Savitzky Golay algorithm; ii) Centroiding: min. peak width at half high: 4, centroid top, 80 %.

The peak list of fragment spectra was searched against the NCBI non-redundant database (NCBIInr) with a mass accuracy of 0.2 Da for the parent ion (MS) and 0.2 Da for the fragment ions (MS/MS). Mascor was used as search engine. The peptides were constrained to be tryptic with a maximum of 1 missed cleavage. Carbamidomethylation of cysteines was considered as a fixed modification whereas oxidation of methionine residues was considered as a variable modification. The highest scoring peptide from each protein as well as single hit peptides entry has been manually inspected to eliminate any false positives in the data-set.

Protein expression and antibody production

For the production of GST-fusion proteins, plasmids containing fragments of Ago1, Ago2, full length TRBP or full length SART-3 were transformed into E.coli strain BL21(DE3) and grown as over night cultures. GST-tagged proteins were purified using glutathione sepharose according to the manufacturer's instructions (Amersham). For antibody production in rabbits, 200 μ g of recombinant protein was repeatedly injected into rabbits. After 5 injections, rabbits were euthanized and the final serum was obtained. Lou/C rats were immunized subcutaneously and intraperitoneally with a mixture of Ago1-GST fusion protein (50 μ g), 5 nmol CPG oligonucleotide (ODN 2006, TIB Molbiol, Berlin, Germany), 500 μ l PBS and 500 μ l IFA. After a six-week interval a final boost without adjuvant was given three days before fusion of the rat spleen cells with the murine myeloma cell line P3X63-Ag8.653 (ZIT). Hybridoma supernatants were tested in an ELISA using bacterial extracts from E. coli

expressing either the Ago1 fusion protein or an irrelevant GST fusion protein. Monoclonal antibodies reacting with Ago1-GST fusion protein and not with an irrelevant GST fusion protein were analysed in western blotting. Ago1-4B8 (rat IgG2a) recognized the protein specifically in western blotting and was used for this study. The Ago1-4B8 antibody is commercially available from Ascenion (<http://www.ascenion.de/>).

Western blotting

For Western blotting, gradient fractions were separated by 10% SDS-PAGE and transferred to a nitrocellulose membrane (Amersham-Biosciences) by semi-dry electroblotting. The following primary antibodies were used: anti-HA (Upstate), anti-hnRNP-U, hnRNP-C1/2, anti-DDX5, anti-YB-1 (all from Abcam), anti-NF-90, anti-NF-45, anti-rpS6 (Cell signaling tech.), DDB1 (Serotec), anti-ZBP-1, anti-ZBP-3, anti-SART3 (see main text), anti-TRBP (see main text) anti-Ago1 (see main text) and anti-Ago2 (Upstate and see above). As secondary antibody, peroxidase-conjugated anti-mouse (Sigma), anti-goat (Abcam), anti-rat (Jackson labs) and anti-rabbit (Sigma) antibodies were used.

RT-PCR

For detection of miRNA association with Ago complexes, lysates from cells transfected with FLAG/HA-tagged Ago1 or Ago2 were separated on sucrose gradients as described. Ago complexes were immunoprecipitated from single fractions using FLAG beads. RNA was isolated and reverse transcription and semi-quantitative PCR was performed using mirVana qRT-PCR miRNA Detection Kit (Ambion) and miRVana qRT-PCR Primer Sets for the miRNAs let-7a and miRNA-16 (Ambion).

For qRT-PCR experiments a reporter construct containing the KRAS 3' UTR (Johnson et al., 2005) was transfected together with FLAG/HA-Ago1 into HEK 293 cells. Lysates were separated by gradient centrifugation and total RNA was extracted from each individual

fraction using phenol/chloroform. Extracted RNA was incubated with 1U DNaseI (Fermentas) and 20U RNasin (Promega) in 1x DNaseI buffer (Fermentas) for 30 min at 37°C followed by enzyme inactivation for 10 min at 70°C. Reverse transcription was performed using a first strand cDNA synthesis kit (Fermentas) according to manufacturer's instructions. The PCR reaction consisted of 2 µl cDNA, 5 nmol primer each and 1x LightCycler 480 SYBR Green I Master buffer (Roche). After initial activation (95°C, 10 min), the target sequence was amplified in 40 PCR cycles (95°C, 15 s; 55°C, 15 s; 72°C, 30 s). PCR Products were tested for specificity by melting point analysis. Quantification was carried out using the LightCycler 480 Basis Software (Roche).

For siRNA knockdown validation, HeLa cells were transfected with siRNAs using the conditions that were employed for the reporter assays. After 5d, total RNA was isolated using the PrepEase Kit (USB corporation, Cleveland, USA). Reverse transcription was performed with random hexamer primers using a first strand cDNA synthesis kit (Fermentas) according to manufacturer's instructions. qPCR was carried out using 500 nM primer each and 1x iQ SYBR Green supermix (Biorad). After initial activation (95°C, 3 min), the target sequence was amplified in 40 PCR cycles (95°C, 10 s; 55°C, 30 s) on a MYiQ (Biorad). Data were evaluated using the iQ5 software (Biorad). The levels of specific mRNAs were normalized to corresponding levels of GAPDH mRNA.

RNA cleavage experiments

In vitro transcribed cleavage substrates used in this study were described previously in (Landthaler et al., 2004; Meister et al., 2004). For RISC activity assays, the miR-19b substrate was ³²P-cap labeled as described (Martinez et al., 2002) and 10 µl of Ago complex-containing anti-FLAG beads were incubated in a 25 µl reaction containing 5 nM target RNA, 1 mM ATP, 0.2 mM GTP, 10 U/mL RNasin (Promega), 100 mM KCl, 1.5 mM MgCl₂ and 0.5 mM DTT for 1.5 h at 30°C. The reaction was stopped by adding 200 µl proteinase K buffer

followed by proteinase K treatment (0.2 mg/ml). RNA was isolated with Phenol/Chloroform and analysed by 8% denaturing RNA PAGE. Signals were detected by autoradiography. For Dicer activity assays, 10 μ l of Ago complex-containing anti-FLAG beads were incubated in 20 μ l phosphate buffered saline (PBS) containing 5 mM ATP, 7.5 mM MgCl₂, 10 U/ml RNasin (Promega) and about 100 counts of internally labeled Dicer for 1 h at 37°C. Beads were treated as described above, and the RNA products were analyzed on a 15% denaturing RNA gel. Radioactive signals were detected as described above.

Northern Blotting

For Northern Blotting, HEK 293 cell lysates transfected with FLAG/HA-Ago2 were subjected to FLAG immunoprecipitation and incubated with or without RNase A treatment as described above. Beads were subjected to Proteinase K treatment as described above. RNA was isolated from the beads using Phenol/Chloroform and separated by 15 % denaturing RNA PAGE. Northern Blotting against miR-19b was performed as described before (Lagos-Quintana et al., 2001).

Luciferase reporter assays

HeLa cells were seeded in complete DMEM to 48 well plates 5hrs prior to transfection to give 50% confluency.

siRNAs were transfected at 10mM final concentration using Hiperfect (Qiagen), according to the manufacturer's instructions. After 2d, cells were passaged in a 1:3 ratio and seeded into a 48 well plate. On the following day, medium was changed to OptiMEM (Invitrogen). Cells were subsequently transfected with reporter plasmids using 0.5 μ l/well Lipofectamine (Invitrogen) and 0.125 μ g/well plasmid, according to the manufacturer's instructions. 24hrs later, lipofection complexes were removed and complete DMEM was added. Cells were lysed 1d later in 50 μ l/well passive lysis buffer (Promega). Luminescence was measured in a

Mithras LB 940 luminometer (Berthold Technologies, Bad Wildbad, Germany), using firefly luciferase buffer (470 μ M D-luciferin, 530 μ M ATP, 270 μ M coenzyme A, 33.3 mM DTT, 20 mM Tricine, 5.34 mM magnesium sulfate, 0.1 mM EDTA; pH adjusted to 7.8) and renilla luciferase buffer (1.43 μ M Coelenterazine, 2.2 mM EDTA, 0.22 M potassium phosphate, 0.44 mg/ml BSA, 1.1 M NaCl, 1.3 mM NaN₃; pH adjusted to 5.0). Luciferase substrates were purchased from PJK cryosystems (Kleinblittersdorf, Germany). All samples were assayed in 4-6 replicates. For each siRNA, the firefly/renilla luminescence ratios of the miRNA reporter plasmids were normalized to the corresponding ratio of the empty reporter vector.

For miRNA inhibition experiments 40 pmol of the 2'-O-methyl oligoribonucleotides were pre-transfected into HeLa cells using EscortV (Sigma), according to the manufacturer's instructions. After 6hrs, the medium on the cells was exchanged and cells were transfected with reporter plasmids (0.1 μ g DNA/well) using 1 μ l/well EscortV (Sigma), according to the manufacturer's instructions. Cells were lysed 24hrs later in 50 μ l/well passive lysis buffer (Promega) and luminescence was measured as described above. The firefly/renilla luminescence ratios of the miRNA reporter plasmids were normalized to the corresponding ratio of the empty reporter vector.

For over-expression experiments, cells were co-transfected with reporter plasmids (0.1 μ g DNA/well) and FLAG/HA-tagged constructs as indicated using 1 μ l/well EscortV (Sigma), according to the manufacturer's instructions. After 36 h, cells were lysed in 50 μ l/well passive lysis buffer (Promega) and luminescence was measured as described above. Again, results were normalized to those of the empty vector.

Supplemental references

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Supplementary Figure legends

Supplementary Figure 1. Proteomic analysis of Ago complexes I, II and III. HEK 293 cells were transfected with FLAG/HA-Ago1 or FLAG/HA-Ago2. Lysates were separated by sucrose gradient centrifugation. Fractions 3-8 (complex I), 11-13 (complex II) and 15-17 (complex III) were pooled and subjected to immunoprecipitation using anti-FLAG antibodies. Immunoprecipitated FLAG/HA-Ago1 complexes (lanes 2, 4 and 6) or FLAG/HA-Ago2 complexes (lanes 8, 10 and 12) were separated by SDS PAGE and the proteins were analyzed

by ESI TOF mass spectrometry. Lanes 1, 3, 5, 7, 9 and 11 show molecular weight markers. IgG as well as FLAG/HA-Ago1 and -Ago2 bands are indicated.

Supplementary Figure 2. Proteomic analysis of control samples. Lysates from HEK 293 cells transfected with FLAG/HA-Ago2 were separated by gradient centrifugation and fractions were pooled as in S1. Immunoprecipitation using agarose coupled mouse IgG and proteomic analysis were performed as described in S1. Lanes 2, 4 and 6 show immunoprecipitates from Ago complexes I, II and III, respectively, obtained with control antibody. IgG bands are denoted to the right, lanes 1, 3, 5 and 7 show molecular weight markers.

Supplementary Figure 3. Efficiency test of RNase treatment. (A) Total RNA from HEK 293 cells was incubated with (+) or without (-) RNase A, separated by RNA PAGE and stained with ethidium bromide. (B) Lysates from HEK 293 cells transfected with FLAG/HA-Ago2 were subjected to immunoprecipitation using FLAG beads. Beads were incubated with or without RNase A. Northern Blot against miR-19b was performed with RNA isolated from these beads. As a loading control, 15% of the beads were used for Western blotting against the HA-tag.

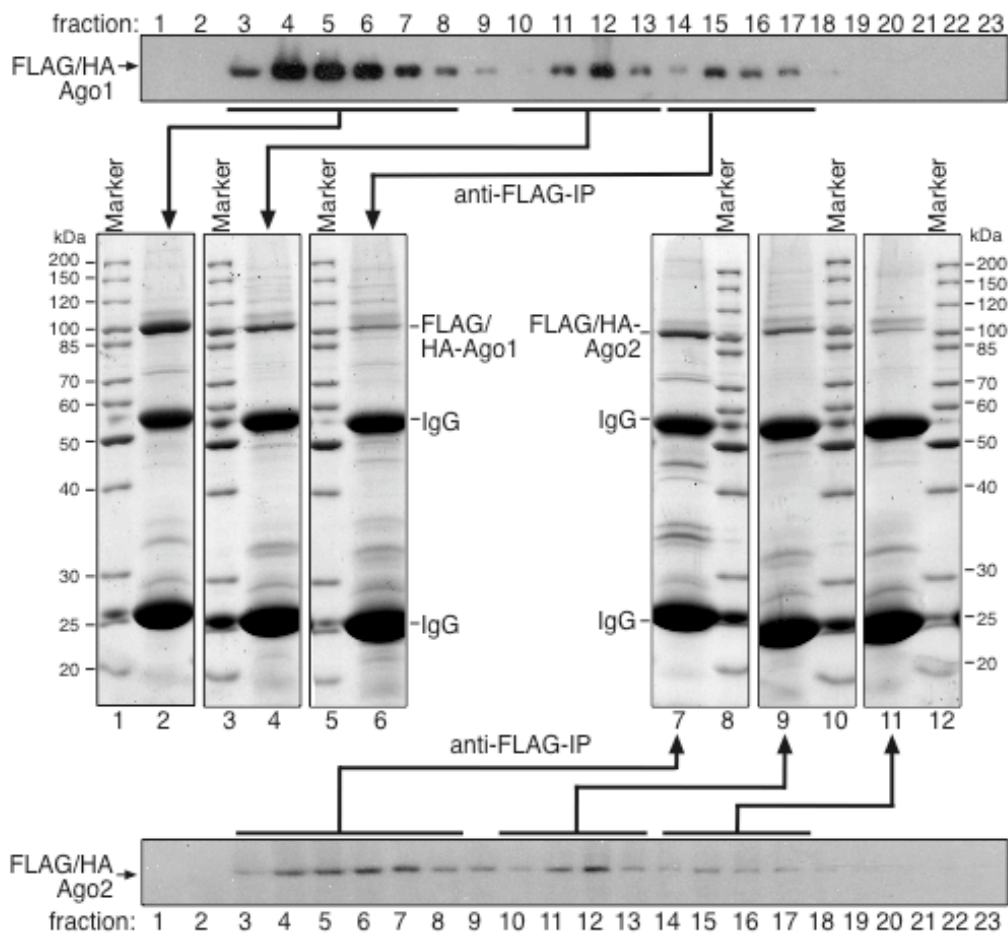
Supplementary Figure 4. SiRNA validation using qRT-PCR. Total RNA was reverse transcribed and cDNA was amplified by qPCR with primers specific to RBM4 (A), YB1 (B), ZBP3 (C) and FMRp (D). mRNA levels relative to GAPDH mRNA were normalized to control transfections. The error bars are derived from three different transfections.

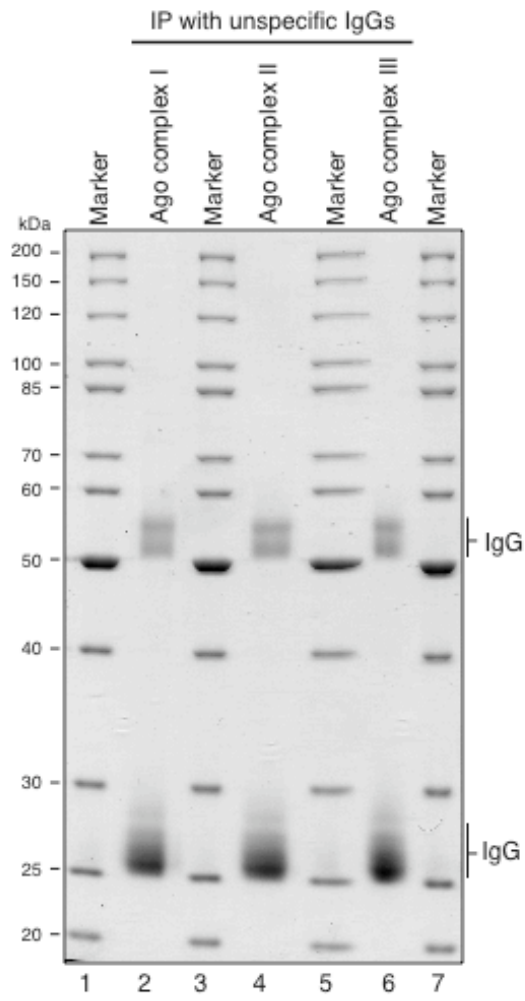
Supplementary Figure 5. Mutant miR-21 cleavage construct shows no significant difference compared to empty vector. SiRNAs against the indicated proteins were pre-transfected into HeLa cells. After 48hrs, a luciferase reporter containing a mutated binding site for miR-21 or

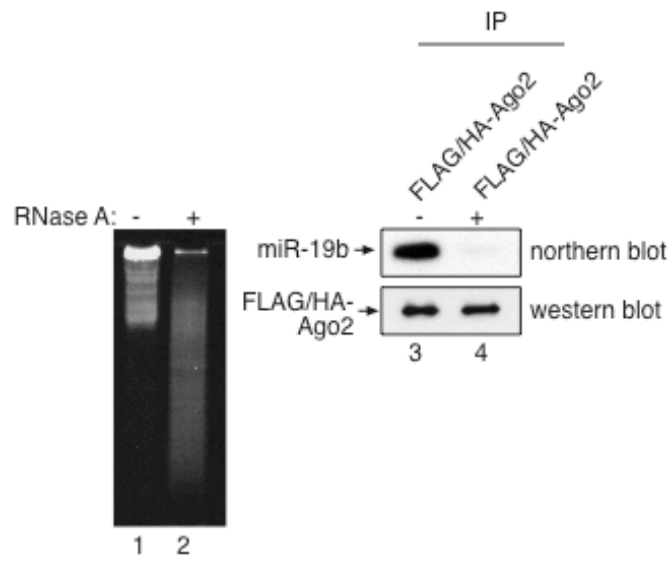
a control vector lacking the miR-21 binding site were transfected. Luciferase assays were done in triplicates. Results from the mutated reporter construct were normalized to those of the empty vector.

Supplementary Figure 6. Overexpression of RBM4 interferes with miRNA-regulated gene silencing. The indicated amounts of plasmids encoding for RBM4 and ZBP3 (control) were co-transfected into HeLa cells together with the KRAS luciferase reporter plasmid. Luciferase assays were done in triplicates. KRAS data were normalized to those of the empty vector.

A







A

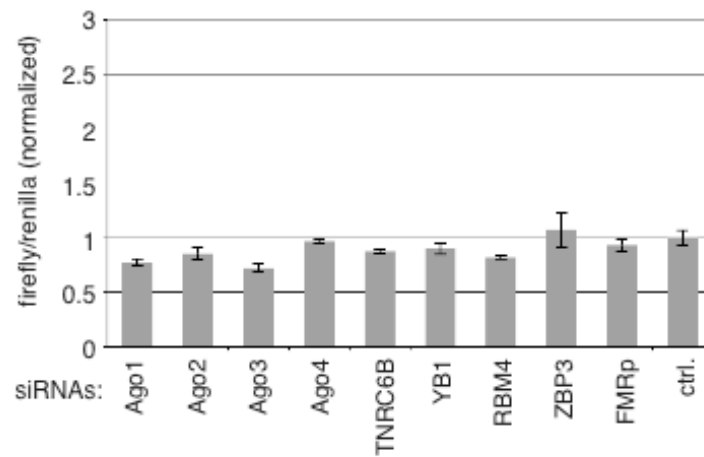
miR-21 cleavage construct

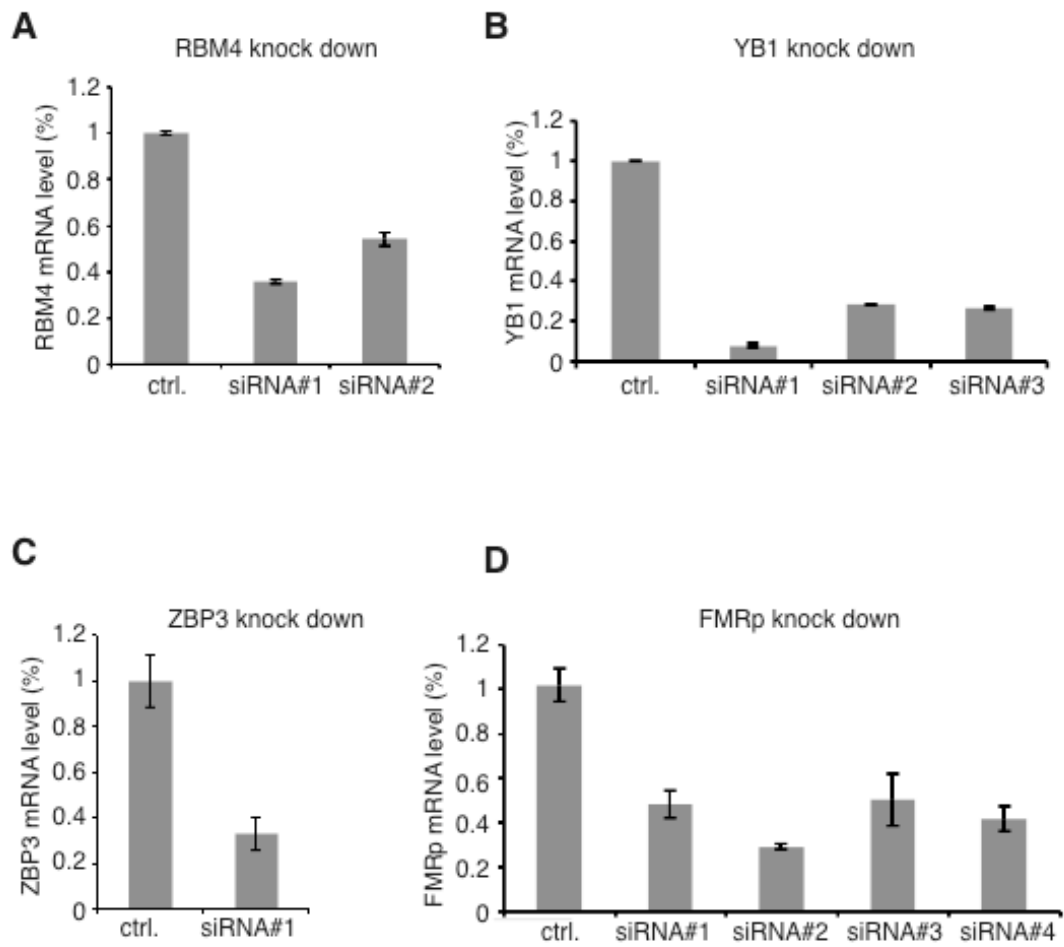
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 5'- TAGCTTATCAGACTGATGTTGA
 miR-21

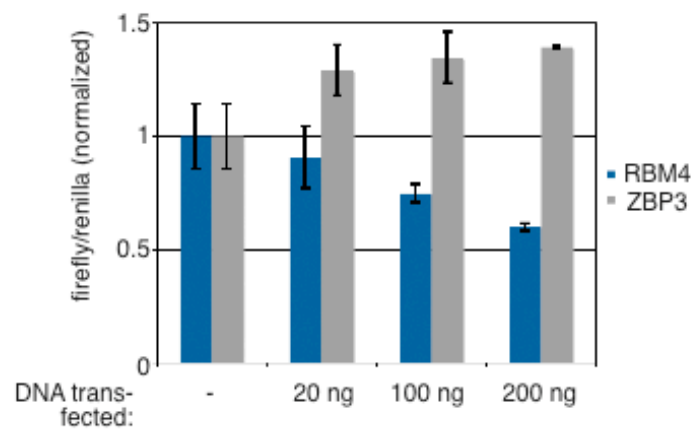
mutant cleavage construct

CGGCCGGACCTCACGCATCCAGATTAAT ATCT CCAC
 5'- TAGCTTATCAGACTGATGTTGA
 miR-21

B







Supplemental table 1: Protein composition of Ago1 complex 1

protein	Acc.No.	Mass	Queries matched	Proteinscore	Seq cov %	Peptidescore	Pep delta	Pep sequence
eukaryotic translation initiation factor 2C, 1 (Ago1) [Homo sapiens]	gi 6912352	97152	26	2058	58	87	-0.0038	K.NASYNLDPYIQEFGIK.V
Dicer	gi 21665773	217490	7	383	5	112	0.1540	K.SNAETATDLVLDLR.Y
squamous cell carcinoma antigen recognized by T cells 3 (SART3) [Homo sapiens]	gi 7961952	109865	6	158	6	40	0.0663	R.YSQVLDLR.Q
Na ⁺ /K ⁺ -ATPase alpha 1 subunit isoform a proprotein [Homo sapiens]	gi 21361181	112824	2	72	1	39	0.0775	R.LNIPYSQVNP.R.D
unnamed protein product [Homo sapiens]	gi 34535987	148788	1	24	-	24	0.0190	K.EPLLHFR.R
DNA damage binding protein 1 (Damage-specific DNA binding protein 1) (DDB p127 subunit) (DDBa) (UV-damaged DNA-binding protein 1) (UV-DDB 1)	gi 418316	126901	4	92	3	50	0.0530	K.LLASINSTR.L
sirtuin 1 [Homo sapiens]	gi 7657575	81630	1	68	1	68	0.0590	R.GDFNQVVP.R.C
HSPC273 [Homo sapiens]	gi 6841196	25891	1	66	4	66	0.0725	R.ELGENLDQLR.A
E1B-55kDa-associated protein (hnRNP U-like) [Homo sapiens]	gi 3319956	95750	1	48	0	48	0.0579	K.INEEISVK.H
programmed cell death 1 precursor [Homo sapiens]	gi 4826890	31687	1	20	2	20	0.0633	R.RTGQPLK.E
heat shock 90kDa protein [Homo sapiens]	gi 56204416	83212	12	686	24	61	0.0464	K.SIYYITGESK.E
Skb1/Its (PRMT5) [Homo sapiens]	gi 48145599	72636	12	749	25	75	0.0108	K.AALPTSIFLTK.K
aralar2 [Homo sapiens]	gi 6523256	74093	3	115	6	86	0.0492	K.TVELLSGVVDQTK.D
chaperonin (HSP60)	gi 306890	60986	7	374	21	55	0.0037	R.VTDALNATR.A
translation initiation factor eIF-2b delta subunit [Homo sapiens]	gi 6563202	57563	1	60	2	60	0.0320	R.VGTALQALVAR.A
elongation factor Tu [Homo sapiens]	gi 31092	50095	5	194	10	54	0.0206	K.IGGIGTVPVGR.V
RuvB-like 2 [Homo sapiens]	gi 5730023	51125	2	89	4	47	0.0332	K.GTEVQVDDIKR.V
Dnj3/Cpr3 [Homo sapiens]	gi 2352904	46277	1	145	10	32	0.0337	R.ELYDRYGEQGLR.E
trans-activation-responsive RNA-binding protein - human (TRBP) (fragment)	gi 107904	38614	1	36	2	36	0.0358	R.FIEIGSGTSK.K
KIAA0115 [Homo sapiens]	gi 473947	50680	1	31	1	31	0.0317	K.SSLNPIFR.G
MEP50 protein (MEP50) [Homo sapiens]	gi 13559060	36701	5	390	22	93	0.0469	K.VVDLAQQVLLSSYR.A
signal sequence receptor, alpha [Homo sapiens]	gi 4507237	32163	1	44	3	44	0.0306	K.GEDFPANNIVR.F
solute carrier family 25 member 3 isoform a precursor [Homo sapiens]	gi 6031192	40069	1	54	3	54	0.0282	R.IQTQPGYANTLR.D
S3 ribosomal protein [Homo sapiens]	gi 7765076	26699	2	206	25	68	0.0322	R.ELAEDGYSGVEVR.V
solute carrier family 25, member 5 [Homo sapiens]	gi 4502099	32874	7	190	22	49	0.0239	K.LLLQVGHASK.Q
Solute carrier family 25, member A6 [Homo sapiens]	gi 15928608	32905	7	204	22	49/48	0.0239/0.0088	R.GNLANVIR.Y
solute carrier family 25 member 4 variant [Homo sapiens]	gi 62089114	29328	3	131	11	47	0.0142	R.GNLANVIR.Y
transmembrane protein 33 [Homo sapiens]	gi 8922491	27933	1	56	4	56	0.0113	R.ALLANALTSALR.L
ribosomal protein L23 [Homo sapiens]	gi 4506605	14856	1	28	5	28	0.0137	K.NLIYISVK.G
Ribosomal protein S27-like protein [Homo sapiens]	gi 13277528	9472	1	58	9	58	0.0133	R.LITEGFSFR.R + Carb.
ribosomal protein L38 [Homo sapiens]	gi 3088356	4291	1	38	27	38	0.0253	K.QSLPPGLAVK.E
ribosomal protein S6	gi 225901	28633	1	28	3	28	0.0224	K.LIEVDDEK.K

Supplemental table 4: Protein composition of Ago2 complex 1

protein	Acc.No.	Mass	Queries matched	Proteinscore	Seq cov %	Peptidescore	Pep delta	Pep sequence
eukaryotic initiation factor 2C2 (Ago2) [Homo sapiens]	gi 29171734	97146	12	1104	40	91	0.0204	R.SVSIPIAPAYIAHLVAFRA
solute carrier family 25 member 3 isoform a precursor [Homo sapiens]	gi 6031192	40069	5	217	14	62	0.0382	R.IQTQPGYANTLR.D
Dicer [Homo sapiens]	gi 5019620	218673	6	353	5	112	0.0632	K.SNAETATDVLVLDLR.Y
KIAA1093 protein [Homo sapiens]	gi 14133235	183156	3	169	3	74	0.0778	K.TGSIAGSIAAR.G
HsGCN1 [Homo sapiens]	gi 2282576	211377	1	41	0	41	0.0405	K.ASLLPDPVPEVR.T
Na ⁺ K ⁺ ATPase [Homo sapiens]	gi 1359715	111901	1	122	3	47	0.0305	R.LNIPVSOVNFPR.D
RNA-binding protein 10 [RNA-binding motif protein 10] (DXS8237E)	gi 12644371	103396	1	40	1	40	0.0326	R.DGLGSDNIGSR.M
HSPC273 [Homo sapiens]	gi 8641186	25891	1	95	10	74	0.1177	R.ELLENDQLR.A
importin 4 [Homo sapiens]	gi 18700635	188642	1	70	1	41	0.0812	R.ELLFLPTEK.I
DNA damage binding protein 1 (Damage-specific DNA binding protein 1) (DDB p127 subunit) (DDBa) (UV-damaged DNA-binding protein 1) (UV-DDB 1)	gi 418316	126901	1	31	2	24	0.1030	K.VTLGTQPTVLR.T
importin 8 [Homo sapiens]	gi 53759103	119861	1	30	0	30	0.1061	K.IINPAPLLR.I
eukaryotic translation initiation factor 2C, 3 (Ago3) [Homo sapiens]	gi 5620478	71165	12	1104	26	91	0.0204	R.SVSIPIAPAYIAHLVAFRA
Heat shock protein 90 [Homo sapiens]	gi 56204416	83212	8	520	13	81	0.0209	R.GVVDSDELPLNISR.E
PREDICTED: similar to liver phosphofructokinase isoform b; 6-phosphofructokinase, liver type, liver-type 1-phosphofructokinase, phosphofructokinase 1	gi 55657570	85437	1	53	-	48	0.0934	R.FDEATQLR.G
initiation factor 4B [Homo sapiens]	gi 288100	69183	1	32	2	32	0.1246	R.AASIFGGAKPVDTAAR.E
MTHSP75	gi 292059	73734	2	25	6	33	0.0469	R.QAVTNPNNTFYATKR.L
Skb [Hs] [Homo sapiens]	gi 82581643	72520	10	710	25	66	0.1128	K.YSQYQQAIIYK.C
aralaz2 [Homo sapiens]	gi 523256	74093	2	100	2	61	0.1153	R.LQIAGETITGPR.V
proteasome subunit p58 [Homo sapiens]	gi 2656092	60968	1	44	2	44	0.0964	R.VYFELDKLDVLR.S
glycoprotein-associated amino acid transporter hb0,+AT1 [Homo sapiens]	gi 5823978	53436	1	32	1	32	0.0358	K.VLSIYVRR.L
Chaperonin [Homo sapiens]	gi 49522865	61016	5	348	10	79	0.0927	K.NAGVEGSLVKE.I
mitogen-activated protein kinase kinase kinase 7 interacting protein 1 isoform alpha [Homo sapiens]	gi 5174703	54510	1	64	2	64	0.1068	K.YGYTIDILSAAK.S
TATA binding protein interacting protein 49 kDa [Homo sapiens]	gi 4506753	50196	1	61	2	61	0.0746	K.QAASGLVQGENAR.E
elongation factor Tu	gi 556301	4	196	10	65	0.0505	K.THNIVVIGHVDSGK.S	
RuvB-like 2 [Homo sapiens]	gi 5730023	51125	2	140	4	82	0.0451	R.SGLDGLLEPR.R
trans-activation-responsive RNA-binding protein - human (TRBP) (fragment)	gi 107904	38914	1	46	2	46	0.0250	R.FIEIGSGTSK.K
unnamed protein product [Homo sapiens]	gi 31092	50095	3	88	6	39	0.0260	K.IGIGITVPVGR.V
HNRPF protein [Homo sapiens]	gi 16876910	45671	1	64	4	51	0.0328	R.VIEVFKSSQEEVRS.S
26S proteasome regulatory chain 4 [validated] - human	gi 345717	49210	1	46	-	46	0.0247	K.OVLYGPPSTGK.T
MEP50 protein (MEP50) [Homo sapiens]	gi 13559060	36701	4	343	21	89	0.0813	K.VWDLAQVQLSSYR.A
brain tumor associated protein LRR4 [Homo sapiens]	gi 14495561	72871	1	47	1	47	0.0122	R.MAEKCR.T
otopetrin 2 [Homo sapiens]	gi 30039714	62195	1	43	1	43	0.0444	R.EAWAIVSIPR.S
Cl channel	gi 228672	25672	1	92	5	92	0.0186	K.SLGTGLVIAEIR.L
7-dehydrocholesterol reductase [Homo sapiens]	gi 3171089	53073	1	38	1	38	0.0129	R.YTAAVPIR.L
oxidase (cytochrome c) assembly 1-like [Homo sapiens]	gi 4826880	55262	1	37	1	37	0.0062	R.NQLEAAR.G
[Human pre-mRNA splicing factor SF2p32, complete sequence], gene product	gi 338043	30888	4	469	33	81	0.0706	R.VSFQGTGESEVW.K
emeirin [Homo sapiens]	gi 4557553	28976	1	40	3	40	0.0357	R.AFGAGLGGQR.Q
Solute carrier family 25, member A6 [Homo sapiens]	gi 15928608	32905	12	420	33	66	0.0461	K.LLLQGVHASK.Q
solute carrier family 25, member 5 [Homo sapiens]	gi 4502099	32874	11	362	32	71	0.0513	K.DFLAGVHAASK.T
transmembrane protein 33 [Homo sapiens]	gi 8922491	27933	2	90	8	60	0.0839	R.ALLANALTSALR.L
ADP-ATP translocase	gi 339721	28042	1	25	4	25	0.0780	R.AAIFYGIYDAK.G
signal sequence receptor, delta [Homo sapiens]	gi 5454000	18887	1	87	13	44	0.0744	R.FFEESYSLR.K
ribosomal protein S20 [Homo sapiens]	gi 3088340	6853	1	38	20	38	0.0502	R.LIHLSPSEVQ.K
ribosomal protein L23 [Homo sapiens]	gi 4509605	14856	2	86	20	35	0.0407	K.GSAITGPVAK.E
NADH dehydrogenase (ubiquinone) 1 alpha subcomplex, 4, 9kDa [Homo sapiens]	gi 75517917	9394	2	52	22	37	0.0467	K.FFSVNWVYSK.L
ribosomal protein S27 [Homo sapiens]	gi 4432748	7886	1	49	17	49	0.0437	-DLLAPSPREEKR.K
ribosomal protein L38 [Homo sapiens]	gi 3088356	4291	1	29	29	29	0.0371	K.OSLPPPLAVK.E
c-myc binding protein [Homo sapiens]	gi 1785851	11945	1	24	10	24	0.0610	K.LAQYEPQEEK.R

