

SUPPLEMENTARY FIGURE LEGENDS

Figure S1 Hydrophaty plots of human (*Hs*) and zebrafish (*Dr*) sialidase proteins.

The amino acid position is plotted as a function of the average hydrophobicity (window size: 11 amino acids) following the Kyte and Doolittle methods. The profile was obtained using the DNA Strider 1.4 program. The shaded area present in *Hs* NEU1 plot indicates the leader peptide. The hydrophaty plots have been arranged based on the level of sequence identity between human and zebrafish sialidases.

Figure S2 *In situ* hybridization of sense probes for *neu3.1* and *neu4* (s-*neu3.1* and s-*neu4*).

A, sense probe for *neu3.1* in a 3 dpf embryo. B-D, sense probe for *neu4* in 21 hpf, 2 dpf and 3 dpf embryos. Embryos are in lateral (A, D) and dorsal (B, C) views with anterior to the left.

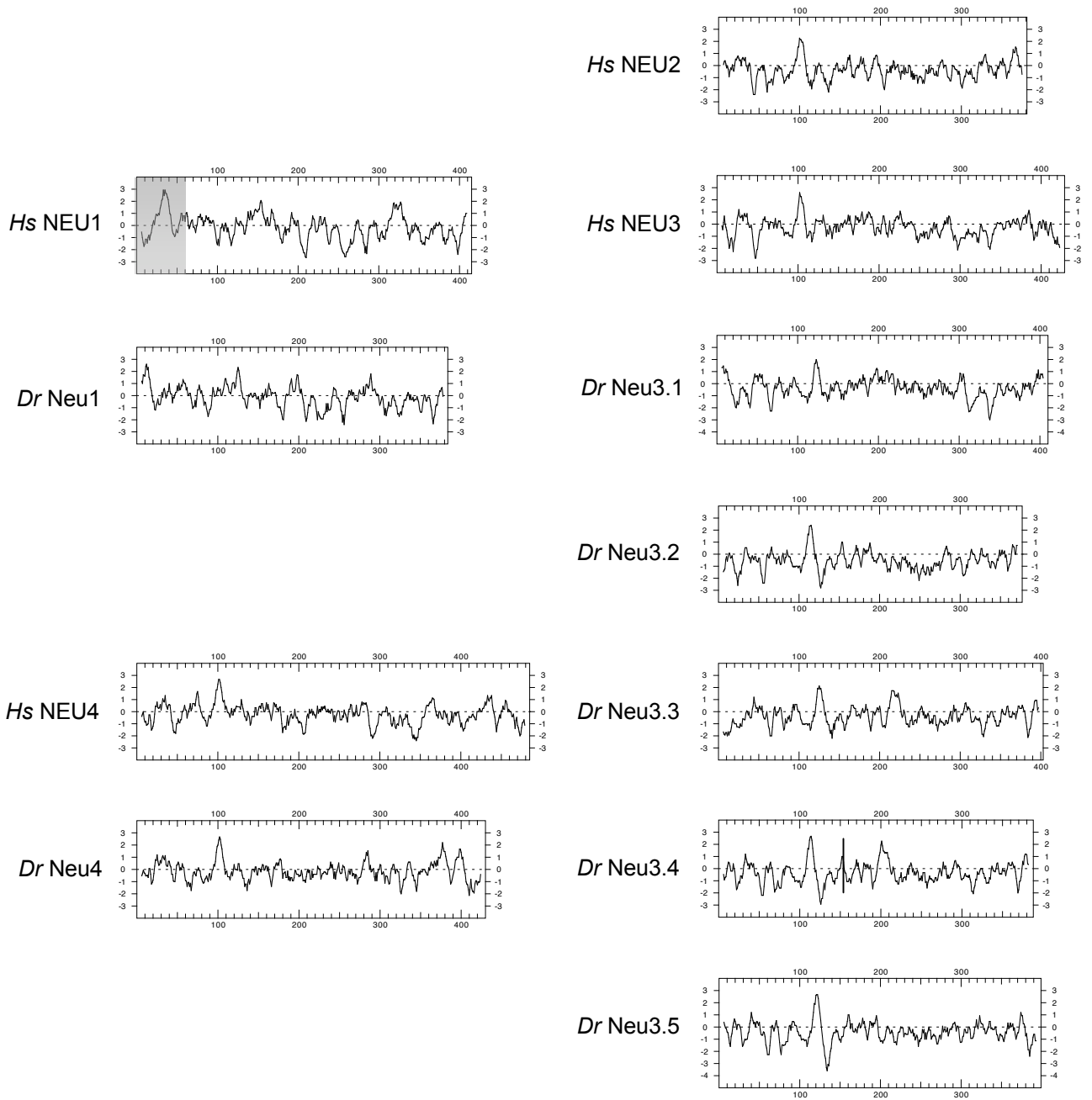
Figure S3 Expression of Neu1, Neu3.1, Neu3.2, Neu3.3 and Neu4 sialidases in COS7 cells.

COS-7 cells were transfected overnight with the constructs encoding Neu3.1-, Neu3.2-, Neu3.3- and Neu4-Myc epitope-tagged enzymes. Sialidase-specific activity toward the artificial substrate 4MU-NeuAc was determined as described in Materials and Methods using as enzymatic source aliquots of the crude cell extracts derived from mock and Neu3 expressing cells. Variations in the observed activity are indicated by the error bar (n = 3).

Figure S4 Subcellular localization of zebrafish sialidase Neu3.1.

COS7 cells expressing the Myc-tagged Neu3.1 sialidase were treated for double indirect immunofluorescence. *Left*: subcellular distribution of Neu3.1; *middle*: labeling of PDI (endoplasmic reticulum marker); *right*: merge of the previous staining patterns. The different primary antibodies were detected using isotype specific secondary antibodies conjugated with Alexa fluor 488 (green) and 555 (red). Specimens were analysed using a Zeiss Axiovert 200 microscope equipped with Confocal Laser System LSM 510 META.

Figure S1



Supplementary Figure

Figure S2

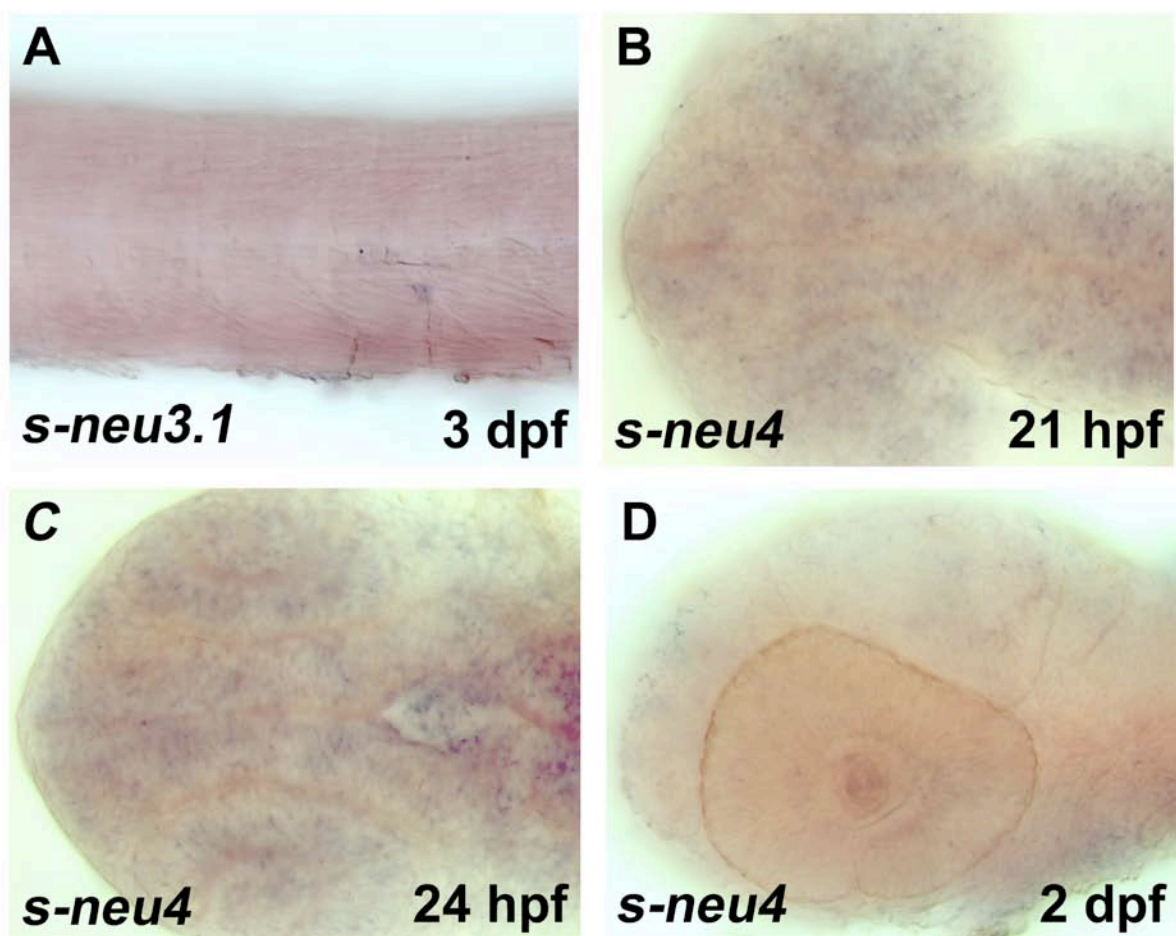


Figure S3

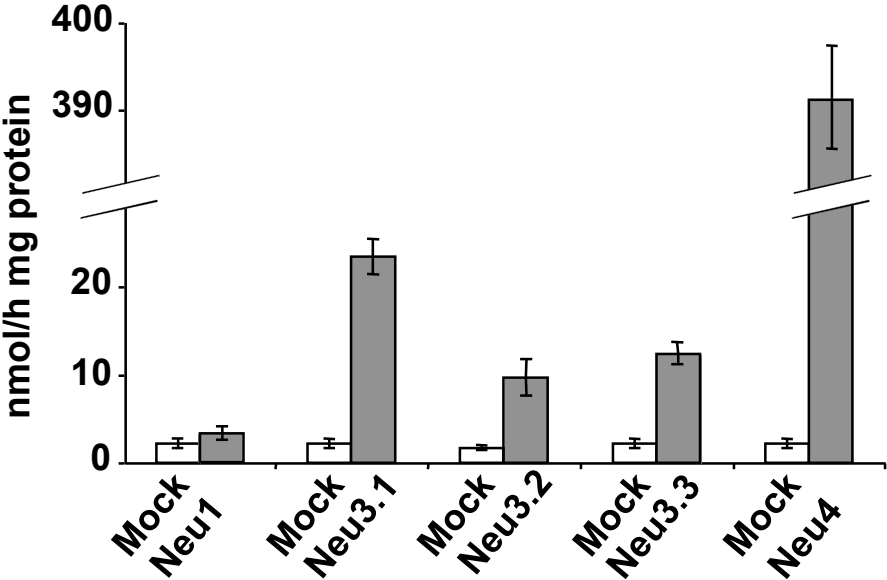


Figure S4

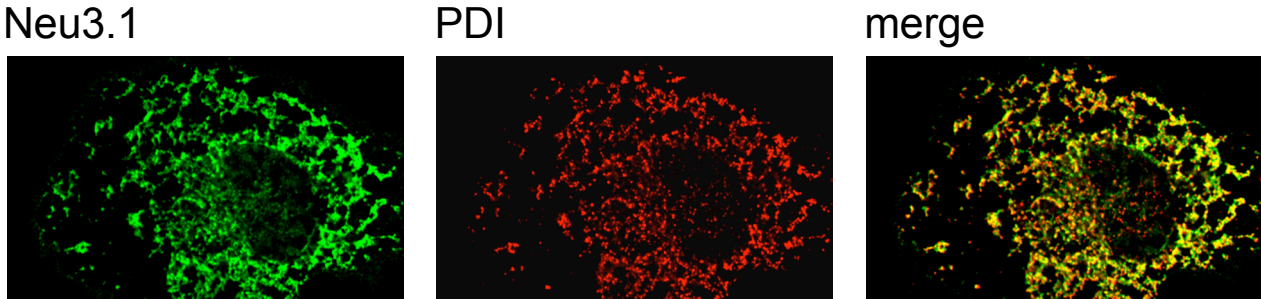


Table S1

General features of zebrafish sialidase genes

Gene name	Encoded polypeptide	Position on Zv6 assembly	Size (bp)
<i>neu1</i>	383	chr19:21918225-21931324	13100
<i>neu3.1</i>	409	chr21:19849217-19854475	5259
<i>neu3.2</i>	376	chr21:19844531-19847289	2759
<i>neu3.3</i>	402	chr21: 19835936-19840817	4882
<i>neu3.4</i>	387	chr21:19827188-19828973	1786
<i>neu3.5</i>	398	chr21:19821489-19822841	1353
<i>neu4</i>	429	chrNA_random:108563257-108570506	7250

Note

chrNA: Whole Genome Shotgun (WGS) contigs that could not be related to any FingerPrint Contig (FPC). These are unmapped scaffolds in random order separated by 1000 Ns between scaffolds. In the Zv7 assembly the *neu4* gene maps on chr 15.

Table S2Amino acid sequence comparison between human (*Hs*) and zebrafish (*Dr*) sialidases

	<i>Hs</i> NEU1	<i>Hs</i> NEU2	<i>Hs</i> NEU3	<i>Hs</i> NEU4
<i>Hs</i> NEU1				
<i>Hs</i> NEU2	18			
<i>Hs</i> NEU3	18	40		
<i>Hs</i> NEU4	20	43	44	
<i>Dr</i> Neu1	58	17	20	21
<i>Dr</i> Neu3.1	15	38	49	41
<i>Dr</i> Neu3.2	15	37	46	42
<i>Dr</i> Neu3.3	17	41	42	40
<i>Dr</i> Neu3.4	17	39	47	40
<i>Dr</i> Neu3.5	15	39	43	39
<i>Dr</i> Neu4	15	42	40	48

Numbers represent amino acid percentage identity calculated with the ClustalW algorithm. The highest values of identity in pairwise alignments are highlighted in bold.

Table S3Amino acid sequence comparison among zebrafish (*Dr*) sialidases

	<i>Dr</i> Neu1	<i>Dr</i> Neu3.1	<i>Dr</i> Neu3.2	<i>Dr</i> Neu3.3	<i>Dr</i> Neu3.4	<i>Dr</i> Neu3.5
<i>Dr</i> Neu3.1	17					
<i>Dr</i> Neu3.2	20	54				
<i>Dr</i> Neu3.3	18	48	59			
<i>Dr</i> Neu3.4	18	55	63	79		
<i>Dr</i> Neu3.5	18	55	73	57	67	
<i>Dr</i> Neu4	21	44	45	41	45	43

Numbers represent amino acid percentage identity calculated with the ClustalW algorithm. The highest values of identity in pairwise alignments are highlighted in bold.