

SI Text

Protein Preparation. The portion of *RNQ1* encoding amino acid residues 153 to 405 was amplified from pKanMXRNQ1–GFP (1) using primers 423 (CCCCATATGCAAGGTCAGGGACAAGGTCAA) and 424 (CCCAAGCTTTTCAGTGATGGTGATGGTGGTAGCGGTTCTGGTTGCCGTT), digested with NdeI and HindIII and ligated to pET-21a(+) (Novagen, Darmstadt, Germany) cut with the same enzymes to form p1178. *E. coli* strain BL21-CodonPlus (DE3)-RIPL carrying p1178 was grown overnight in rich medium containing 100 µg/ml ampicillin, 75 µg/ml streptomycin, and 34 µg/ml chloramphenicol.

Media for labeling were based on described methods (2, 3). Minimal medium contained, per liter, 200 ml of salts (65 g/liter KH₂PO₄, 50 g/liter K₂HPO₄, 45 g/liter Na₂PO₄, 10 g/liter NH₄Cl), 10 ml of trace elements (6 g/liter FeSO₄, 6 g/liter CaCl₂, 1.2 g/liter MnCl₂, 0.8 g/liter CoCl₂, 0.7 g/liter ZnSO₄, 0.3 g/liter CuCl₂, 0.25 g/liter (NH₄)₆Mo₇O₂₄, 5 g/liter EDTA), 10 ml of 1 M MgSO₄, 12 ml of 40% dextrose, 100 ml of TAU (0.3 g/liter thiamine, 2 g/liter adenine sulfate, 2 g/liter uracil), 100 ml of amino acid mix (1 g/liter of each amino acid except tyrosine), 100 ml of 0.1% tyrosine, and 50 µg/ml ampicillin. Cells are grown (500 ml per 2 l flask) at 37C to OD₅₅₀ = 0.5, collected by centrifugation, and suspended in fresh medium with 200 mg/liter of labeled tyrosine or leucine in place of that unlabeled amino acid. To make Ala-3-¹³C labeled protein, 1 g/liter of Ala-3-¹³C replaced alanine in the medium. After 15 min growth, the culture was made 1 mM in IPTG and growth continued overnight. Uniformly ¹³C, ¹⁵N-labeled Rnq1¹⁵³⁻⁴⁰⁵ was prepared by the same method except that at OD₅₅₀ = 0.5, cells were resuspended in media lacking amino acids and containing 2 g/liter ¹⁵NH₄Cl and 0.5 g/liter U-¹³C-glucose. After 15 min of shaking at 37C, 1 mM IPTG and 4.5 g/liter U-¹³C-glucose were added.

Cells from two liters of culture are harvested, suspended in 25 ml of 8 M guanidine HCl, 100 mM Tris Cl pH 8.0, 150 mM NaCl, with one “complete, EDTA-free” protease inhibitor tablet (Roche, Mannheim, Germany) incubated for 2 h to complete lysis, spun in a 45Ti rotor for 60 min at 30,000 rpm. The supernatant was mixed with 4 ml packed of NiNTI volume per liter culture, mixed gently at room temperature for 20 min. The mixture was poured into a column,

washed extensively with 8 M urea, 100 mM Tris Cl pH 8.0, 150 mM NaCl, and the Rnq1¹⁵³⁻⁴⁰⁵ was eluted using the same buffer with 200 mM imidazole. Protein was precipitated by addition of four volumes of methanol, and dissolved for amyloid formation in 4 M urea, 150 mM NaCl, 5 mM KPO₄ pH 7.4. After 2-5 days incubation at room temperature with gentle agitation, amyloid formation was complete. Samples were washed with H₂O four times, and dried by lyophilization.

To examine the effect of dilution, Rnq1¹⁵³⁻⁴⁰⁵ fully labeled with Tyr-1-¹³ was dissolved in 8 M guanidine, 100 mM Tris Cl pH8.0, 150 mM NaCl, and mixed with four times the weight (as determined by OD₂₈₀) of unlabeled material, methanol-precipitated, and dissolved in 4 M urea, 150 mM NaCl, 5 mM KPO₄ pH 7.4 for amyloid formation.

1. Nakayashiki T, Kurtzman CP, Edskes HK, Wickner RB (2005) Yeast prions [URE3] and [PSI⁺] are diseases. *Proc Natl Acad Sci USA* 102:10575-10580.
2. Blanco FJ, Hess S, Pannell LK, Rizzo NW, Tycko R (2001) Solid-state NMR data support a helix-loop-helix structural model for the N-terminal half of HIV-1 Rev in fibrillar form. *J Mol Biol* 313:845-859.
3. Cai ML, Huang Y, Sakaguchi K, Clore GM, Gronenborn AM, Craigie R (1998) An efficient and cost-effective isotope labeling protocol for proteins expressed in *Escherichia coli*. *J Biomol NMR* 11:97-102.

Sequence of Rnq1¹⁵³⁻⁴⁰⁵

(M) GQGQGQGQGQGQGQGQGSFTALASL
ASSFMNSNNNNQOGQONQSSGGSSEFGALASM
ASSFMHSNNNQNNSNNSQOQYQNSYQNGNQN
SQGYNNQOQYQGGNGGYQOQOQOQSGGAFSSL
ASMAQSYLGGGQTQSNQOQYNOQOQONNQOQ
YQOQOQONQYQHQQOQOQOQOQGHSSSF~~S~~ALAS
MASSYLGNNNSNSSSYGGQOQANEYGRPQQ
NGOQOQSNEYGRPQYGGNQNSNGQHESEFNFS
GNFSQONNNGNQNR~~Y~~ (H₆)

Amino acid composition of Rnq1¹⁵³⁻⁴⁰⁵.

Ala	A	13	Asn	N	41
Glu	E	3	Pro	P	2
Phe	F	9	Gln	Q	67
Gly	G	42	Arg	R	3
His	H	4	Ser	S	39
Leu	L	7	Thr	T	2
Met	M	5	Tyr	Y	15
