Supporting Information

A Phosphorylated Prodrug for the Inhibition of Pin1

Song Zhao, Felicia A. Etzkorn*

Department of Chemistry, Virginia Tech, Blacksburg, VA, 24061-0212.

Correspondence: fetzkorn@vt.edu

Experimental

General

Unless otherwise indicated, all reactions were carried out under N₂ in flame-dried glassware. THF and CH₂Cl₂ were dried by passage through alumina. Anhydrous (99.8%) DMF was available commercially and used directly from SureSealTM bottles. Dimethyl sulfoxide (DMSO) was anhydrous and dried with 4 Å molecular sieves. Triethylamine (TEA) was distilled from CaH₂, and oxalyl chloride (COCl)₂ was distilled before use each time. Diisopropylethylamine (DIEA) was distilled from CaH₂ under a N₂ atmosphere. Brine, NaHCO₃, and NH₄Cl refer to saturated aqueous solutions unless otherwise noted. Flash chromatography was performed on 32-63 µm or 230-400 mesh, ASTM silica gel with reagent grade solvents. NMR spectra were obtained at rt in CDCl₃ unless otherwise noted. Proton (400 MHz) NMR spectra were obtained for compounds 4-10, and carbon-13 (75 MHz) NMR spectra for compounds 4-10. Proton (500 MHz) NMR spectra, and carbon-13 (125 MHz) NMR spectra were obtained for compounds 1 and 2. Phosphorus (75 MHz) NMR spectra was obtained for compound 1, 2, 10. Rotamer peaks are indicated by listing ¹H chemical shifts separately; for ¹³C the minor rotamer peak is listed in parentheses. Coupling constant J values are given in Hz. Electrospray ionization (ESI-MS) was carried out on a triple quadrupole ThermoFinnigan TSQ MS. Human Pin1 recombinant protein was prepared as described.¹ Analytical reverse phase liquid chromatography (RP-HPLC) was performed on a RP C18, 250 \times 4.4 mm, 5 µm column (Varian Solaris). Preparative HPLC was performed using on a RP C18, 250 × 21.4 mm, 5 µm (Varian Solaris). HPLC solvents were A: water, B: CH₃CN. UV detection was performed at 220 nm unless otherwise noted.

Lactone 4. Fmoc-Ser- $\Psi[(Z)$ CH=C]-Pro-OH, **3** (40 mg, 0.096 mmol) and imidazole (33 mg, 0.48 mmol) were dissolved in DMF (2.0 mL), and TBSCl (29 mg, 0.19 mmol) was added. The mixture was stirred for 16 h, and NH₄Cl (2 mL) was added. The mixture was stirred for an additional 50 min, and diluted with EtOAc (20 mL), washed with NH₄Cl (2 × 10 mL), dried with Na₂SO₄, and concentrated. Chromatography on silica gel with 2% methanol in chloroform gave **4** (28 mg, 55%) as white powder. ¹H NMR (CDCl₃) δ 7.77 (d, *J* = 8.0, 2H),

7.57 (d, J = 8.0, 2H), 7.40 (app. t, J = 7.5, 2H), 7.32 (app. t, J = 6.5, 2H), 5.50 (br s, 1H), 4.73 (d, J = 8.5, 1H), 4.52? (br s, 1H), 4.46 (d, J = 6.0, 2H), 4.39 (t, J = 12, 1H) 4.22 (m, 2H), 3.92 (m, 1H), 2.37 (br s, 2H), 2.27 (two d, J = 7.1, 7.7, 1H), 1.97 (two d, J = 6.0, 7.1, 1H), 1.73 (two d, J = 6.0, 6.1, 1H), 1.60 (two d, J = 5.9, 7.8, 1H). ¹³C-NMR (CDCl₃) δ 173.2, 155.3, 143.6, 142.4, 141.3, 127.8, 127.0, 124.9, 120.0, 66.7, 66.3, 49.0, 47.1, 43.9, 35.1, 29.3, 24.4 ppm. HRMS calculated for C₂₄H₂₃NO₄ (MH⁺) m/z = 390.1705, found m/z = 390.1715.

Boc-Ser(TBS)-Ψ[(*Z***)CH=C]-Pro-OH 7.** Boc-Ser-Ψ[(*Z*)CH=C]-Pro-OH 6 (synthesized by the published method)² (111 mg, 0.388 mmol) and imidazole (136 mg, 2.00 mmol) were dissolved in DMF (2.0 mL), and TBSCl (151 mg, 1.00 mmol) was added with stirring. The reaction was stirred for 18 h at rt, then NH₄Cl (5 mL) was added. The mixture was stirred for an additional 60 min, and then diluted with EtOAc (20 mL), washed with NH₄Cl (2 × 10 mL), and brine (10 mL). The organic layer was dried over MgSO₄, and concentrated by rotary evaporation. Chromatography on silica gel with 30% EtOAc in hexane gave 150 mg (70%) of 7 as pale yellowish oil. ¹H-NMR (CDCl₃) δ 11.10 (br s, 1H), 5.96 and 4.91 (br s, 1 H), 5.42 (d, *J* = 5.8, 1H), 4.23 (br s, 1H), 3.61 and 3.59 (d, *J* = 4.4, 1H), 3.55 and 3.48 (br s, 2H), 2.41 (m, 1H), 2.23 (m, 1H), 2.04 (br s, 1H), 1.90 (m, 1H), 1.80 (br s, 1H), 1.55 (m, 1H), 1.37 (s, 9H), 0.83 (s, 9H), - 0.01 (s, 6H). ¹³C-NMR (CDCl₃) δ 178.5, 155.4 (157.0), 144.1 (145.4), 122.2, 79.1 (79.9), 65.3, 51.8 (52.7), 45.8, 33.6, 31.1, 28.2, 25.7, 24.0, 18.2, -5.5 ppm. HRMS calculated for C₂₀H₃₈NO₅Si (MH⁺) *m/z* = 400.2519, found *m/z* = 400.2485.

Boc-Ser(TBS)-Ψ[(Z)CH=C]-Pro-(2)-N-(3)-ethylaminoindole 8. Boc-Ser(TBS)- Ψ [(Z)CH=C]-Pro-OH, 7 (150 mg, 0.376 mmol) was dissolved in DMF (20 mL), and cooled to 0 °C for 10 min. HOAt (101 mg, 0.751 mmol), HATU (287 mg, 0.751 mmol) and DMAP (10 mg, 0.075 mmol) were added. Then DIEA (260 µL, 1.50 mmol) was added to the stirred solution dropwise. Tryptamine (120 mg, 0.751 mmol) was added slowly. The mixture was stirred for 6 h, diluted with EtOAc (200 mL), washed with water (2 × 50 mL), and brine (20 mL). The aqueous layer was back-extracted with CH_2Cl_2 (2 × 75 mL). The organic layers were combined, dried with Na₂SO₄ and concentrated. Chromatography on silica gel with 2% MeOH in CHCl₃ gave 184 mg (90%) of **8** as colorless oil. ¹H-NMR (CDCl₃), δ 8.82 (br s, 1H), 7.71 (br s, 1H), 7.65 (d, J = 7.6, 1H), 7.33 (d, J = 8.1, 1H), 7.13 (app t, J = 7.6, 1H), 7.05 (app t, J = 7.5, 1H), 6.97 (s, 1H), 5.32 (d, J = 8.8, 1H), 5.05 (s, 1H), 4.08 (m, 1H), 3.64 (m, 1H), 3.55 (m, 1H), 3.50 (two d, J = 6.2, 6.3, 2H), 3.35 (d, J = 7.0, 1H), 3.00 (app t, J = 7.7, 1H) 2H), 2.31 (m, 2H), 2.15 (m, 1H), 1.79 (m, 1H), 1.54 (m, 2H), 1.44 (s, 9H), 0.88 (s, 9H), 0.04 (s, 6H). ¹³C-NMR (CDCl₃) δ 171.9, 156.0, 144.3, 136.2, 127.5, 123.8, 121.9, 121.4, 118.7,

113.1, 111.1, 79.5, 64.9, 52.6, 47.5, 40.5, 38.5, 36.4, 32.8, 30.8, 28.3, 25.7, 25.0, 23.3, 18.2, -5.6. HRMS calculated for $C_{30}H_{48}N_3O_4$ (MH⁺) m/z = 542.3414, found m/z = 542.3403.

Boc-Ser-Ψ[(*Z***)CH=C]-Pro-(2)-***N***-(3) ethylaminoindole 9. Boc-Ser(TBS)-Ψ[(***Z***)CH=C]-Pro-(2)-***N***-(3) ethylaminoindole, 8** (92 mg, 0.17 mmol) was dissolved in THF (2.5 mL), and cooled to 0 °C for 10 min. A solution of TBAF (117 mg, 0.342 mmol) in THF (2.5 mL) was added dropwise at 0 °C. The mixture was stirred at rt for 4 h. The reaction was quenched with NH₄Cl (25 mL), and extracted with EtOAc (2 × 80 mL). The organic layer was washed with brine (20 mL), dried with Na₂SO₄ and concentrated. Chromatography on silica gel with 2% MeOH in CHCl₃ gave 85 mg (85%) of **9** as colorless oil. ¹H-NMR (CDCl₃), δ 8.60 (br s, 1H), 7.73 (br s, 1H), 7.63 (d, *J* = 7.8, 1H), 7.33 (d, *J* = 8.1, 1H), 7.15 (app t, *J* = 7.5, 1H), 7.07 (app t, *J* = 7.4, 1H), 6.99 (s, 1H), 5.33 (s, 1H), 5.30 (d, *J* = 8.8, 1H), 4.04 (m, 1H), 3.87 (br s, 1H), 3.60 (m, 1H), 3.53 (m, 2H), 3.45 (m, 2H), 1.41 (s, 9H). ¹³C-NMR (CDCl₃) δ 173.5, 156.5, 144.4, 136.1, 127.5, 123.4, 122.1, 121.7, 119.1, 118.8, 113.1, 111.1, 79.7, 64.7, 53.4, 47.4, 40.5, 33.4, 31.6, 28.3, 24.7, 23.8. HRMS calculated for C₂₄H₃₄N₃O₄ (MH⁺) *m/z* = 428.2549, found *m/z* = 428.2553.

Fmoc-Ser- Ψ **[**(*Z*)**CH=**C**]**-**Pro-**(2)-*N*-(3)-ethylaminoindole 5. Boc-Ser- Ψ **[**(*Z*)**CH=**C**]**-**Pro-**(2)-N-(3)-ethylaminoindole 9 (62 mg, 0.14 mmol) was dissolved in CH₂Cl₂ (3 mL), and cooled to 0 °C for 10 min. TFA (1 mL) was added dropwise. The mixture was stirred at 0 °C for 10 min and cold bath was removed. The mixture was stirred at rt for an additional 45 min and the solvent was evaporated. CH_2Cl_2 was added and evaporated (3 × 20 mL). The remaining TFA was removed under high vacuum overnight. Without further purification, the crude product was dissolved in a mixture solution of 10% aq. Na₂CO₃ and NaHCO₃ (3:1, 2 mL), then cooled to 0 °C for 10 min. A solution of Fmoc-Cl (43 mg, 0.17 mmol) in dioxane (2 mL) was added dropwise. After stirring at 0 °C for 20 h, the mixture was diluted with water (20 mL) and extracted with EtOAc (2×25 mL). The aqueous layer was acidified with 1M HCl to pH 3-4 and extracted with EtOAc (3×30 mL) and CH₂Cl₂ (3×30 mL). The organic layers were combined, washed with brine (20 mL), dried with Na₂SO₄, and concentrated. Chromatography on silica gel with 10 % MeOH in CHCl₃ gave 63 mg (78 %) of 5 as a colorless oil. ¹H-NMR (CDCl₃), δ 7.95 (br s, 1H), 7.77 (d, J = 7.5, 2H), 7.57 (d, J = 7.8, 2H), 7.53 (d, J = 6.2, 2H), 7.41 (app t, J = 7.5, 2H), 7.32 (app t, J = 6.8, 2H), 7.24 (s, 1H), 7.10 (app t, J = 7.5, 1H), 6.97 (app t, J = 7.4, 1H), 6.83 (br s, 1H), 5.23 (m, 2H), 4.37 (dd, J = 6.9, 10.5, 1H), 4.26 (d, J = 7.0, 9.3, 1H), 4.15 (app t, J = 6.6, 1H), 3.83 (m, 1H), 3.5-3.37 (m, 4H),

3.23 (d, J = 6.8, 1H), 3.00 (m, 1H), 2.86 (m, 1H), 2.32 (m, 1H), 2.19 (m, 2H), 1.84 (m, 1H), 1.54 (m, 2H). ¹³C-NMR (CDCl₃) δ 172.9, 156.7, 145.5, 143.9, 141.4, 136.2, 127.9, 127.3, 125.2, 125.1, 122.6, 122.2, 121.8, 120.2, 119.3, 118.9, 113.4, 111.2, 66.7, 64.6, 53.7, 47.9, 47.2, 40.7, 33.6, 31.7, 24.6, 24.0. HRMS calculated for C₃₄H₃₆N₃O₄ (MH⁺) m/z = 550.2706, found m/z = 550.2711.

Fmoc-Ser(PO(OH)₂)-Ψ[(Z)CH=C]-Pro-(2)-N-(3)-ethylaminoindole 1. Fmoc-Ser- $\Psi[(Z)CH=C]$ -Pro-(2)-N-(3)-ethylaminoindole, 5 (31 mg, 0.056 mmol) was dissolved in THF (3 mL). Tetrazole (36 mg, 0.22 mmol) and $(tBuO)_2P(N-iPr)_2$ (40 μ L, 0.11 mmol) were added. The mixture was stirred at rt for 20 h, then cooled to -40 °C for 10 min. tert-Butyl hydroperoxide (5 M in decane, 22 µL, 0.11 mmol) was added dropwise. The mixture was stirred at -40 °C for 40 min. The cold bath was removed and the reaction was stirred at rt for an additional 30 min. The mixture was cooled to 0 °C and 10% aq. Na₂S₂O₃ (3 mL) was added. After stirring for 10 min, the mixture was transferred to a separatory funnel using Et₂O (3 × 30 mL). The combined organic layers were washed with 10 % aq. Na₂S₂O₃ (2 × 20 mL) and brine (20 mL), dried with Na₂SO₄, and concentrated to give 40 mg of the crude $Fmoc-Ser(PO(O-tBu)_2)-\Psi[(Z)CH=C]-Pro-(2)-N-(3)-ethylaminoindole, 10, as a colorless oil.$ ¹H-NMR (CDCl₃), δ 8.30 (s, 1H), 7.70 (t, J = 10, 2H), 7.50-7.40 (m, 3H), 7.35 (m, 2H), 7.25 (m, 3H), 7.06 (app t, J = 7.5, 1H), 6.92 (m, 2H), 5.67 (m, 1H), 5.30 (m, 1H), 4.27 (d, J = 9.0, 1H), 4.10 (m, 2H), 3.90 (m, 2H), 3.50 (m, 3H), 3.35 (m, 1H), 2.95 (m, 2H), 2.22 (m, 3H), 1.90 (m, 1H), 1.45 (s, 9H). ³¹P-NMR (CDCl₃) δ –9.65 (s). ESI-MS gave the molecular ion $[M+H]^+$ m/z = 742.3, $[M+Na]^+$ m/z = 764.3. Without further purification (decomposes on silica gel), Fmoc-Ser(PO(OtBu)₂)- Ψ [(Z)CH=C]-Pro-(2)-N-(3)-ethylaminoindole (40 mg, 0.054mmol) was dissolved in CH₂Cl₂ (4 mL), and cooled to 0 °C. TFA (1mL) was added to the reaction mixture slowly, followed by the addition of water (0.2 mL) as scavenger. After stirring at 0 °C for 10 min, the cold bath was removed and the reaction mixture was stirred at rt for an additional 30 min, and the solvent was evaporated. CH₂Cl₂ was added and evaporated (5 \times 20 mL). The remaining TFA was removed under high vacuum overnight until constant weight was obtained. The crude product was purified by semipreparative C_{18} HPLC at 15 mL/min, 10 % to 90 % B over 20 min. Purified 1 (12 mg, 70% yield) eluted at 19.6 min as a white solid. Purity was 98.8 % by analytical C₁₈ HPLC (2 mL/min, 10 % to 90 % B over 13 min, retention time 11.79 min). ¹H-NMR (CD₃OD), δ 7.73 (dd, J = 7.4, 5.1, 2H), 7.55 (app t, J = 7.9, 2H), 7.39 (d, J = 7.3, 1H), 7.32 (m, 3H), 7.21 (dd, J = 7.0, 13.4, 2H), 7.03 (app t, J = 7.3, 1H), 6.97 (s, 1H), 6.90 (app t, J = 7.7, 1H), 5.43 (d, J = 9.3, 1H), 4.40 (m, 1H),

4.24 (m, 2H), 4.08 (app t, J = 6.6, 1H), 3.91 (m, 1H), 3.86 (m, 1H), 3.47 (m, 1H), 3.41 (app t, J = 7.6, 2H), 2.91 (m, 2H), 2.41 (m, 1H), 2.30 (m, 1H), 1.99 (m, 2H), 1.72 (m, 1H), 1.57 (m, 1H). ¹³C-NMR (CD₃OD) δ 147.3, 145.4, 145.2, 142.5, 138.1, 128.8, 128.7, 128.1, 126.3, 126.2, 125.0, 123.4, 122.3, 120.8, 119.5, 119.4, 113.3, 112.2, 68.1, 67.9, 52.7, 41.7, 35.2, 35.1, 33.1, 30.8, 26.0, 25.5. ³¹P- NMR (CD₃OD) δ -1.16 (s). ESI-MS [M+H]⁺ m/z = 630.3, [M+Na]⁺ m/z = 652.2, [M-H₃PO₄]⁺ m/z = 532.2. HRMS calcd for C₃₄H₃₇N₃O₇P (MH⁺) m/z = 630.2369, found m/z = 630.2340.

Fmoc-Ser(PO(OPOM)₂)-Ψ[(Z)CH=C]-Pro-(2)-N-(3)-ethylaminoindole 2. BisPOM phosphate was synthesized by a modification of the published procedure.³ Oxalyl chloride (90 µL, 1.9 mmol) was added dropwise to CH₂Cl₂ (1.5 mL) at 0 °C. DMF (7 µL) was added in one portion. A solution of hydrogen bisPOM phosphate (62 mg, 0.19 mmol) in CH₂Cl₂ (1.5 mL) was added dropwise at 0 °C over 15 min. The reaction mixture was stirred at rt for 2 h. The solvent and (COCl)₂ were removed by rotary evaporation. The remaining oxalyl chloride was removed in vacuo until constant weight was obtained. The product was obtained as slightly yellowish oil (50 mg, 80%). Without further purification, the bisPOMphosphoryl chloride was used immediately in the next step. ¹H-NMR (CDCl₃) δ 5.71 (m, 4H), 1.23 (s, 18H). ¹³C-NMR (CDCl₃) δ 83.52, 83.45, 31.0, 27.0. ³¹P-NMR (CDCl₃), δ 3.80 (s). Fmoc-Ser- $\Psi[(Z)CH=C]$ -Pro-(2)-N-(3)-ethylaminoindole, 5 (21 mg, 0.038 mmol) was dissolved in THF (1.5 mL) and cooled to -40 °C. Pyridine (0.75 mL) was added, and the mixture was stirred at -40 °C for 20 min. DMAP (2.5 mg) was added, then a solution of bisPOM phosphoryl chloride (50 mg, 0.14 mmol) in THF (0.9 mL) was added to the reaction mixture dropwise via syringe at -40 °C. The mixture was stirred at -40 °C for 2 h. A second batch of bisPOM phosphoryl chloride solution, (20 mg, 0.058 mmol) in THF (0.5 mL) was added. The mixture was stirred for 30 min. The cold bath was removed and the reaction mixture was stirred at rt for 6 h. Water (2 mL) was added to quench the reaction and the reaction mixture was diluted with EtOAc (40 mL). The organic layer was washed with water (2×20 mL). The aqueous layer was back-extracted with EtOAc (3×20 mL). The combined organic layers were washed with brine (20 mL), dried with Na₂SO₄, and concentrated. The crude product was purified using semipreparative C₁₈ HPLC (15 mL/min, 60% B for 5 min, then 60% to 95% B over 20 min). Purified 2 (3 mg, 20% yield) eluted at 23.0 min as a white solid. Purity > 95%by analytical C₁₈ HPLC (1.5 mL/min, 10% B for 5 min, then 10% to 90% B over 20 min, retention time 24.7 min). ¹H-NMR (CDCl₃), δ 8.04 (s, 1H), 7.76 (d, J = 7.1, 2H), 7.55 (m, 3H), 7.40 (m, 2H), 7.30 (m, 2H), 7.10 (app t, J = 8.1, 1H), 6.97 (app t, J = 7.4, 2H), 6.88 (s,

1H), 5.62 (m, 4H), 5.42(br s, 1H), 5.25 (m, 2H), 4.29 (d, J = 7.0, 1H), 4.16 (app t, J = 6.4, 1H), 4.11 (br s, 1H), 3.98 (m, 1H), 3.49 (m, 2H), 3.45-3.25 (m, 2H), 2.93 (m, 2H), 2.4-2.0 (m, 4H), 1.83 (m, 1H), 1.25 (s, 9H), 1.23 (s, 9H). ¹³C-NMR (CDCl₃) δ 177.2, 172.6, 156.1, 148.0, 144.0, 141.4, 136.3, 127.9, 127.3, 125.2, 122.2, 122.1, 120.3, 120.1, 119.4, 118.9, 113.4, 111.3, 83.0, 69.0, 67.0, 51.3, 48.0, 47.2, 40.4, 38.9, 33.6, 31.6, 27.0, 24.7, 23.9. ³¹P-NMR (CDCl₃) δ -3.93 (s). ESI-MS gave the molecular ion [M+H]⁺ m/z = 858.25; [M+Na]⁺ m/z = 880.37; [M+K]⁺ m/z = 896.47. HRMS calculated for C₄₆H₅₇N₃O₁₁P (MH⁺) m/z = 858.3731, found m/z = 858.3805.

Pin1 inhibition assay

Pin1 inhibition assay of **1** was performed as published.¹ The inhibitor **1** (10 μ L at concentrations of 800 μ M, 1.6, 2.4, 3.2, 4.0, 6.0, 8.0, 10.0 mM in 1:2 DMSO: H₂O) was preequilibrated in the 1.0 mL quartz cuvette at 4 °C for 10 min. For each concentration, the assay was performed in duplicate, and all of the data were used for calculation of IC₅₀. The plot of % inhibition vs. log[I] (μ M) produced a sigmoidal curve (SI Figure 1). The concentration of **1** for 50% inhibition of Pin1 activity (IC₅₀) was obtained by fitting all the experimental data to a dose response curve (95% confidence level) using equation (1) in TableCurve (version 3 for win32), where [I] is the inhibitor concentration (μ M).

%Inhibition =
$$a + \frac{b}{\{1 + (\log[I]/c)^d\}}$$
 (1)

A2780 cell based assay

Antiproliferative activity towards the A2780 human ovarian cancer cell line was measured as published.^{4, 5} The concentrations of **1** used were 190.8, 159.0, 95.4, 79.5, 39.7, 19.9, 9.9 μ M (duplicates), and the concentrations of **2** were 140.0, 116.7, 58.3, 29.2, 14.6, 7.3, 5.8 μ M (duplicates). The plot of % inhibition of proliferation activity against A2780 ovarian cancer cells vs. log[I] (μ M) produced a sigmoidal curve for each inhibitor (SI Figures 2 and 3). The concentrations of **1** and **2** for 50% inhibition of Pin1 activity (IC₅₀) were obtained by fitting all the experimental data to a dose response curve (95% confidence level) using equation (1) in TableCurve (version 3 for win32). Where [I] is the inhibitor concentration (μ M).



SI Figure 1. Dose response curve for inhibition of Pin1 by compound **1** (IC₅₀ = $28.3 \pm 2.1 \mu$ M). In the equation, a = 4.60, b = 113.29, c = 1.58, and d = 4.71; r² = 0.989. IC₅₀ = $28.3 \pm 2.1 \mu$ M.



SI Figure 2. Dose response curve for the inhibition of A2780 ovarian cancer cells proliferation activity of 1. For 1, from equation (1), a = 7.74, b = 94.51, c = 1.72, and d = 6.59; $r^2 = 0.996$. IC₅₀ = 46.2 ± 3.0 μ M.



SI Figure 3. Dose response curve for the inhibition of A2780 ovarian cancer cells proliferation activity of 2. For 2, from the equation, a = 11.04, b = 75.29, c = 1.41, and d = 6.35; $r^2 = 0.998$. IC₅₀ = $26.9 \pm 1.5 \mu$ M.



SI Figure 4. ³¹P-NMR study of the phosphorylation step. **a**: $(COCl)_2$ was added to **11**, followed by DMF; **b**: **11** was added very slowly to the mixture of $(COCl)_2$ and DMF. **10**: BisPOM phosphoryl chloride; **11**: bisPOM phosphate; **12**: $(POM)_4$ pyrophosphate; **2**: Fmoc-Ser $(PO(OPOM)_2)$ - $\Psi[(Z)CH=C]$ -Pro-(2)-N-(3)-ethylaminoindole.

Base	Temperature	Yield
Et ₃ N	-40 °C	< 20%
Et ₃ N	rt	< 20%
DIEA	rt	< 20%
collidine	rt	< 20%
NMM	rt	< 20%
Pyridine (8 equivalents)	-40 °C	20%
Pyridine (8 equivalents)	rt	25%
Pyridine (large excess)	-40 °C	22%
Pyridine (large excess)	rt	30%

SI Table 1. Yields for the phosphorylation step using different bases.

SI References

1. Wang, X. J.; Xu, B.; Mullins, A. B.; Neiler, F. K.; Etzkorn, F. A. J. Am. Chem. Soc. 2004, 126, 15533.

2. Wang, X. J.; Hart, S. A.; Xu, B.; Mason, M. D.; Goodell, J. R.; Etzkorn, F. A. *J. Org. Chem.* **2003**, *68*, 2343.

3. Hwang, Y.; Cole, P. A. Org Lett 2004, 6, 1555.

4. Abdel-Kader, M.; Berger, J. M.; Slebodnick, C.; Hoch, J.; Malone, S.; Wisse, J. H.; Werkhoven, M. C.; Mamber, S.; Kingston, D. G. *J. Nat. Prod.* **2002**, *65*, 11.

5. Kapustin, G.; Fejér, G.; Gronlund, J. L.; Seto, E.; McCafferty, D. G.; Etzkorn, F. A. *Org. Lett.* **2003**, *5*, 3053.

























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