Supporting Information

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Fig. S1. Insoluble inclusion bodies isolated from cells expressing LacS to different levels. Expression of LacS by *Escherichia coli* MC1061 cells was induced with the percentages of L-arabinose indicated above the panels. Molecular masses (in kDa) and positions of the marker proteins are indicated on the left of each panel. (*Left*) Coomassie-stained gel. (*Right*) Immunoblot decorated with anti-His tag antibody.



Fig. S2. Size-exclusion chromatography (SEC) on FC-12-solubilized cells. *E. coli* MC1061 cells were induced with 1×10^{-1} % (wt/vol) L-arabinose for 4 h at 25°C (LacS and LacY), and *E. coli* C43 cells were induced with 0.4 mM IPTG for 16 h at 17°C (GltP). Disrupted cells were solubilized with 1% (wt/vol) FC-12, and after ultracentrifugation the supernatant was analyzed by SEC. Samples taken before (–) and after (+) ultracentrifugation, and relevant fractions from the SEC were analyzed by *in gel* fluorescence and immunodetection using an anti-His tag antibody. Black and white arrows indicate the positions of nonfluorescent and fluorescent species of the GFP fusion proteins, respectively.



Fig. S3. Isolation of membrane vesicles. *E. coli* MC1061 cells were induced with 1×10^{-1} % (wt/vol) L-arabinose for 4 h at 25°C to allow expression of LacS. Cells were disrupted by two passes through a French pressure cell. (*Left*) The disrupted cells before centrifugation and the supernatant (LS supernatant) and pellet (LS pellet) after fractionation by low-speed centrifugation were analyzed. (*Right*) The supernatant of the low-speed centrifugation was further fractionated by sucrose density centrifugation. The top (low density; 5 ml) and bottom (high density; 1 ml) fractions of the sucrose gradient were analyzed.

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Table S1. Proteins analyzed in this study

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| Protein | Function | Source organism | Transporter family | Family nr. | Size, kDa |
|------------|--|-----------------|--------------------|------------|-----------|
| LacY | Lactose/H ⁺ symporter | E. coli | MFS | 2.A.1 | 46.5 |
| GltP | Glutamate/H ⁺ symporter | E. coli | DAACS | 2.A.23 | 47.2 |
| EcClC | Cl ⁻ /H ⁺ antiporter | E. coli | CIC | 2.A.49 | 50.3 |
| NhaA | Na ⁺ /H ⁺ antiporter | E. coli | NhaA | 2.A.33 | 41.4 |
| DctA | Dicarboxylate/H ⁺ symporter | E. coli | DAACS | 2.A.23 | 45.4 |
| LacS(∆IIA) | Lactose/H ⁺ symporter | S. thermophilus | GPH (MFS) | 2.A.2 | 52.3 |
| YdjN | Unknown | E. coli | DAACS | 2.A.23 | 48.7 |
| SstT | Serine/Na ⁺ symporter | E. coli | DAACS | 2.A.23 | 43.5 |
| GlpF | Glycerol facilitator | E. coli | MIP | 1.A.8 | 29.8 |

MFS, Major Facilitator Superfamily; DAACS, Dicarboxylate/Amino Acid:Cation (Na⁺ or H⁺) Symporter Family; ClC, Chloride Carrier/Channel Family; NhAA, Na⁺:H⁺ Antiporter (NhaA) Family; GPH, Glycoside-Pentoside-Hexuronide:Cation Symporter Family; MIP, Major Intrinsic Protein Family. Family names and numbers are derived from the Transport Classification Database [Saier MH, Jr (1988) Molecular phylogeny as a basis for the classification of transport proteins from bacteria, archaea, and eukarya. *Adv Micro Physiol* 40:81–136].