

Supporting Information

Feng *et al.* 10.1073/pnas.0711961105

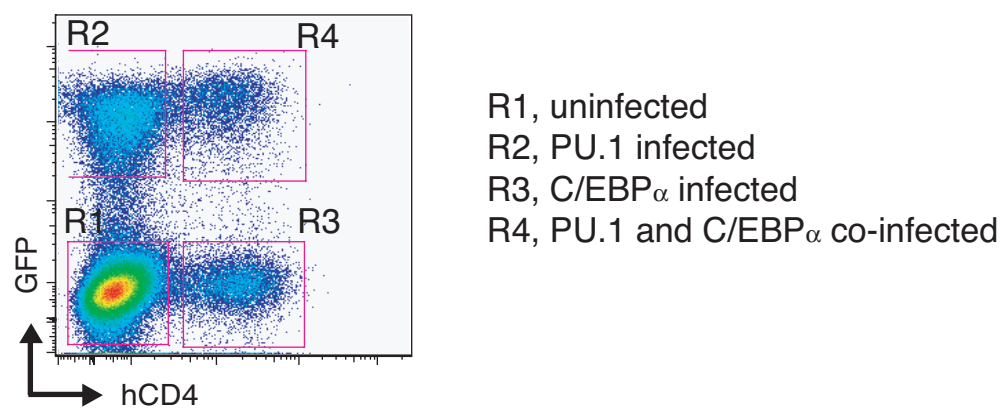


Fig. S1. FACS plot of NIH 3T3 cells infected with PU.1-GFP and C/EBP α viruses for 7 days. The gates show uninfected, singly infected, and doubly infected cells.

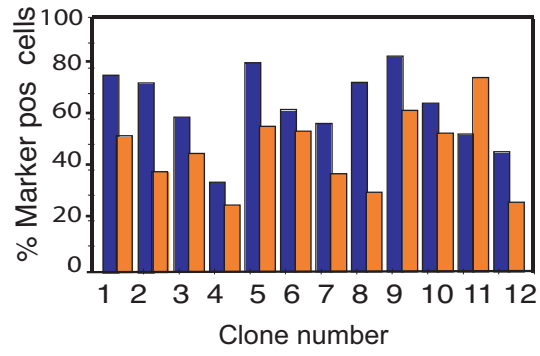


Fig. S3. Experiment showing that subclones of NIH 3T3 cells are susceptible to transcription factor-induced conversion. Shown is Mac-1 (blue bars) and CD45 (orange bars) expression in NIH 3T3 subclones coexpressing PU.1 and C/EBP α viruses 7 days after infection. None of the subclones expressed Mac-1 or CD45 before infection.

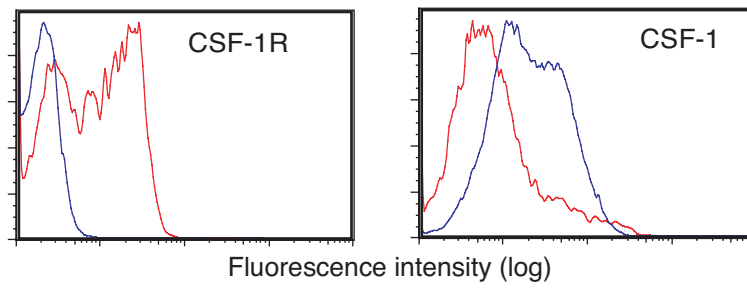


Fig. 57. FACS analysis of CSF-1R and CSF-1. Blue lines, 3T3 cells; red lines, PC2.3 cells.

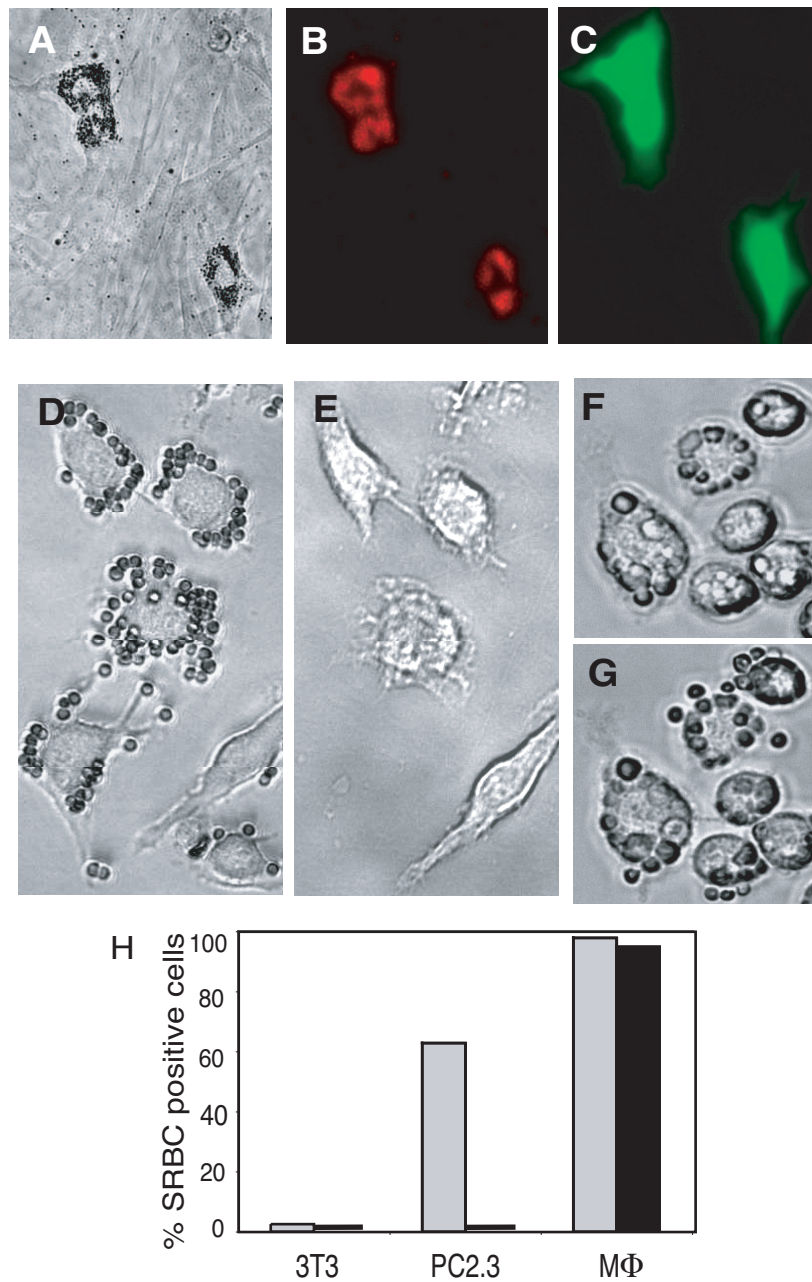


Fig. 58. Phagocytic capacity of PC2.3 cells. (A–C) Brightfield and fluorescent images of primary mouse embryo fibroblasts infected with PU.1-EGFP and C/EBPa-hCD4 viruses for 11 days, incubated with red fluorescent beads, and washed. (B) Red fluorescent beads. (C) GFP fluorescence. (D–G) phagocytosis of sheep red blood cells (SRBCs). (D) PC2.3 cells incubated with antibody coated SRBCs, showing formation of rosettes. (E) Cells in D after treatment with lysis buffer to remove noninternalized SRBCs. (F and G) BAC macrophages incubated with opsonized SRBCs before and after lysis, respectively. (H) Quantification of the data, with the percentage of rosette forming cells in gray and cells that ingested SRBCs in black.

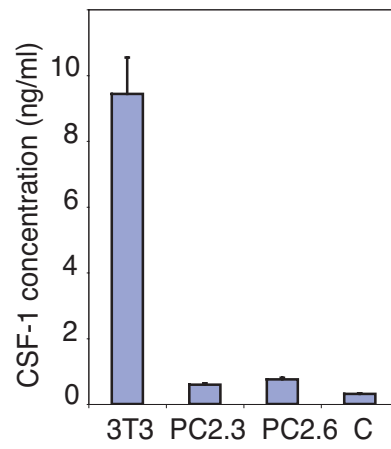


Fig. S10. CSF-1 production. NIH 3T3 cells and PC2.3 and BAC macrophages were washed and incubated overnight with growth medium, and the CSF-1 concentration in the supernatants was tested by RIA. Bars indicate standard deviation of triplicate samples.

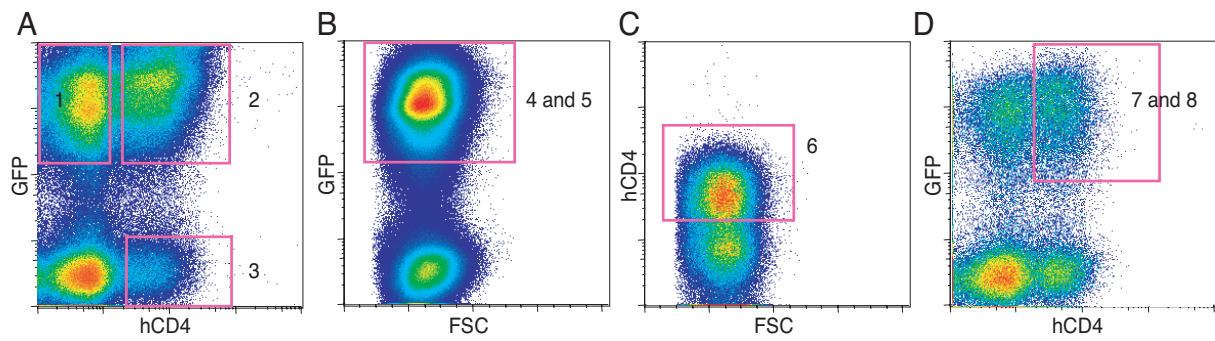


Fig. S11. (A) FACS plots of NIH 3T3 cells infected for 4 days with a combination of control vectors (A), PU.1 alone (B), C/EBP α alone (C), and a combination of PU.1 and C/EBP α (D). The numbers next to the red squares indicate gates used to sort fractions for isolation of mRNA analyzed in Fig. 5D (profiles of fraction 5 and 8 cells are not shown). FSC, forward scatter.

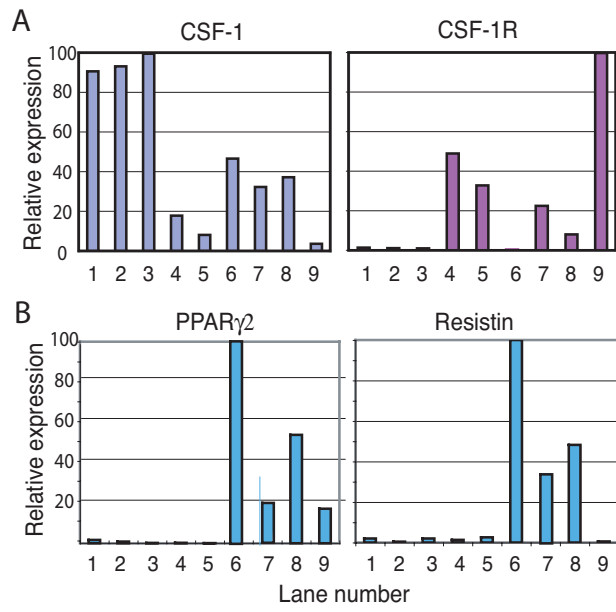


Fig. S12. Quantification of the data shown in Fig. 5D. Gels were scanned by densitometry. Values were plotted relative to the highest number obtained.

Table S1. Number of genes that are up or down-regulated at least 2-fold in BAC1.2F5 macrophages relative to NIH 3T3 cells

Variables	Up-regulated in BAC MΦ	Down-regulated in BAC MΦ
No. of genes	960	1,052
Fold change	5.3	0.34
No. of genes also changed in PC2.3 cells	620	400
Fold change	3.9	0.38

Comparisons of relative gene expression, based on two-color cDNA array analysis, were made for BAC macrophages versus 3T3 cells. The table shows the number of genes that are up- or down-regulated at least 2-fold in BAC1.2F5 macrophages relative to NIH 3T3 cells. The table also indicates the average fold expression changes.

Table S2. Number of genes that are up or down-regulated at least 2-fold in PC2.3 cells relative to NIH 3T3 cells

Variables	Up-regulated in PC2.3 cells	Down-regulated in PC2.3 cells
No. of genes	954	400
Fold change	5.3	0.38
No. of genes also changed in BAC cells	597	231
Fold change	5.6	0.49

Comparisons of relative gene expression, based on two-color cDNA array analysis, were made for PC2.3 cells versus 3T3 cells. The table shows the number of genes that are up- or down-regulated at least 2-fold in PC2.3 cells relative to NIH 3T3 cells. The table also indicates the average fold expression changes.