Marker		Primers	Fragment Size (bp)	Restriction enzyme
T0337	For	5′-TGCAGTCCAATCAAATGTCATGTCCA-3′	983	N/A***
	Rev	5′-GTATCTCATGGTGAATGCACTTGTCGTA-3′		
T0487	For	5′-GACAGCGTGAAATGATTCTCGAGGCA-3′	996	HhaI
	Rev	5′-GCTGCCACTATTGAGGTACGCAACAAC-3′		
CT265	For	5′-GCGAGGCAACACTATTCACTTGATGT-3′	959	BstXI
	Rev	5′-GGAAGAACTGGTCATCTTTGGGCGCA-3′		
20K14T7	For	5′-GGGTCCACTCAGAGATGCAACTTCC-3′	716	HinFI
	Rev	5′-GAACGTTACTAGAGTATCCAAGGAA-3′		
CT148	For	5′-GTACAGCCTAGTAAGTAACACGC-3′	830	HinDIII
	Rev	5′-GCATCATGAATGTGAATGATACAGA-3′		
T1581	For	5′-GCTACTAGAAGAACCTAGTCAGC-3′	470	αTaqI
	Rev	5′-GTAATCGCCGGTGTCGTAAGCAGC-3′		
C_2At5g47390	For	5′-TGGTGGCTCTGTTGATGGTTATGC-3′	1202	αTaqI
	Rev	5′-ACATCCTATGCTCCTCCTCAGTCC-3′		

Table S1. Development of PCR based markers flanking the *gf* locus.

RFLP markers were converted to PCR based markers using the forward (For) and reverse (Rev) primers. Following DNA sequence analysis of cloned fragments from the parents of our F2 mapping population, *S. lycopersicum* (*gf/gf*) and the *S. pennellii* introgression line (IL8-3) restriction enzymes were chosen that yielded polymorphic fragments between the parents that could be utilized as genetic markers. Fragment sizes are given for amplification products from *S. lycopersicum*.

***For T0337 the amplification product from IL8-3 was approximately 150 bp larger than from *S. lycopersicum* allowing detection of the polymorphic fragments by agarose gel electrophoresis without a restriction enzyme digest.