Isolation and Genetic Analysis of Amber *uvrA* and *uvrB* Mutants

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Genetic properties of amber uvrA and uvrB mutants of *Escherichia coli* K-12 are described. The isolation of three amber uvrA and two amber uvrB mutants indicates that the products of these genes are proteins.

The genes uvrA, -B, and -C are involved in releasing pyrimidine dimers from ultraviolet (UV)-irradiated deoxyribonucleic acid. Recently it has been reported that the *uvrA* and *uvrB* genes affect the activity of an endonuclease specific for UV-irradiated deoxyribonucleic acid (1, 16) and that in the *uvrC* mutant strain the number of nicks formed by incising UVirradiated deoxyribonucleic acid is increased slowly with time after irradiation (7). It is, however, not known yet whether the products of the *uvr* genes are protein or ribonucleic acid, although Kondo et al. (8) suggested that the product of the *uvrB* gene was ribonucleic acid, or a protein whose synthesis is hardly inhibited by chloramphenicol. To answer this question, we have tried to isolate amber mutations in the uvr genes.

The methods for isolating UV-sensitive mutants were essentially the same as those described by Howard-Flanders and Theriot (5). To obtain a uvr amber mutation, a derivative of strain KN250 carrying a temperature-sensitive amber suppressor, sup-126 (11), and amber trpand tyr mutations was used. A log-phase culture was treated with 0.5% ethyl methane sulfonate for 1 h at 37 C and washed to remove ethyl methane sulfonate. The cells were resuspended in fresh medium, divided into many tubes, cultured to stationary phase, and then plated on NBT (nutrient broth + 50 μ g of thymine per ml) agar plates together with about 10^9 T3 or λ phages irradiated with 1,000 ergs of UV light per mm². Colonies from the survivors were selected, purified successively by single colony isolation, and tested for sensitivity to UV by irradiating with appropriate doses and incubating at both 30 and 42 C. The ability to reactivate UV-irradiated bacteriophage was also examined. Out of 133 UV-sensitive strains isolated, 94 mutants showed the Hcr⁻ phenotype. Among these Hcr⁻ mutants, 54 independent mutants were classified into uvrA, -B, and -C groups by examining UV sensitivity of heterodiploid strains constructed by transferring F112 (9), F450 (2), and F4102 (10) carrying the $uvrA^+$, $-B^+$, and $-C^+$ genes, respectively. The uvrD mutation (12) was identified by examining the Hcr phenotype, X-ray sensitivity, and the UV sensitivity of Arg⁺ recombinants after conjugation with Hfr J4 (6) for 30 min. Results are shown in Fig. 1. In these uvr^- strains three uvrA (uvr-24, uvr-43, and uvr-72) and two uvrB mutations (uvr-46 and uvr-59), shown as underlined numbers in Fig. 1, were concluded to be amber mutations by two different types of experiments (Table 1).

First, upon transferring plasmid KHF6 carrying $supD^-$ into these mutants they became UV^R, Trp⁺, and Tyr⁺ simultaneously, whereas upon transferring plasmid KHF17 not carrying $supD^-$ they remained as they were.



FIG. 1. Location of uvr mutations on the linkage map of Escherichia coli K-12 (15). Isolation and genetic mapping of uvr mutations were described in the text. Underlined numbers represent amber uvr mutations. The arrows represent origin and direction of transfer of F-prime factors and an Hfr strain used in the experiments. The approximate extent of the chromosomal segments carried by the various F-prime factors are indicated by their length and position.

530 NOTES

J. BACTERIOL.

Secondly, when the spontaneous back mutants from Trp amber to Trp prototroph or from Tyr amber to Tyr prototroph were isolated, they also showed the $UV^{R}Tyr^{+}$ or $UV^{R}Trp^{+}$ phenotype, respectively, an indication that the three phenomena, from Trp and Tyr amber to Trp and Tyr prototroph and from UV^{S} to UV^{R} , were reversed by a single mutation, namely, an amber suppressor mutation. Note that under these

conditions the spontaneous reversion frequency ranged from 10^{-6} to 10^{-8} .

The mutation sites of these amber $uvr^$ strains were analyzed by complementation and by P1 transduction. The F-prime factor, F112 carrying $uvrA^+$ or uvrA6 (13), was transferred to strains EA512 (uvr-24), EA1803 (uvr-43), and EA3601 (uvr-72). The result with strain EA3601 is shown in Fig. 2 (a and b). The mero-

Strain no.	uvr	Hcr	KHF17 trans- ferred		KHF6 trans- ferred			Trp ⁺ back mutants			Tyr ⁺ back mutants			
			UV	Trp	Tyr	υv	Trp	Tyr	Frequency	UV ^R Tyr ⁺	UV ^s Tyr ^{- a}	Frequency	UV¤ Trp⁺	UV ^s Trp ⁻ "
EA512	A24	-	s	-	-	R	+	+	2.1×10^{-7}	58	5	2.7 × 10 ⁻⁷	55	8
EA1803	A43	-	S	-	-	R	+	+	9.3 × 10 ⁻⁹	7	0	9.3 × 10 ⁻⁹	8	0
EA3601	A72	-	S	-	- 1	R	+	+	3.0×10^{-7}	27	0	_0		
										(Tyr ⁻)				
EA2401	B46	-	s	- 1	_	R	+	+	3.1×10^{-7}	61	0	2.9×10^{-7}	64	0
EA3103	B59	. –	s	-	-	R	+	+	1.5×10^{-6}	59	0	1.7×10^{-6}	64	0

TABLE 1. Characterization of UV-sensitive amber mutants

^a The back mutants UV^sTyr⁻ and UV^sTrp⁻ seem to be the true backs of amber *trp* and amber *tyr*, respectively. ^b Since EA3601 had acquired another mutation besides amber in *tyr*, Tyr⁺ back mutants could not be obtained.



FIG. 2. (a) UV survival curves of merodiploid strains F112 (uvr⁺)/EA3601 (\bigcirc , $\textcircled{\bullet}$) and F112 (uvrA6)/ EA3601 (\triangle , \clubsuit) at 30 (open symbols) and 42 C (closed symbols). Overnight cultures in a minimal medium were diluted into M9 buffer to a density of about 10⁷ cells/ml, irradiated with various doses of UV, and plated on NBT agar at 30 and 42 C. The fraction surviving was determined from the number of visible colonies after incubation overnight at 42 C or for 2 days at 30 C. (b) Host-cell reactivation of λ_{vir} by F112 (uvr⁺)/EA3601 (\bigcirc), F112 (uvrA6)/EA3601 (\bigcirc), EA3601 (\square), a wild-type strain (\triangle), and AB1886 (\blacktriangle).

diploid strain carrying F112 ($uvrA^+$) became as resistant as a wild-type strain and showed a host-cell reactivation of UV-irradiated bacteriophage λ , whereas the merodiploid carrying F112 (uvrA6) remained as sensitive as a parent strain and showed the Hcr⁻ phenotype. Similar results were also obtained for the other two



FIG. 3. Host-cell reactivation of λ_{vir} (open symbols) and $\lambda b2$ bio⁺uvrB⁺ (closed symbols) by various uvrB mutants. Symbols: Wild-type strain (\diamond , \blacklozenge); E54 (su⁻uvrB46) (\triangle , \blacktriangle); E95 (su⁻uvrB59) (\bigcirc , \blacklozenge); and AB1885 (uvrB5) (\Box , \blacksquare).

amber mutants. This information demonstrates that the mutation occurs in the *uvrA* gene in these three amber mutants. When the *sup*⁺ derivatives of strains EA2401 (*uvr-46*) and EA3103 (*uvr-59*) as well as strain AB1885 (4) were tested for the Hcr of λ and λ b2 bio⁺*uvrB*⁺, they reactivated much more UV λ b2 bio⁺*uvrB*⁺ than λ bacteriophage (Fig. 3). Then the mutation site *uvr-59* was mapped by P1 transduction (Table 2); the genes *uvr-59* and *uvrB5* of strain AB1885 were linked to gal at frequencies of 38 and 31%, respectively. Hence, mutations *uvr-59* and *uvr-46* are concluded to be *uvrB* mutations.

The effect of temperature-sensitive suppressor sup-126 on the UV sensitivity and on a hostcell reactivation of amber uvr^- mutants was also examined (Table 3). All these amber mutants showed a higher survival after UV irradiation on agar plates at 30 C than after that at 42 C and partially reactivated UV-irradiated bacteriophage λ when in the presence of sup-126, whereas in substituting sup-126 with $supD^+$ by conjugation they became all extremely sensitive to UV even at 30 C and showed the Hcr⁻ phenotype.

Previously we have reported the isolation of a temperature-sensitive uvrA mutant (13, 14). This information and the isolation of amber uvrA mutants presented in this communication demonstrate that the product of the uvrA gene is a protein. Similarly, the isolation of amber uvrB mutants also indicates that the product of the uvrB gene is a protein.

In view of the fact that only three out of 24 uvrA mutants and only two out of 19 uvrB mutants carried the amber lesion, the eight uvrC and three uvrD mutants examined might

Donor	Recipient	No. of Gal ⁺ transductants	Frequency of unselected marker (%)		
		scored	URR	UVs	
W3876 (gal ⁺ uvrB ⁺)	E50 (gal ⁻ uvr-59)	111	38	62	
W3876 (gal+ uvrB+)	AB1885 (gal ⁻ uvrB5)	94	31	69	

TABLE 2. Co-transduction of uvrB with gal by P1kc phage

TABLE 3. Effect of a temperature-sensitive amber suppressor sup-126 on UV survival and on Hcr of λ phage

	Surv	vival after irradia	Survival of λ irradiated with 600				
Mutations	sup	-126	SI	u ⁻	ergs/mm ²		
		42 C	30 C	42 C	sup-126	su⁻	
uvrA24	4.0×10^{-4}	1.2×10^{-5}	3.8×10^{-5}	2.7×10^{-4}	6.7×10^{-4}	7.8×10^{-5}	
uvrA43	$2.1 imes 10^{-2}$	8.2×10^{-5}	$7.5 imes 10^{-6}$	1.4×10^{-4}	4.8×10^{-3}	1.1×10^{-5}	
uvrA72	8.5×10^{-5}	1.0×10^{-5}	8.6×10^{-6}	$1.5 imes 10^{-5}$	$2.3 imes10^{-3}$	3.0×10^{-5}	
uvrB46	3.0×10^{-2}	4.6×10^{-4}	6.8×10^{-4}	$1.5 imes 10^{-4}$	2.3×10^{-2}	1.7×10^{-4}	
uvrB59	$2.0 imes 10^{-2}$	4.8×10^{-3}	5.5×10^{-5}	2.8×10^{-4}	1.6×10^{-2}	1.3×10^{-4}	

be not a large enough group to obtain an amber mutation.

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LITERATURE CITED

- Braun, A., and L. Grossman. 1974. An endonuclease from *Escherichia coli* that acts preferentially on UVirradiated DNA and is absent from the *uvrA* and *uvrB* mutants. Proc. Natl. Acad. Sci. U.S.A. 71:1838-1842.
- Freifelder, D. 1968. Studies on *Escherichia coli* sex factors. IV. Molecular weight of the DNA of several F' elements. J. Mol. Biol. 35:95-102.
- Horiuchi, T., and T. Nagata. 1973. Mutations affecting growth of the *Escherichia coli* cell under a condition of DNA polymerase l-deficiency. Mol. Gen. Genet. 123:89-110.
- Howard-Flanders, P., R. P. Boyce, and L. Theriot. 1966. Three loci in *Escherichia coli* K-12 that control the excision of pyrimidine dimers and certain other mutagenic products from DNA. Genetics 53:1119-1136.
- Howard-Flanders, P. and L. Theriot. 1962. A method for selecting radiation-sensitive mutants of *Escherichia coli*. Genetics 47:1219-1224.
- Jacob, F., and E. L. Wollman. 1957. Analyse des groupes de liason génétique de différentes souche donatrice d'*Escherichia coli* K-12. C. R. Acad. Sci. 245:1840-1843.

- Kato, T. 1972. Excision repair characteristics of recB⁻res⁻ and uvrC⁻ strains of Escherichia coli. J. Bacteriol. 112:1237-1246.
- Kondo, S., H. Ichikawa, and T. Kato. 1969. Genetic reactivation of DNA damage in UV-sensitive strains of *Escherichia coli*. Jpn. J. Genet. 44(Suppl. 2):47-48.
- 9. Low, K. B. 1972. Escherichia coli K-12 F-prime factors, old and new. Bacteriol. Rev. 36:587-607.
- Marinus, M. G. 1973. Location of DNA methylation genes on the *Escherichia coli* K-12 genetic map. Mol. Gen. Genet. 127:47-55.
- Nagata, T., and T. Horiuchi. 1973. Isolation and characterization of a temperature-sensitive amber suppressor mutant of *Escherichia coli* K-12. Mol. Gen. Genet. 123:77-88.
- Ogawa, H., K. Shimada, and J. Tomizawa. 1968. Studies on radiation-sensitive mutants of *E. coli*. I. Mutants defective in the repair synthesis. Mol. Gen. Genet. 101:227-244.
- Shimazu, Y., M. Morimyo, and K. Suzuki. 1975. Genetic analysis of ultraviolet sensitive mutants of *E. coli* K-12 URT-43 (*urt-1*) and KMBL99 dar3 carrying temperature sensitive mutations at the locus *uvrA*. Mutat. Res. 30:1-8.
- Suzuki, K., E. Saito, and M. Morimyo. 1969. A mutant of *Escherichia coli* exhibiting varying ultraviolet sensitivities depending on the temperature of incubation after irradiation. Photochem. Photobiol. 9:259-272.
- Taylor, A. L., and C. D. Trotter. 1972. Linkage map of Escherichia coli strain K-12. Bacteriol. Rev. 36:504– 524.
- Waldstein, E. A., R. Sharm, and R. Ben-Ishai. 1974. Role of ATP in excision repair of ultraviolet radiation damage in *Escherichia coli*. Proc. Natl. Acad. Sci. U.S.A. 71:2651-2654.