

## Supporting information

### **Silver(I) Tetrafluoroborate as a Potent Promoter for Chemical Glycosylation**

Sophon Kaeothip, Papapida Pornsuriyasak, and Alexei V. Demchenko\*

*Department of Chemistry and Biochemistry, University of Missouri – St. Louis, One  
University Boulevard, St. Louis, MO 63121, USA*

*demchenkoa@umsl.edu*

#### Contents:

S-2	General methods
S-3/S-5	Transformation of the STaz moiety into other leaving groups: synthesis of glycosyl donors, Synthesis of <b>1</b> , <b>3a</b> , <b>4a</b> , <b>6</b> , <b>8-11</b>
S-6/S-9	General Glycosylation Procedures: Synthesis of O-Glycosides and Oligosaccharides
S-10/S-11	References
S-12/S-13	Spectra of the trisaccharide <b>25</b>
S14/S-24	<sup>1</sup> H Spectra of known compounds

**General Methods.** Column chromatography was performed on silica gel 60 (70-230 mesh), reactions were monitored by TLC on Kieselgel 60 F<sub>254</sub>. The compounds were detected by examination under UV light and by charring with 10% sulfuric acid in methanol. Solvents were removed under reduced pressure at <40 °C. Dichloromethane and 1,2-dichloroethane were distilled from CaH<sub>2</sub> directly prior to application. Methanol was dried by refluxing with magnesium methoxide, distilled and stored under argon. Pyridine was dried by refluxing with CaH<sub>2</sub> and then distilled and stored over molecular sieves (3Å). Molecular sieves (3Å), used for reactions, were crushed and activated *in vacuo* at 390 °C during 8 h in the first instance and then for 2-3 h at 390 °C directly prior to application. AgBF<sub>4</sub> was used as received. Optical rotations were measured at 'Jasco P-1020' polarimeter. <sup>1</sup>H-NMR spectra were recorded at 500MHz, <sup>13</sup>C-NMR spectra were recorded at 125MHz. HRMS determinations were made with the use of JEOL MStation (JMS-700) Mass Spectrometer.

**Transformation of the STaz moiety into other leaving groups: synthesis of glycosyl donors**

**1-O-Acetyl-2,3,4,6-tetra-O-benzyl-D-glucopyranose (1).** Pd(OAc)<sub>2</sub> (35 mg, 0.156 mmol) was added to a mixture of 2-thiazoliny 2,3,4,6-tetra-O-benzyl-1-thio-β-D-glucopyranoside (**5a**, 50 mg, 0.078 mmol) and activated molecular sieves 3Å (150 mg) in dry (ClCH<sub>2</sub>)<sub>2</sub> (0.5 mL). The reaction mixture was stirred under argon for 24 h at 55 °C. Upon completion, the solid was filtered-off and the filtrate was concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (ethyl acetate/ hexane gradient elution) to afford 1-O-acetate **1** as colorless syrup in 72% yield (α/β = 1.6/1). Analytical data for **1** were essentially the same as reported previously.<sup>1</sup>

**2-Benzoxazolyl 2,3,4,6-tetra-O-benzyl-1-thio-β-D-glucopyranoside (3a).** A mixture of **5a** (50 mg, 0.078 mmol) and activated molecular sieves 3Å (40 mg) in dry CH<sub>2</sub>Cl<sub>2</sub> (1.2 mL) was stirred under argon for 1 h. Freshly prepared solution of Br<sub>2</sub> in dry CH<sub>2</sub>Cl<sub>2</sub> (0.75 mL, 1/165, v/v) was then added dropwise over the period of 5 min at rt. Quantitative TLC estimates were made based on the accumulation of 2,3,4,6-tetra-O-benzyl-α-D-glucopyranosyl bromide **2a**. After that, the solvent was evaporated under the reduced pressure at rt. The residue containing crude **2a** was then treated dissolved in dry acetone (0.5 mL) and potassium salt (KSBox,<sup>2</sup> 22 mg, 0.117 mmol) and 18-crown-6 (4 mg, 0.012 mmol) were added. The reaction mixture was stirred under argon for 1 h at rt. After that, the mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> (20 mL) and washed with water (10 mL), 1% aq. NaOH (10 mL) and water (3 x 10 mL), the organic phase was separated, dried, and concentrated *in vacuo*. The residue was purified by silica gel column chromatography (ethyl acetate/hexane gradient elution) to afford the title compound **3a** as a white foam in 82% yield. Analytical data for **3a** were essentially the same as reported previously.<sup>2</sup>

**2-Benzothiazolyl 2,3,4,6-tetra-O-benzyl-1-thio-β-D-glucopyranoside (4a).** Crude glycosyl bromide **2a** (~48 mg, 0.078 mmol), obtained as described for the synthesis of **3a**, was dissolved in dry acetone (0.5 mL) and potassium salt of 2-mercaptobenzotriazole<sup>3</sup> (24 mg, 0.117 mmol) and 18-crown-6 (4 mg, 0.012 mmol) were added. The reaction mixture was stirred under argon for 1 h at rt, then diluted with CH<sub>2</sub>Cl<sub>2</sub> (20 mL) and washed with water (10 mL), 1% aq. NaOH (10 mL), and water (3 x 10 mL). The organic phase was separated, dried, and concentrated *in vacuo*. The residue was purified by silica gel column chromatography (ethyl acetate/hexane gradient elution) to afford the title compound **4a** as a white foam in 75% yield. Analytical data for **4a** were essentially the same as reported previously.<sup>4</sup>

**Ethyl 2,3,4,6-tetra-O-benzyl-1-thio- $\beta$ -D-glucopyranoside (6).** A mixture of ethane thiol (17  $\mu$ L, 0.225 mmol) and sodium hydride (1.0 mg, 0.025 mmol) was added to a solution of **2a** (~95 mg, 0.15 mmol), prepared as described for the synthesis of **3a**, in dry  $\text{CH}_2\text{Cl}_2$  (1.0 mL) at 0  $^\circ\text{C}$  and the resulting mixture was stirred under argon for 1 h at rt. The reaction mixture was then diluted with  $\text{CH}_2\text{Cl}_2$  (20 mL) and washed with water (10 mL), 20% aq.  $\text{NaHCO}_3$  (10 mL), and water (3 x 10 mL), the organic phase was separated, dried, and concentrated *in vacuo*. The residue was purified by silica gel column chromatography (ethyl acetate/hexane gradient elution) to afford the title thioglycoside **6** as a white foam in 92% yield. Analytical data for **6** were essentially the same as reported previously.<sup>5</sup>

**2,3,4,6-Tetra-O-benzyl- $\alpha$ -D-glucopyranosyl chloride (8).** To a stirred solution of **5a** (100 mg, 0.15 mmol) and dimethylformamide (6  $\mu$ L, 0.075 mmol) in dry  $\text{CH}_2\text{Cl}_2$  (1.2 mL) under argon thionyl chloride (34  $\mu$ L, 0.45 mmol) was added dropwise. The reaction mixture was kept for 1 h and then concentrated under the reduced pressure. The residue was purified by short-path silica gel column chromatography (ethyl acetate/hexane gradient elution) to afford the title compound **8** as a colorless syrup in 84% yield. Analytical data for **8** were essentially the same as reported previously.<sup>6</sup>

**2,3,4,6-Tetra-O-benzyl-D-glucopyranosyl fluoride (9).** To a stirred solution of **5a** (100 mg, 0.15 mmol) in THF (1.2 mL) diethylaminosulfur trifluoride (DAST, 72  $\mu$ L, 0.55 mmol) was added at -30  $^\circ\text{C}$  under argon. The external cooling was then removed and the reaction mixture was stirred for 24 h at rt. After that, the solution was cooled to -30  $^\circ\text{C}$  and methanol (0.1 mL) was added, the resulting mixture was warmed to rt and the volatiles were evaporated off *in vacuo*. The residue was dissolved in  $\text{CH}_2\text{Cl}_2$  (20 mL) and washed successively with water (10 mL), 20% aq.  $\text{NaHCO}_3$  (10 mL), and water (3 x 10 mL). The

organic phase was separated, dried, and concentrated *in vacuo*. The residue was purified by short-path silica gel column chromatography (ethyl acetate/hexane gradient elution) to afford the title fluoride **9** as a white foam in 88% yield ( $\alpha/\beta = 1/2.8$ ). Analytical data for **9** were essentially the same as reported previously.<sup>7,8</sup>

**2,3,4,6-Tetra-O-benzyl-D-glucopyranose (10)**. A solution of **5a** (65 mg, 0.10 mmol) and N-bromosuccinimide (45 mg, 0.20 mmol) in acetone/water (1.5 mL, 9/1, v/v) was stirred for 16 h at rt. The reaction mixture was then diluted with CH<sub>2</sub>Cl<sub>2</sub> (20 mL) and washed with water (10 mL), 20% aq. NaHCO<sub>3</sub> (10 mL), and water (3 x 10 mL). The organic phase was separated, dried, and concentrated *in vacuo*. The residue was purified by silica gel column chromatography (ethyl acetate/hexane gradient elution) to afford the title hemiacetal **10** as a colorless syrup in 90% yield ( $\alpha/\beta = 1/1$ ). Analytical data for **10** were essentially the same as for the commercial sample.

**2,3,4,6-Tetra-O-benzyl- $\alpha$ -D-glucopyranosyl trichloroacetimidate (11)**. NaH (8 mg, 0.37 mmol) was added portionwise to a stirred solution of compound **10** (100 mg, 0.185 mmol) and trichloroacetonitrile (85  $\mu$ L, 0.84 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (2 mL). After stirring for 3 h at rt, the reaction mixture was concentrated under the reduced pressure. The residue was purified by column chromatography on silica gel (ethyl acetate/ hexane gradient elution) to afford the title compound **11** as a colorless syrup in 82% yield. Analytical data for **11** were essentially the same as reported previously.<sup>9</sup>

## **General Glycosylation Procedures: Synthesis of O-Glycosides and Oligosaccharides.**

*Method A – Activation with AgBF<sub>4</sub>. (see the article)*

*Method B – Activation of glycosyl donor 1 with BF<sub>3</sub>·Et<sub>2</sub>O.* A mixture containing the glycosyl donor (0.11 mmol), glycosyl acceptor (0.10 mmol), and freshly activated molecular sieves (3 Å, 200 mg) in (ClCH<sub>2</sub>)<sub>2</sub> (2 mL) was stirred under argon for 1.5 h. BF<sub>3</sub>·Et<sub>2</sub>O (40 µL, 0.33 mmol) was added and the reaction mixture was stirred for 16 h at rt. It was then diluted with CH<sub>2</sub>Cl<sub>2</sub>, the solid was filtered-off and the residue was washed with CH<sub>2</sub>Cl<sub>2</sub>. The combined filtrate (30 mL) was washed with 20% aq. NaHCO<sub>3</sub> (10 mL) and water (3 x 10 mL). The organic phase was separated, dried with MgSO<sub>4</sub>, and concentrated in vacuo. The residue was purified by column chromatography on silica gel (ethyl acetate/hexane gradient elution) to afford a disaccharide derivative.

*Method C- Activation of 1 with BF<sub>3</sub>·Et<sub>2</sub>O/AgBF<sub>4</sub>.* A mixture of the glycosyl donor (0.11 mmol), glycosyl acceptor (0.10 mmol), and freshly activated molecular sieves (3 Å, 200 mg) in (ClCH<sub>2</sub>)<sub>2</sub> (2 mL) was stirred under argon for 1.5 h. BF<sub>3</sub>·Et<sub>2</sub>O (40 µL, 0.33 mmol) and AgBF<sub>4</sub> (10 mg, 0.05 mmol) were added and the reaction mixture was stirred for 5 h at rt. Upon completion, it was diluted with CH<sub>2</sub>Cl<sub>2</sub>, the solid was filtered-off and the residue was washed with CH<sub>2</sub>Cl<sub>2</sub>. The combined filtrate (30 mL) was washed with 20% aq. NaHCO<sub>3</sub> (10 mL) and water (3 x 10 mL). The organic phase was separated, dried with MgSO<sub>4</sub>, and concentrated in vacuo. The residue was purified by column chromatography on silica gel (ethyl acetate/hexane gradient elution) to afford a disaccharide derivative.

*Method D - Activation of 6 and 7 with NIS.* A mixture of the glycosyl donor (0.11 mmol), glycosyl acceptor (0.10 mmol), and freshly activated molecular sieves (3 Å, 200 mg) in

(ClCH<sub>2</sub>)<sub>2</sub> (2 mL) was stirred for 1.5 h under argon. NIS (50 mg, 0.22 mmol) was then added and the reaction mixture was stirred for 24 h at rt. Upon completion, the solid was filtered-off and the residue was washed with CH<sub>2</sub>Cl<sub>2</sub>. The combined filtrate (30 mL) was washed with 20% aq. Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> (10 mL) and water (3 x 10 mL). The organic phase was separated, dried over MgSO<sub>4</sub>, and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (ethyl acetate/ hexane gradient elution) to afford a disaccharide derivative.

*Method E - Activation of 6 and 7 with NIS/AgBF<sub>4</sub>.* A mixture of the glycosyl donor (0.11 mmol), glycosyl acceptor (0.10 mmol), and freshly activated molecular sieves (3 Å, 200 mg) in (ClCH<sub>2</sub>)<sub>2</sub> (2 mL) was stirred for 1.5 h under argon. NIS (50 mg, 0.22 mmol) and AgBF<sub>4</sub> (10 mg, 0.05 mmol) were added and the reaction mixture was stirred for 10-20 min at rt. Upon completion, the solid was filtered-off and the residue was washed with CH<sub>2</sub>Cl<sub>2</sub>. The combined filtrate (30 mL) was washed with 20% aq. Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> (10 mL) and water (3 x 10 mL). The organic phase was separated, dried over MgSO<sub>4</sub>, and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (ethyl acetate/ hexane gradient elution) to afford a disaccharide derivative.

**Pent-4-en-1-yl 2,3,4,6-tetra-O-benzyl-1-thio-β-D-glucopyranoside (7).** The title compound was synthesized from glycosyl bromide **2a**, obtained as described for the synthesis of **3a**, and 4-penten-1-ol by Method A in 89% yield as a white foam. Analytical data for **7** were essentially the same as reported previously.<sup>10</sup>

**Methyl 4-O-(2,3,4,6-tetra-O-benzyl-D-glucopyranosyl)-2,3,6-tri-O-benzyl-α-D-glucopyranoside (16a).** The title compound was obtained by Method A from **8** and **12**<sup>11</sup> in

91% yield ( $\alpha/\beta = 1/3.4$ ) or **11** and **12** in 65% yield ( $\alpha/\beta = 1/5.4$ ). Analytical data for **16a** were essentially the same as reported previously.<sup>12</sup>

**Methyl 6-O-(2,3,4,6-tetra-O-benzyl-D-glucopyranosyl)-2,3,4-tri-O-benzyl- $\alpha$ -D-glucopyranoside (17a).** The title compound was obtained by Method A from glycosyl donors **2a**, **3a**, **5a**, **8**, **9** or **11** and glycosyl acceptor **13**<sup>13</sup> in 92% ( $\alpha/\beta = 2.4/1$ ), 89% ( $\alpha/\beta = 1.2/1$ ), 95% ( $\alpha/\beta = 1.3/1$ ), 97% ( $\alpha/\beta = 1.6/1$ ), 81% ( $\alpha/\beta = 2/1$ ) or 80% ( $\alpha/\beta = 1.5/1$ ) yield, respectively. In addition, the title compound was obtained by Method B from **1** and **13** in 91% yield ( $\alpha/\beta = 1.4/1$ ), by Method C - from **1** and **13** in 95% yield ( $\alpha/\beta = 2/1$ ), by Method D - from **6** or **7** and **13** in 91% ( $\alpha/\beta = 1.2/1$ ) or 89% ( $\alpha/\beta = 1/1$ ), and by Method E - from **6** or **7** and **13** in 92% ( $\alpha/\beta = 1.2/1$ ) or 76% ( $\alpha/\beta = 1.2/1$ ), respectively. Analytical data for **17a** were essentially the same as reported previously.<sup>14</sup>

**Methyl 2-O-(2,3,4,6-tetra-O-benzoyl- $\beta$ -D-glucopyranosyl)-3,4,6-tri-O-benzyl- $\alpha$ -D-glucopyranoside (17b).** The title compound was obtained by Method A from **2b** and **14**<sup>15</sup> in 99% yield. Analytical data for **17b** were essentially the same as reported previously.<sup>16</sup>

**Methyl 6-O-(3,4,6-tri-O-acetyl-2-O-benzyl-D-glucopyranosyl)-2,3,4-tri-O-benzyl- $\alpha$ -D-glucopyranoside (17c).** The title compound was obtained by Method A from **3c** or **5c** and **13** in 91% ( $\alpha/\beta = 1.6/1$ ) or 75% yield ( $\alpha/\beta = 1.2/1$ ), respectively. Analytical data for **17c** were essentially the same as reported previously.<sup>17</sup>

**Methyl 2-O-(2,3,4,6-tetra-O-benzyl-D-glucopyranosyl)-3,4,6-tri-O-benzyl- $\alpha$ -D-glucopyranoside (18a).** The title compound was obtained by Method A from **5a** and **14** in



92% yield ( $\alpha/\beta = 1.4/1$ ). Analytical data for **18a** were essentially the same as reported previously.<sup>17</sup>

**Methyl 2-O-(2,3,4,6-tetra-O-benzoyl- $\beta$ -D-glucopyranosyl)-3,4,6-tri-O-benzyl- $\alpha$ -D-glucopyranoside (18b).** This compound was obtained by Method A from **2b** and **14** in 82% yield. Analytical data for **19a** were essentially the same as reported previously.<sup>17</sup>

**Methyl 2-O-benzyl-3-O-(2,3,4,6-tetra-O-benzyl-D-glucopyranosyl)-4,6-O-benzylidene- $\alpha$ -D-glucopyranoside (19a).** This compound was obtained by Method A from **4a** or **9** and **15**<sup>18</sup> in 87% ( $\alpha/\beta = 7.4/1$ ) or 81% yield ( $\alpha/\beta = 2.3/1$ ), respectively. Analytical data for **19a** were essentially the same as reported previously.<sup>19</sup>

**Pent-4-en-1-yl 3-O-(2,3,4,6-tetra-O-benzyl-D-glucopyranosyl)-4,6-O-benzylidene-2-deoxy-2-(2,2,2-trichloroethoxycarbamoyl)- $\beta$ -D-galactopyranoside (21).** This compound was obtained by Method A from **3a**, **4a**, **5a**, or **9** and **20**<sup>20</sup> in 86% ( $\alpha/\beta = 1.1/1$ ), 85% ( $\alpha/\beta = 3.3/1$ ), 83% ( $\alpha/\beta = 1/1.2$ ), or 65% yield ( $\alpha/\beta = 1.6/1$ ), respectively. Analytical data for **21** were essentially the same as reported previously.<sup>17</sup>

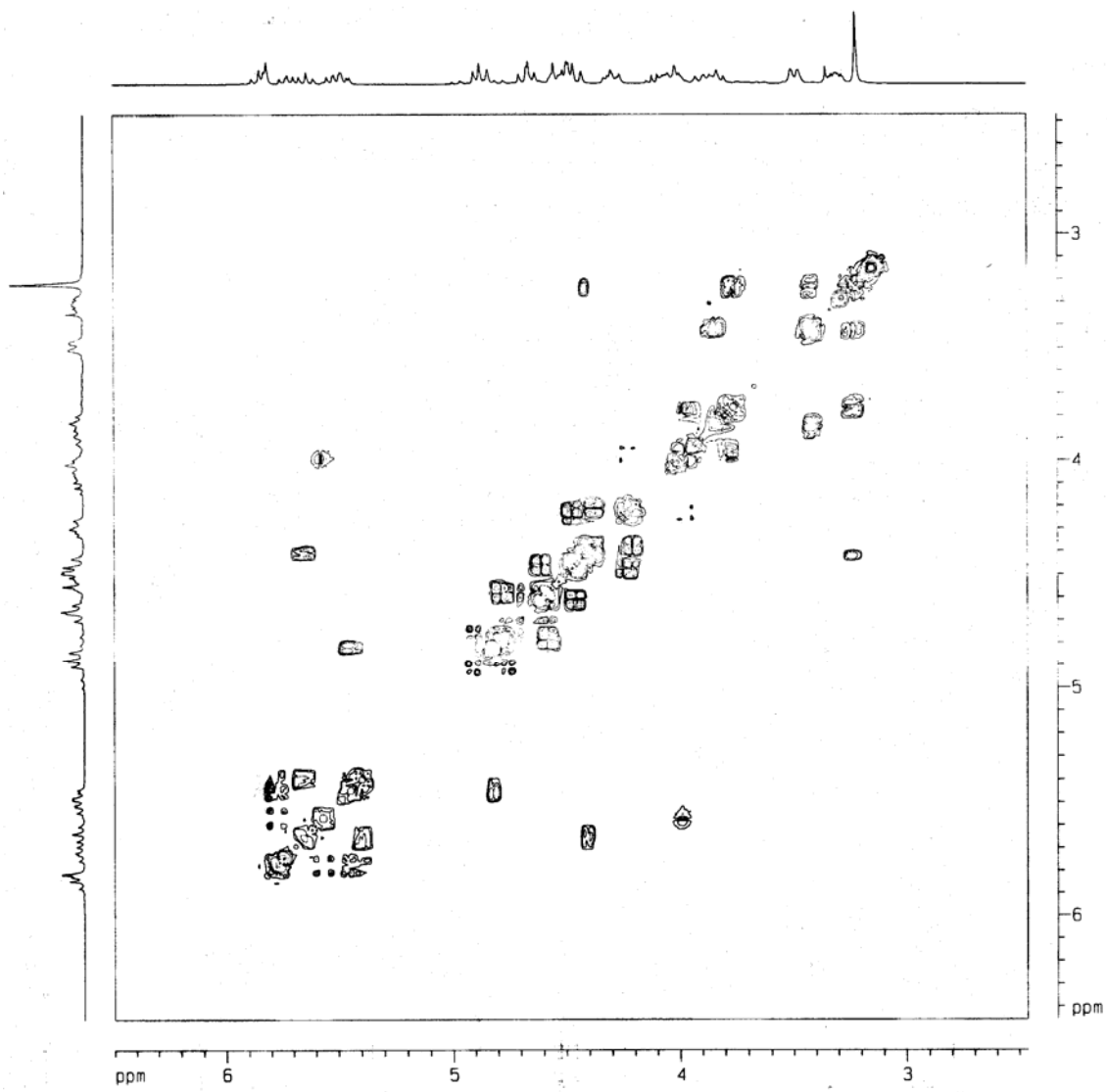
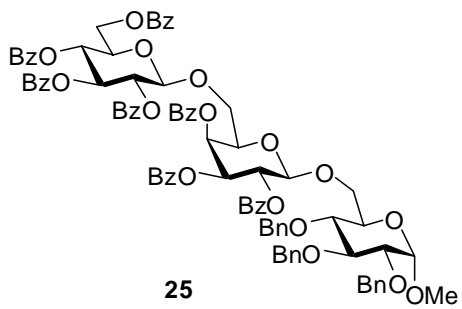
**Ethyl 6-O-(2,3,4,6-tetra-O-benzyl-D-glycopyranosyl)-2,3,4-tri-O-benzyl-1-thio- $\beta$ -D-glucopyranoside (23).** This compound was obtained by Method A from **2a**, **3a**, **5a**, **8** or **9** and **22**<sup>21</sup> in 84% ( $\alpha/\beta = 1.6/1$ ), 87% ( $\alpha/\beta = 2/1$ ), 89% ( $\alpha/\beta = 2.4/1$ ), 84% ( $\alpha/\beta = 2.2/1$ ) or 83% yield ( $\alpha/\beta = 2.4/1$ ), respectively. Analytical data for **23** were essentially the same as reported previously.<sup>22</sup>

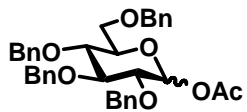
## References

- (1) Schmidt, R. R.; Michel, J. *J. Org. Chem.* **1981**, *46*, 4787-4788.
- (2) Kamat, M. N.; Rath, N. P.; Demchenko, A. V. *J. Org. Chem.* **2007**, *72*, 6938-6946.
- (3) Ramakrishnan, A.; Pornsuriyasak, P.; Demchenko, A. V. *J. Carbohydr. Chem.* **2005**, *24*, 649-663.
- (4) Bogusiak, J.; Szeja, W. *Chem. Lett.* **1988**, 1975-1976.
- (5) Andersson, F.; Fugedi, P.; Garegg, P. J.; Nashed, M. *Tetrahedron Lett.* **1986**, *27*, 3919-3922.
- (6) Ernst, B.; Winkler, T. *Tetrahedron Lett.* **1989**, *30*, 3081-3084.
- (7) Kovac, P.; Yeh, H. J. C.; Jung, G. L. *J. Carbohydr. Chem.* **1987**, *6*, 423-439.
- (8) Nguyen, H. M.; Chen, Y. N.; Duron, S. G.; Gin, D. Y. *J. Am. Chem. Soc.* **2001**, *123*, 8766-8772.
- (9) Schmidt, R., R.; Stumpp, M. *Liebigs Ann. Chem.* **1983**, 1249-1256.
- (10) Ratcliffe, A. J.; Fraser-Reid, B. *J. Chem. Soc., Perkin Trans I* **1989**, 1805-1810.
- (11) Garegg, P. J.; Hultberg, H. *Carbohydr. Res.* **1981**, *93*, C10-C11.
- (12) Pougny, J. R.; Nassr, M. A. M.; Naulet, N.; Sinay, P. *Nouveau J. Chem.* **1978**, *2*, 389-395.
- (13) Kuester, J. M.; Dyong, I. *Justus Liebigs Ann. Chem.* **1975**, 2179-2189.
- (14) Eby, R.; Schuerch, C. *Carbohydr. Res.* **1975**, *39*, 33-38.
- (15) Sollogoub, M.; Das, S. K.; Mallet, J.-M.; Sinay, P. *C. R. Acad. Sci. Ser. 2* **1999**, *2*, 441-448.
- (16) Hashimoto, S.; Honda, T.; Ikegami, S. *J. Chem. Soc., Chem. Commun.* **1989**, 685-687.
- (17) Pornsuriyasak, P.; Demchenko, A. V. *Chem. Eur. J.* **2006**, *12*, 6630-6646.
- (18) Kondo, Y. *Agric. and Biol. Chem.* **1975**, *39*, 1879-1881.
- (19) Dasgupta, F.; Garegg, P. J. *Carbohydr. Res.* **1990**, *202*, 225-238.

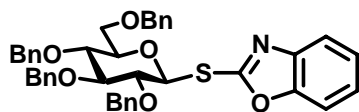
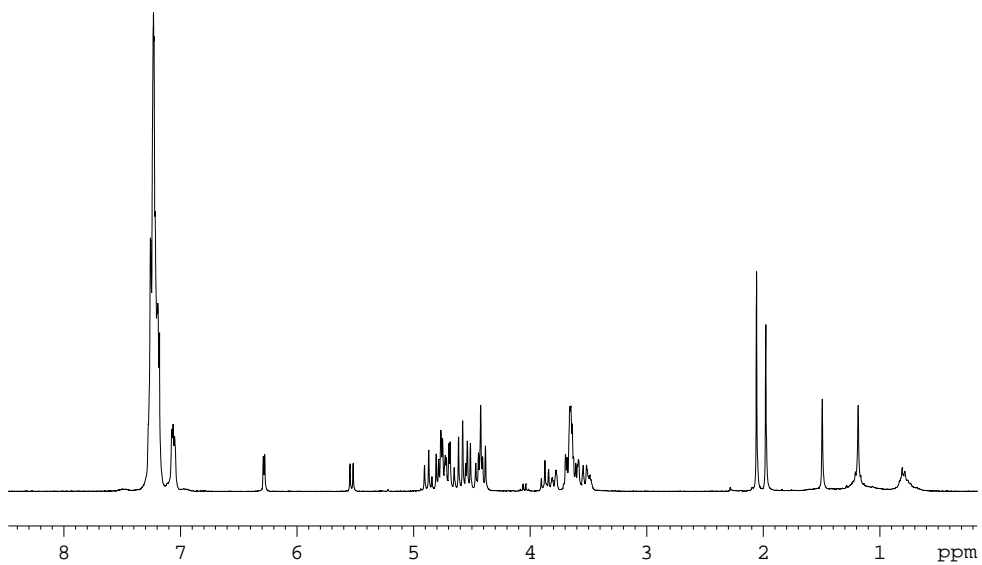
- (20) Pornsuriyasak, P.; Demchenko, A. V. *Carbohydr. Res.* **2006**, *341*, 1458-1466
- (21) Ray, A. K.; Roy, N. *Carbohydr. Res.* **1990**, *196*, 95-100.
- (22) Mehta, S.; Pinto, B. M. *J. Org. Chem.* **1993**, *58*, 3269-3276.



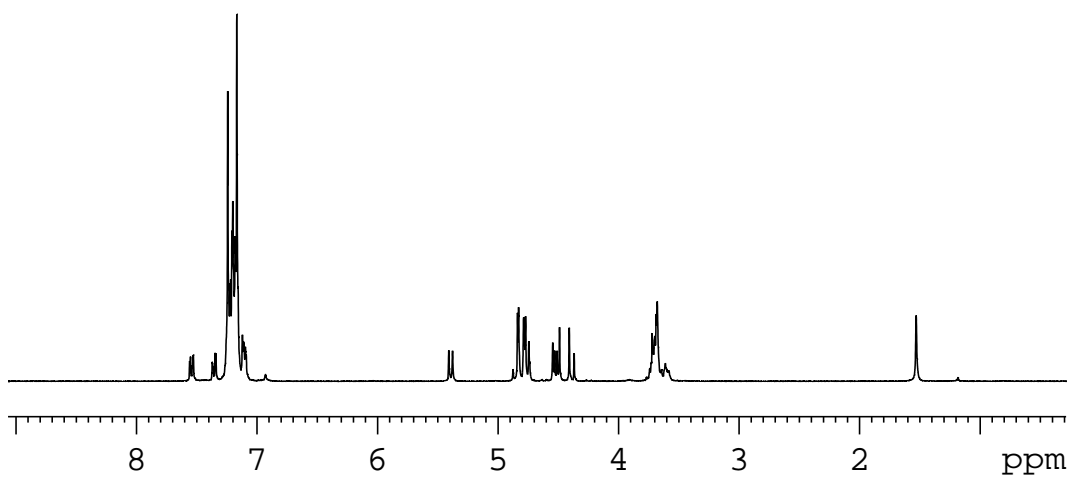


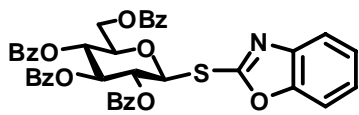


1

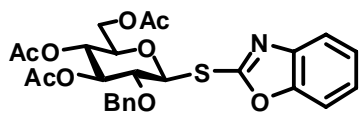
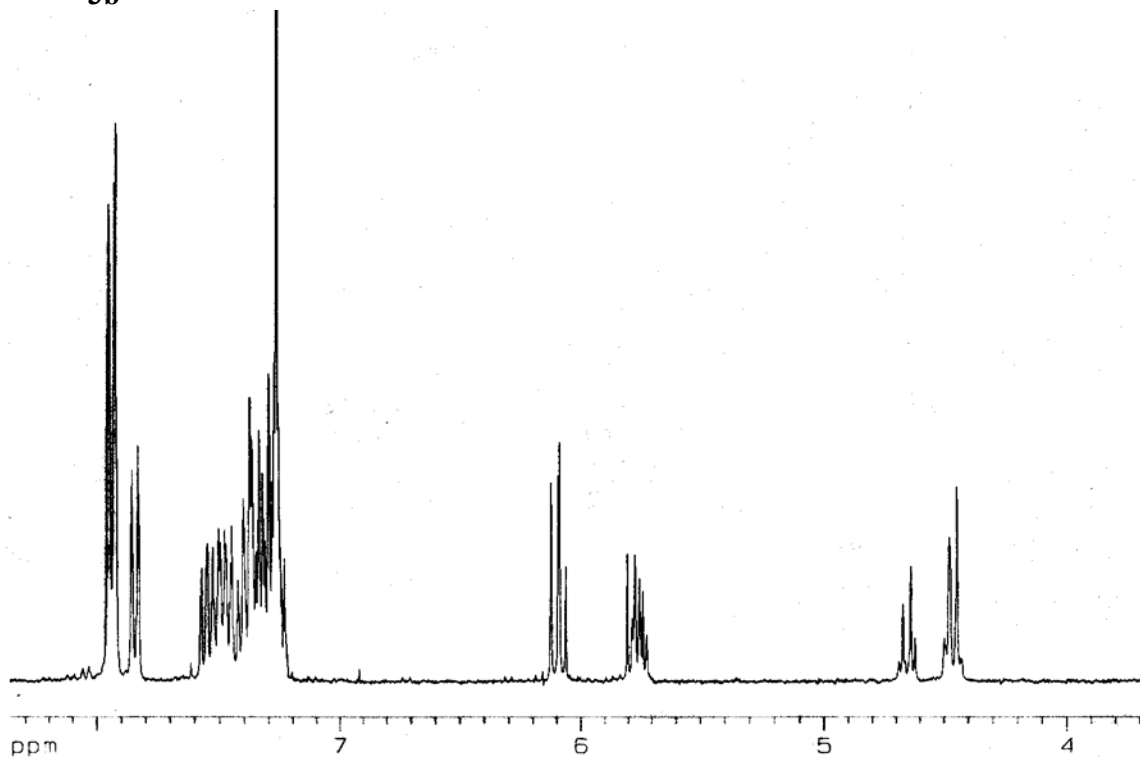


3a

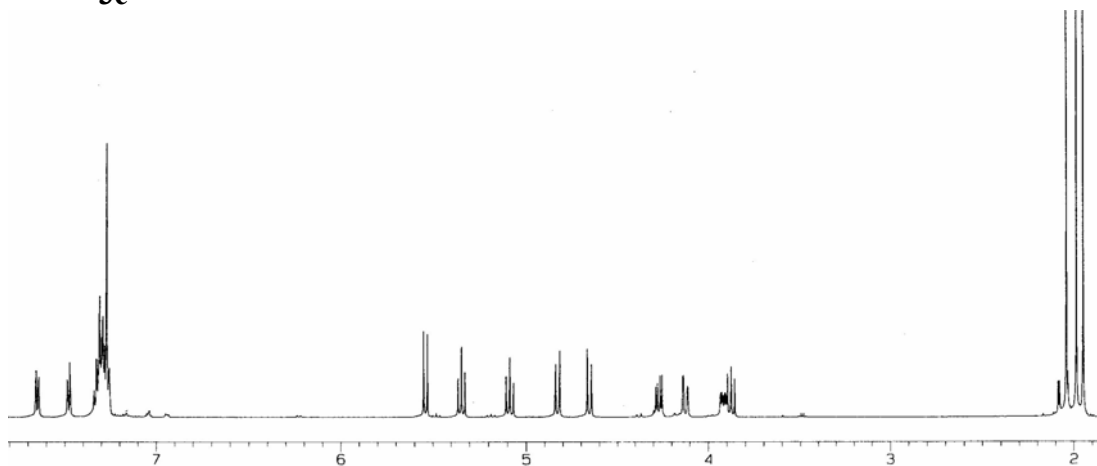


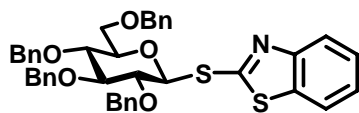


3b

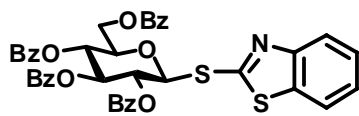
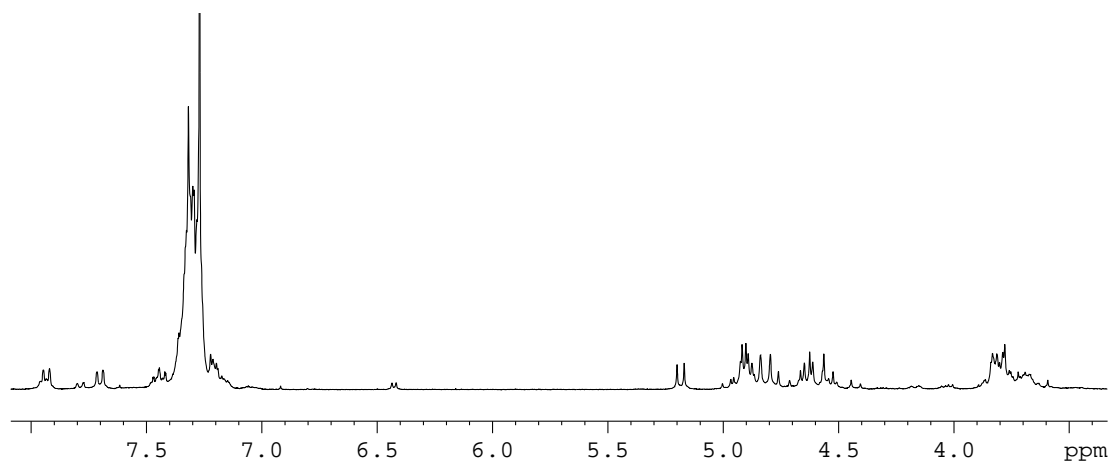


3c

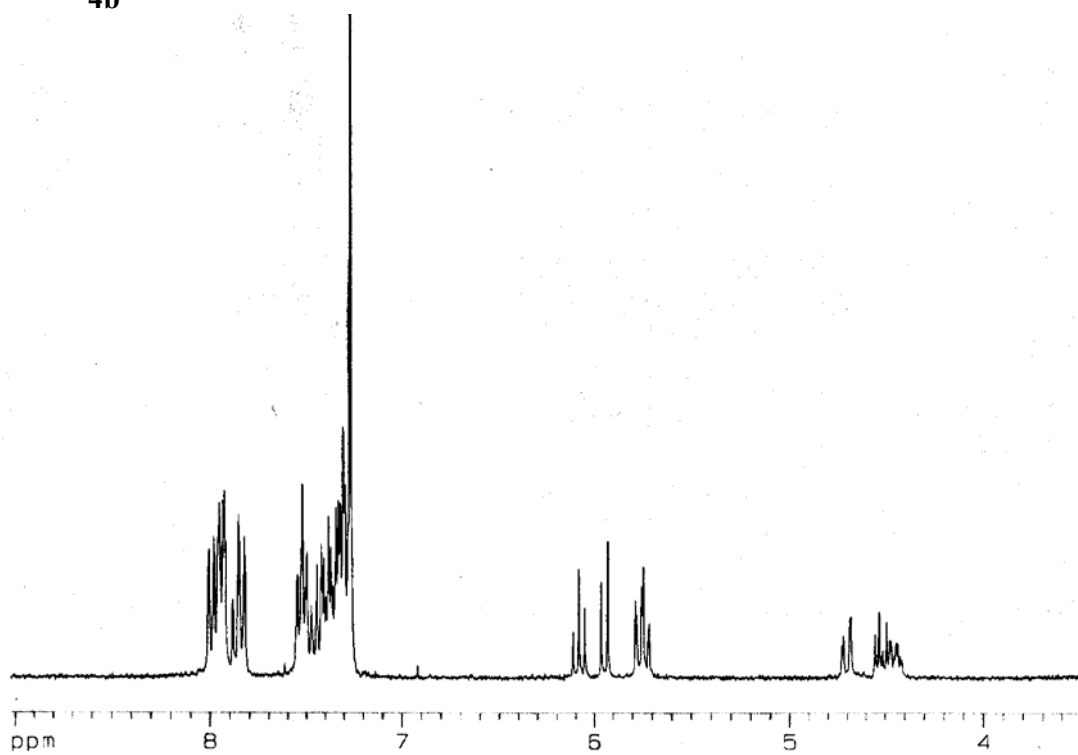




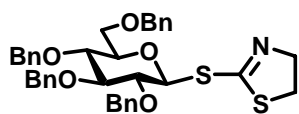
4a



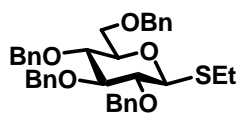
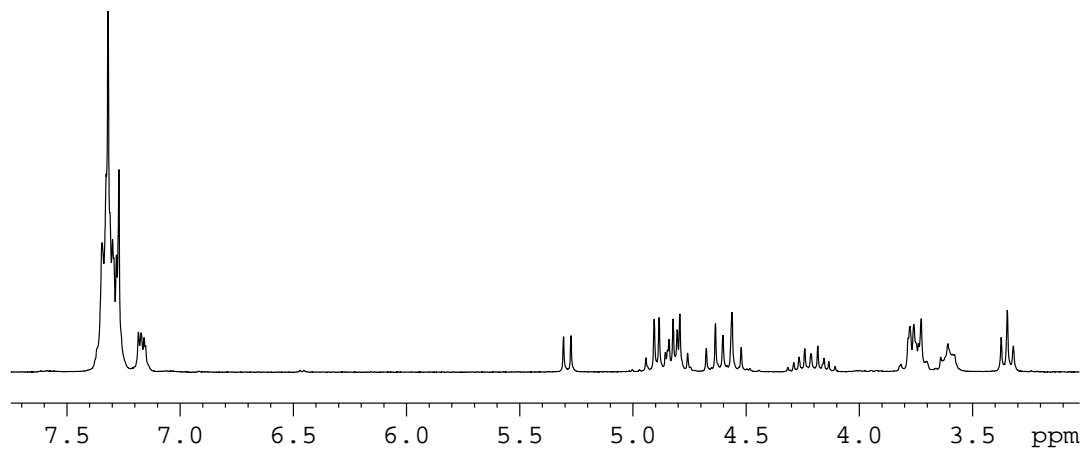
4b



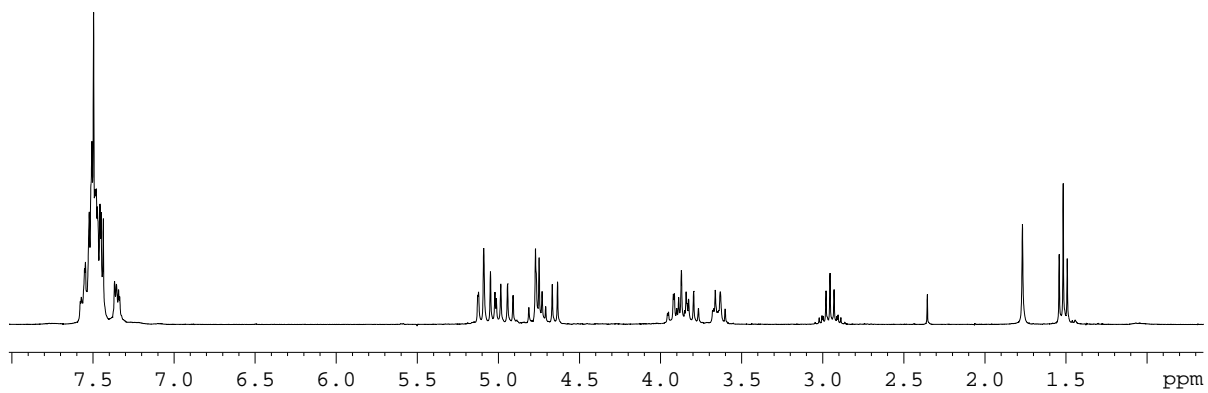


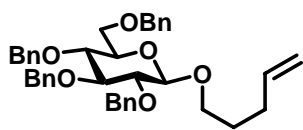


5a

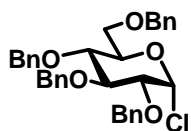
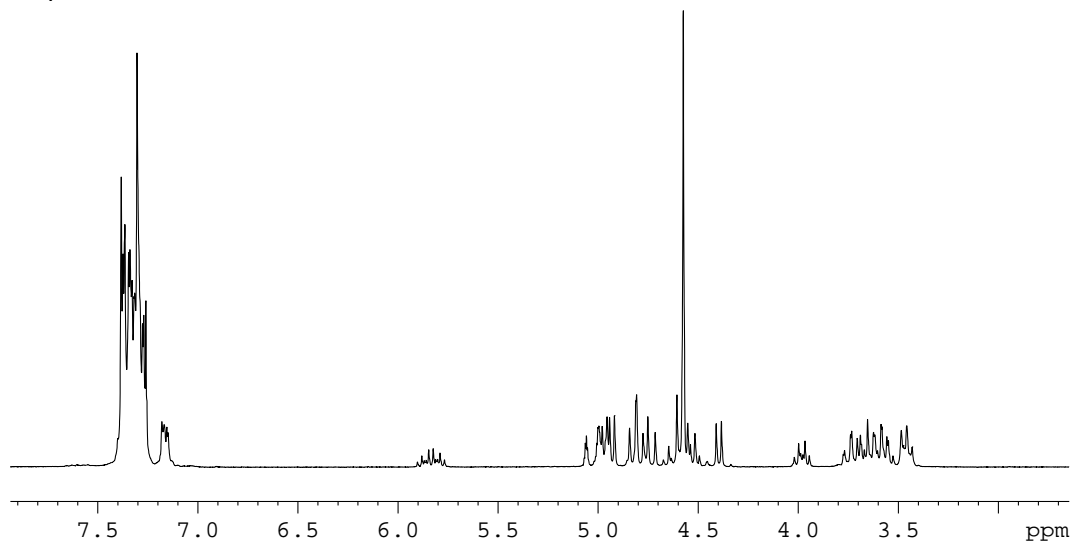


6

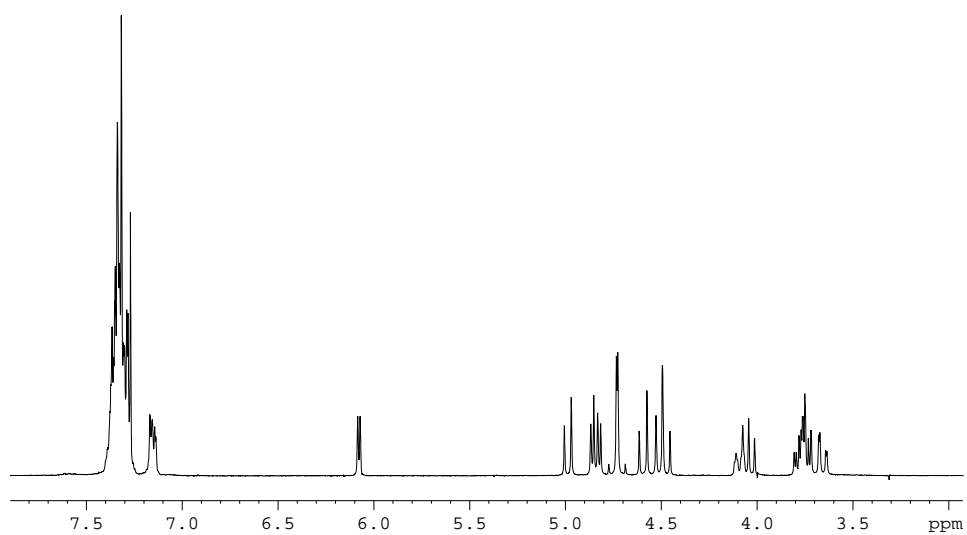


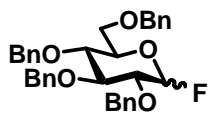


7

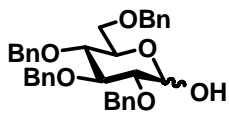
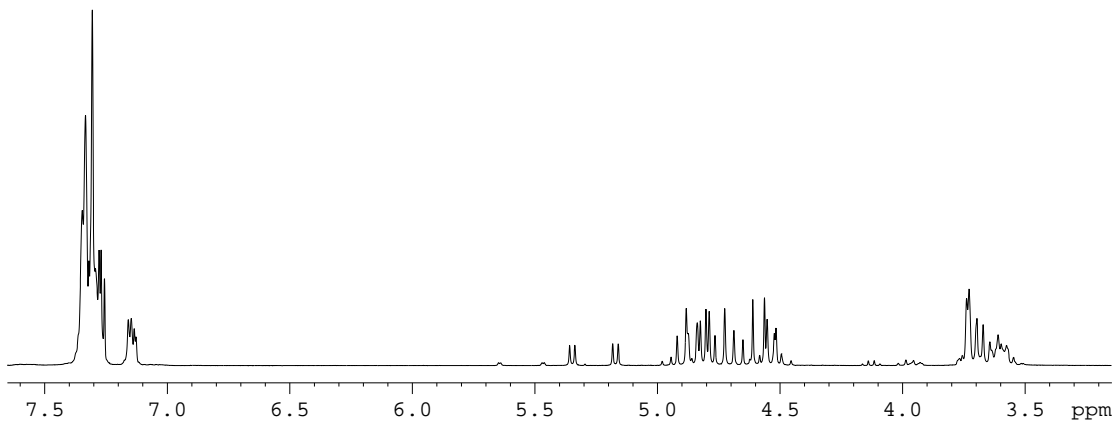


8

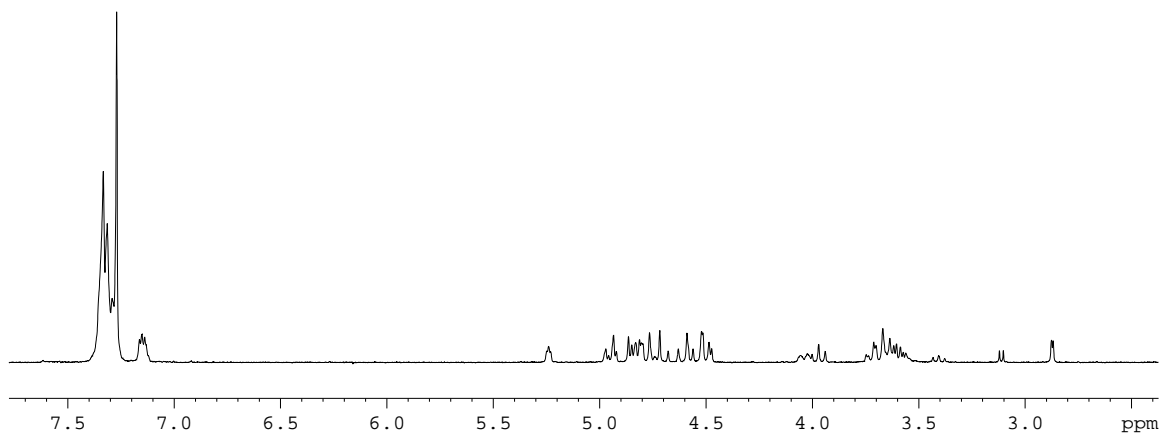


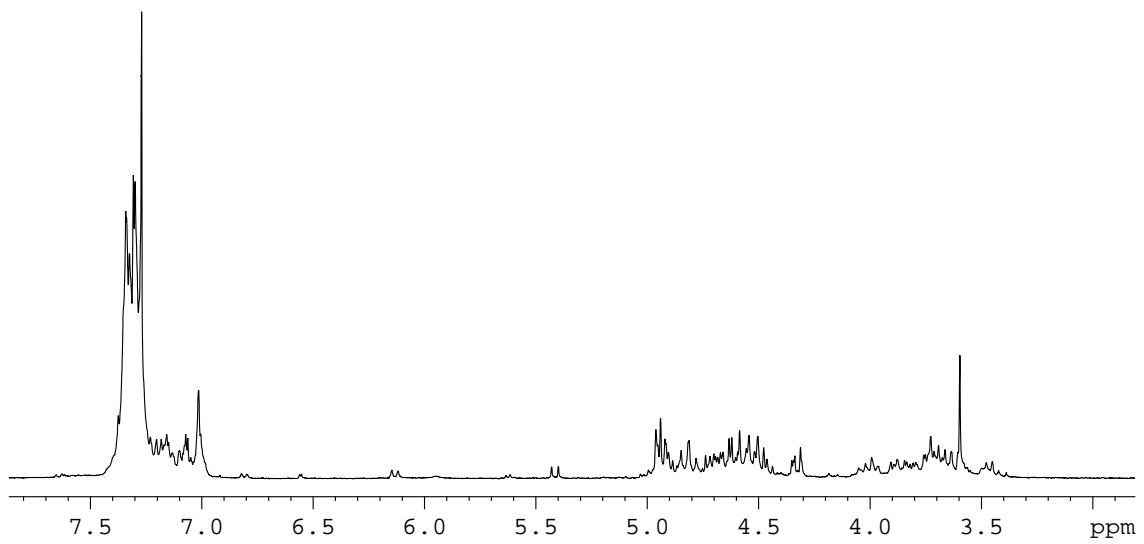
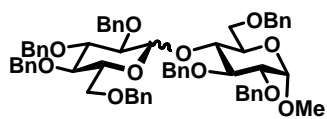
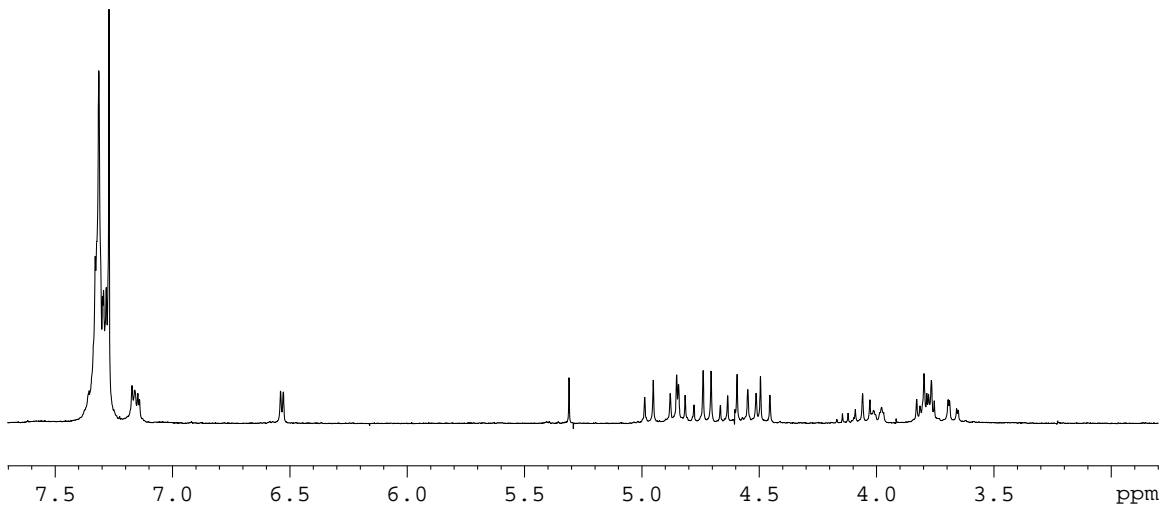
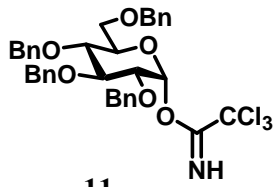


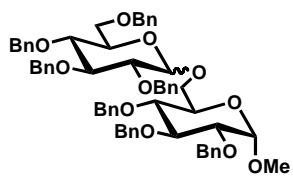
9



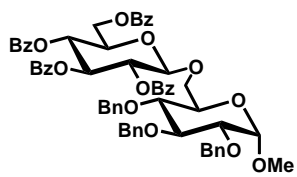
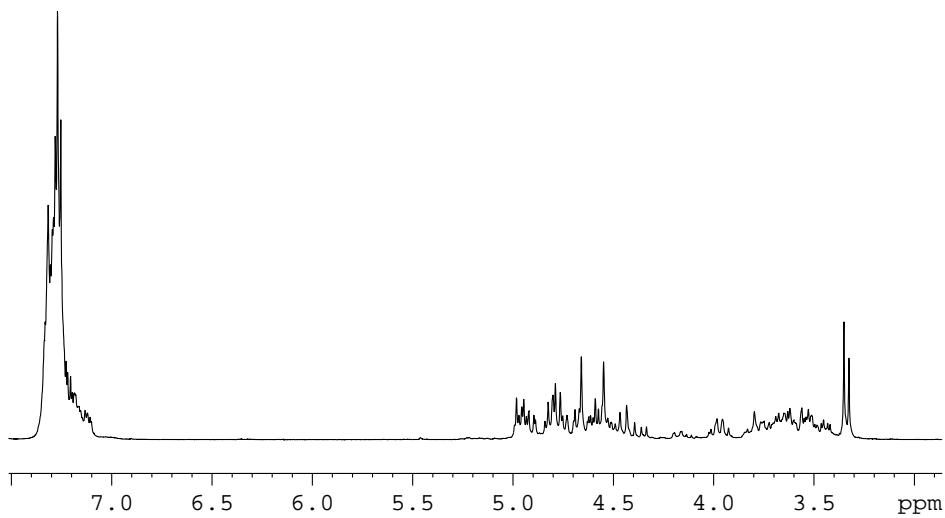
10



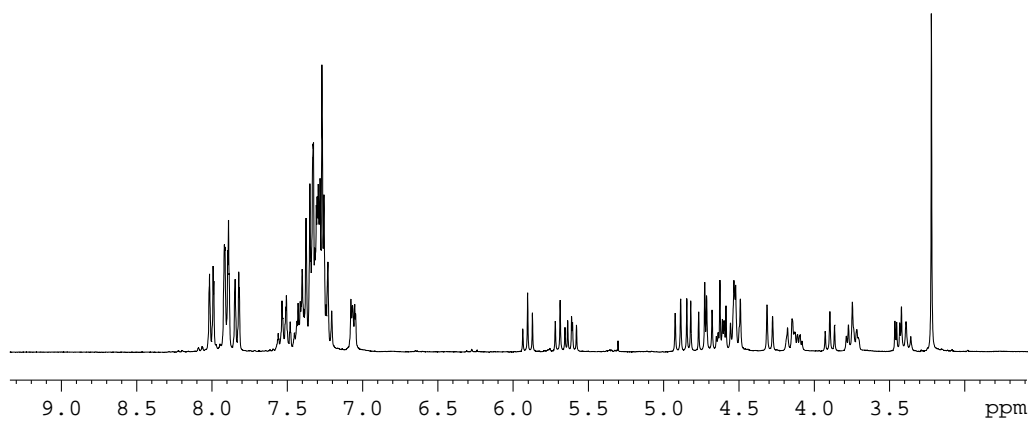


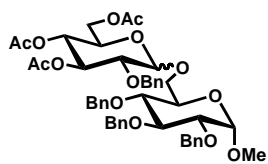


17a

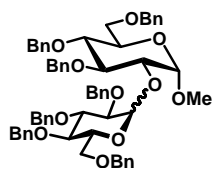
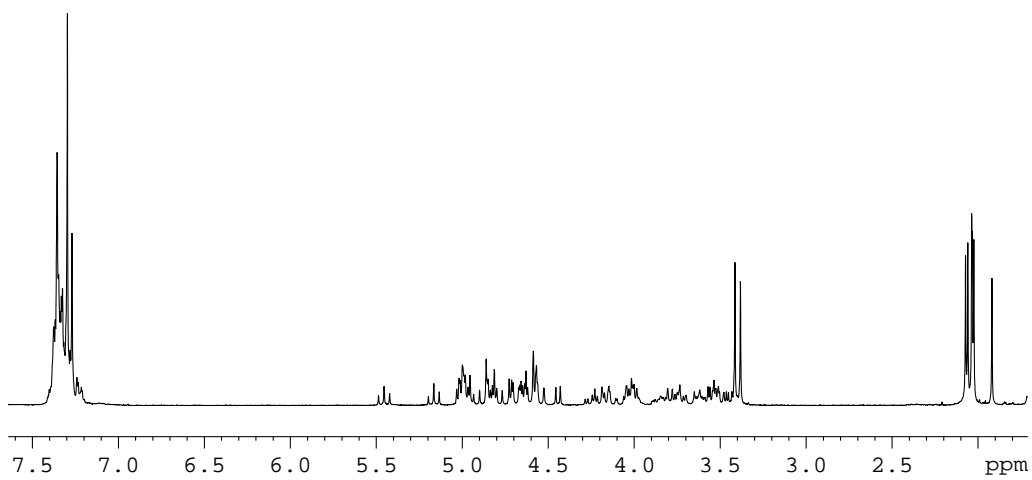


17b

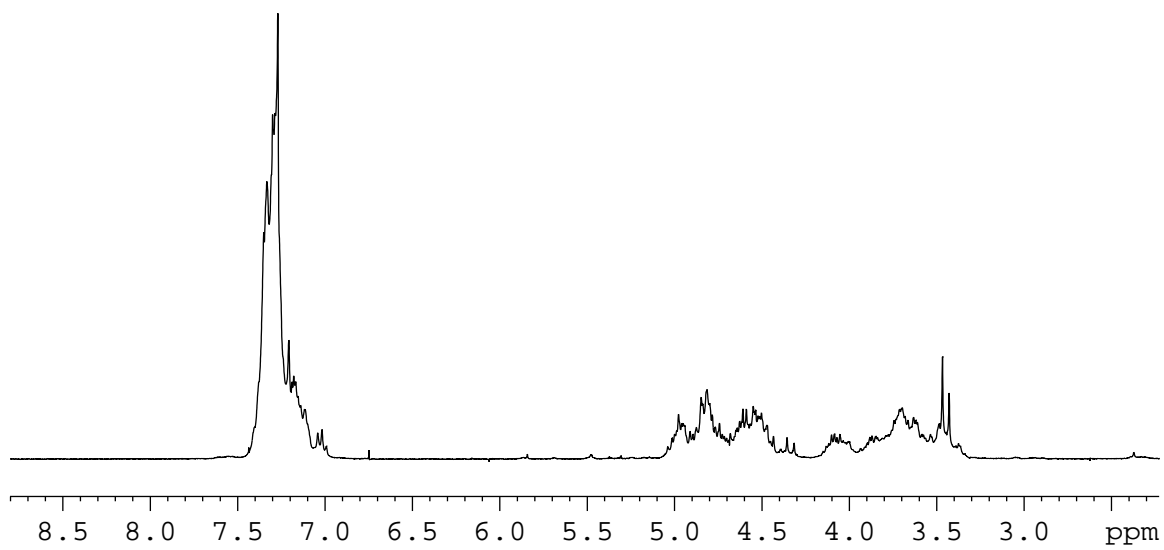




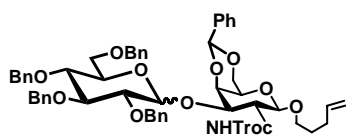
17c



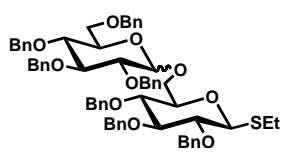
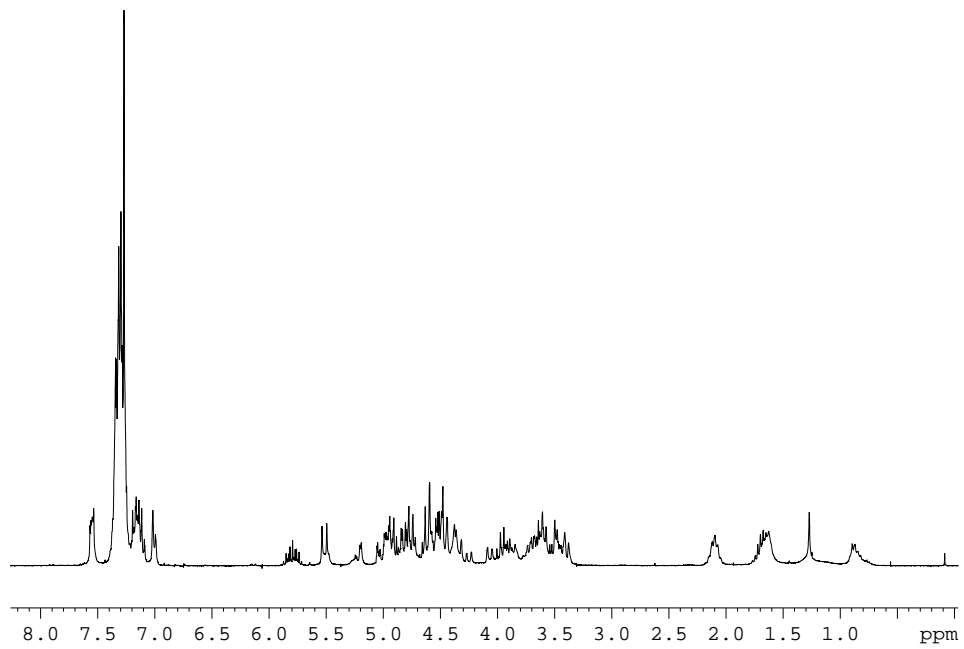
18a







21



23

