T Helper Cell Activation and Human Retroviral Pathogenesis

KAREN F. T. COPELAND^{1*} AND JONATHAN L. HEENEY^{2*}

Molecular Virology and Immunology Programme, Department of Pathology, Health Sciences Centre, McMaster University, Hamilton, Ontario, Canada, and Laboratory of Viral Pathogenesis, Department of Virology, Biomedical Primate Research Centre, Rijswijk, The Netherlands²

INTRODUCTION	
T HELPER CELL BIOLOGY	723
T Helper Cell Development and Maturation	723
Activation of T Helper Cells	723
CONSEQUENCES OF RETROVIRAL INFECTION ON IMMUNE ACTIVATION	
HIV Infection	725
HTLV Infection	727
EFFECT OF T HELPER CELL ACTIVATION ON RETROVIRAL EXPRESSION	
T HELPER CELL ACTIVATION AND DISEASE PROGRESSION	729
HIV and AIDS	729
HTLV and ATL	730
EFFECT OF HUMAN RETROVIRUS-ENCODED PROTEINS ON APOPTOSIS	
APOPTOSIS AND DISEASE PROGRESSION	732
CONCLUSIONS	734
ACKNOWLEDGMENTS	734
REFERENCES	734

INTRODUCTION

The understanding of disease caused by human retroviral infections has been hampered by our lack of understanding of the complexity of the interactions of these viruses with the human immune system. In fact, it is in part through the study of these infections that important insights into the workings of the immune system itself have been made. Human retroviruses have the common characteristics of causing chronic persistent infection and a long and variable asymptomatic period during which overt clinical disease is not frequently manifested. Substantial evidence exists documenting that the human retroviruses, human immunodeficiency virus (HIV) and human T-cell leukemia virus (HTLV), take advantage of activated T helper (Th) cells to initiate permanent infection (117, 312, 364). Furthermore, the transcriptional signals used by Th cells to regulate cell function are also used by these retroviruses to regulate virus production. In effect, after establishing infection, human retroviruses appear to benefit from an active immune system to subsequently proliferate. This is, however, not without consequences. The way in which these viruses respond to the intracellular signals produced in Th cells following contact with common antigens eventually leads to a distortion in Th-cell function, and depending on which virus is involved, this may lead to two extremely different disease outcomes. Infection with HTLV results in dysregulated Th-cell proliferation, sometimes causing a disease of excess Th cells commonly known as adult T-cell leukemia (ATL). In contrast, infection with HIV causes a disease of profound Th-cell loss, resulting in systemic immunosuppression and AIDS. Thus, in both instances, disease progression is intimately linked to a disturbance of normal Th-cell growth and function, although the disease outcome represents two different extremes in Th-cell numbers.

Activation occurs following the interaction of Th cells with specialized cells (antigen-presenting cells [APC]), which present foreign antigen. When appropriate costimulatory signals are delivered during this interaction, the Th cell becomes activated and can proceed to differentiate and proliferate. In addition to activation, two other normally occurring alternative outcomes of APC-Th-cell interaction, anergy and apoptosis, may occur. Anergy is a state of nonresponsiveness which may occur to protect the host from inappropriate Th-cell responses if the proper coregulatory signals are not available. Apoptosis is a fundamental process which assists in the regulation and normal physiologic development and balance of cell populations. In populations of immunologic cells, apoptosis is a natural process aimed at the regulated removal of unwanted or self-reactive cells. Apoptosis is also observed to be increased in a large number of disease states, often reflecting disturbances in normal physiological processes. In addition to anergic cells, highly elevated levels of apoptosis have, for instance, been observed in HIV-infected individuals, occurring in Th cells and other lymphocyte populations. In contrast, HTLV infection, which is also intimately linked to cellular activation, is associated with a protection from apoptosis, possibly culminating in the development of Th-cell cancer. In this review, the active involvement of immune activation, anergy, and apoptosis and specifically the consequences of dysregulated Th-cell functions in the pathogenesis of human retroviral infections will be discussed with particular regard to their significance to aberrant Th-cell function and disease progression.

^{*} Corresponding author. Mailing address for Jonathan L. Heeney: Laboratory of Viral Pathogenesis, Department of Virology, Biomedical Primate Research Centre, Lange Kleiweg 151, Rijswijk, The Netherlands. Phone: 33 15 284 2661. Fax: 33 15 284 3986. Electronic mail address: heeney@bprc.nl. Mailing address for Karen F. T. Copeland: Rm. 3N11, Department of Pathology, Health Sciences Centre, McMaster University, 1200 Main St. West, Hamilton, Ontario L8N 3Z5, Canada. Phone: (905) 525-9140 Ext. 22494. Fax: 905-521-2613. Electronic mail address: copelndk@fhs.mcmaster.ca.

T HELPER CELL BIOLOGY

T Helper Cell Development and Maturation

Many activities of Th cells are to a large degree mediated by the production of cytokines. A number of events determine which type of cytokines these cells will produce, depending on the type of Th cell they become (Th0, Th1, Th2, or Th3). The events which determine this are largely influenced by the nature of the infectious agent, the type of APC involved and the local microenvironment. Depending on the costimulating signals that the Th cell receives, it may undergo appropriate activation, proliferation, and cellular differentiation into an antigen-specific Th1/0 or Th2 population. Current research in this area is revealing other subpopulations such as Th3 cells. However, for the purposes of simplifying this discussion, we will confine ourselves to Th1 and Th2 cells. If the appropriate costimulatory signals are not provided, the Th cell may enter a nonresponsive state (anergy) or undergo programmed cell death (apoptosis) (reviewed in reference 306). Activation, anergy, and apoptosis are all normal physiologic processes which regulate T-cell responses. Stimulation of Th cells with antigen presented by the class II major histocompatibility complex (MHC) APC, followed by appropriately polarized costimulatory signals and additional cytokines, commonly results in a specific proliferation and further development into a dichotomy of the two extremes referred to as Th1 or Th2 (143). The cytokines (type 1 and type 2) produced by Th1 and Th2 cells, respectively, serve to regulate the function of specific immune system effector responses of cytotoxic T lymphocytes (CTL) and antibodies by B cells, respectively, in response to foreign antigen. The Th phenotype produced is influenced by the cytokines in the microenvironment of the Th cell, where it interacts with particular APC, influenced by surrounding cell types and their cytokines at the time of antigen presentation (236, 348).

Differentiation of naive Th cells into the Th1 phenotype is promoted by gamma interferon (IFN-γ). Th1 cells are characterized by their ability to produce interleukin-2 (IL-2), IFN-y and tumor necrosis factors alpha and beta (TNF- α and - β), which are not made by Th2 cells. In contrast, Th2 (but not Th1) cells synthesize IL-4, IL-5, IL-6, and IL-10 (reviewed in references 274, 285, and 297). Th1 and Th2 cells also differ markedly in the signals required for their development. IL-2 is required for the development of the Th1 population, and these cells drive the cell-mediated arm of the immune response, including transient antibody production, the activation of macrophages, and delayed-type hypersensitivity responses. The Th2 phenotype is induced by IL-4, and restimulation of these cells results in high-level IL-4 secretion (298, 322). The humoral arm of the immune response is driven by Th2 cells, resulting in sustained production of antibodies, including immunoglobulin E (IL-4) and activation of eosinophils (IL-5) and mast cells (IL-3, IL-4, and IL-10) (274, 285, 297). In the absence of clear polarizing signals, a Th cell designated Th0 may develop. These cells produce a cytokine profile intermediate between those of Th1 and Th2 cells and can undergo partial differentiation to the Th1 or Th2 phenotype (1, 93, 313). A Th3 phenotype which produces high levels of transforming growth factor β (TGF- β) has also been described (44).

The development of a particular Th phenotype may be favored depending upon the exogenous stimulus received by the host. In response to infections by intracellular organisms (viruses, bacteria, and protozoa), the Th1 population is favored (69, 129, 284, 287, 363). The induction of a Th1 response by intracellular bacteria and viruses reflects their ability to stim-

ulate IFN- α/β and IL-12 production by macrophages (285). These cytokines induce IFN- γ production by natural killer (NK) cells and T cells, which in turn inhibits the development of IL-4-producing cells (65). When IFN- γ levels are absent or limited, IL-4 production favors the development of Th2 cells, which predominate in response to particulate antigens and extracellular organisms including allergens and helminth infections (69, 261, 284, 288). The conditions favoring a particular Th shift are thus dependent upon the cytokines elicited during the initial immune response to an exogenous agent, and this will affect any subsequent specific immune response.

Activation of T Helper Cells

Th cells are specialized in that they express cell surface receptors which recognize and bind foreign antigen in the form of short peptides displayed in the context of MHC class II by APC (B cells, macrophages, and follicular dendritic cells). Activation of the Th cell in the proper context results in the expansion of Th populations and the elimination of inappropriately responding Th cells by apoptosis. The new population, in the appropriate environment, provides the necessary cytokines for the proliferation and expansion of other populations of effector cells, such as B cells or CD8⁺ CTL, enabling them to utilize their effector mechanisms to remove foreign antigen. Full activation requires T-cell receptor (TCR)-CD3 complex recognition of peptide antigens presented by APC, as well as nonantigenic costimulatory signals provided by the APC. Costimulation is provided by the engagement of the B7 molecule on APC with the constitutively expressed CD28 receptor on the Th cell (110, 132, 159). Activation profoundly induces the expression of cytolytic T-lymphocyte-associated sequence (CTLA-4) (29), a receptor which shares 28% amino acid identity with CD28 (133) and which binds B7 with a 20-fold greater avidity than CD28 does (207). B lymphocytes express three distinct CTLA-4 counterreceptors designated B7-1, B7-2, and B7-3 (25). While the interaction of B7 molecules at the CTLA-4 receptor can enhance proliferative responses of resting Th cells costimulated with anti-CD3 and anti-CD28 antibodies (208), anti-CTLA-4 antibodies inhibited proliferation when cross-linked or presented in an immobilized form together with anti-CD3 and anti-CD28 antibodies (9, 25, 135, 182, 202, 207). This inhibition results from a block in the transition from the G_1 to the S phase of the cell cycle (185). Furthermore, anti-CTLA-4 antibodies can induce cell death of activated T cells (120). The contribution of CTLA-4 to Th cell proliferation is thus more complex than was originally reported and is an area requiring further study. A novel receptor (SLAM) involved in T-cell activation has been reported to enhance the proliferation of and cytokine production by CD4⁺ T cells in the absence of any other stimuli (57). Signalling via SLAM potentiates the development of a Th0/Th1 cytokine profile in a CD28-independent manner (57).

The outcomes of APC–Th-cell interaction and the development of the Th1 and Th2 phenotypes are presented in Fig. 1. Appropriate costimulation results in a cascade of intracellular signalling pathways required for the induction of IL-2 and Th-cell proliferation. In one of these pathways, the activity of phospholipase C-γ1 is enhanced by phosphorylation, allowing the cleavage of membrane-bound phosphatidylinositol 4,5-bisphosphate to generate the second messengers inositol triphosphate and diacylglycerol (55). The production of inositol triphosphate triggers a dramatic bimodal elevation of free calcium levels within the cell (55, 102, 105). Diacylglycerol directly activates protein kinase C (PKC) (61). Both second-messenger signals are essential for the production of IL-2,

T-helper cells in health

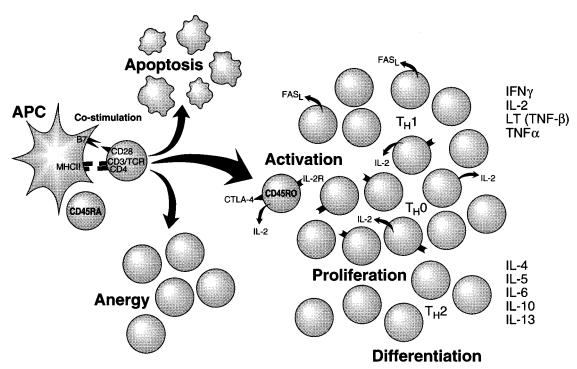


FIG. 1. Activation, proliferation, and differentiation to Th1 and Th2 phenotypes. The events following costimulation of a Th cell by interaction with an APC include activation, anergy, and apoptosis. Following appropriate costimulation, the Th cell proliferates and differentiates into Th subpopulations dependent upon the nature of the stimulatory signal and cytokines in the microenvironment.

which in turn is required for the proliferation of the activated cell and for the maintenance of memory. The availability of IL-2 allows the Th cell to remain in the cell cycle and also protects it against apoptosis. Repeated activation of T cells may result in a loss of IL-2 synthesis and the subsequent elimination of these cells by apoptosis.

While Th1 cells proliferate in response to IL-2, Th2 cells proliferate in response to IL-2 or IL-4, provided that IL-1 is made available by the APC (348). In contrast, Th1 cells express little or no IL-1 receptor (39). Mitogen stimulation results in much higher levels of cyclic AMP (cAMP) in Th2 cells than in Th1 cells (238). Inducers of cAMP inhibit Th1 cell proliferation but do not inhibit Th2-cell proliferation (131). Furthermore, agents which elevate cAMP levels block IL-2 and IFN-γ production by Th1 cells but not IL-4 production by Th2 cells (20). The level of cAMP generated in the Th cell may differentially regulate the lymphokines produced by Th cells, and the higher levels of cAMP induced in the activated Th2 cell may interfere with several transcription factors including NF-κB and AP-1. This could explain the contrasting cytokine profiles of Th1 and Th2 cells (8, 43, 247, 332). Lower NF-kB levels are noted in Th2 cells (198), and the cytokines produced by both Th1 and Th2 cells significantly modulate levels of the NF-κB inhibitor protein, IκBα (197). Furthermore, the relative abundance of the IL-4-inducible transcription factor STAT6 correlates with differentiation to the Th2 phenotype (199). The influence of cAMP levels on STAT production in Th2 cells has not been reported. Another Th2 lymphokine, IL-10, inhibits antigen-stimulated proliferation of Th1 cells (91, 92), probably by modulating the synthesis of B7 molecules (73). It seems

likely that Th2 cells do not require costimulation via CD28/CTLA-4 for activation. Thus, the immune response generated to an external stimulus not only is limited by the lymphokines produced by the Th cell and their downstream effects on Th-cell activation genes but also may be influenced by the availability of accessory signals required during activation. Th-cell activation may result in the coupling of the Fas receptor (CD95) with its ligand. CD95 is a member of the TNF/nerve growth factor receptor superfamily, and ligation of CD95 with Fas ligand (FasL) causes rapid apoptosis (programmed cell death) in sensitive cells (315). Apoptosis is characterized by the activation of a Ca²⁺-dependent endonuclease which cleaves chromosomal DNA between nucleosomes (301, 354). The preferential expression of FasL on Th1 cells (279) arms these cells to kill other cell types or other Th1 cells by apoptosis.

CONSEQUENCES OF RETROVIRAL INFECTION ON IMMUNE ACTIVATION

Human retroviruses cause persistent infections characterized by long, clinically asymptomatic periods prior to disease progression. HTLV is known primarily as the etiologic agent of ATL, a malignancy of mature Th lymphocytes (140, 141, 268, 361). A hallmark of HTLV infection is the induction and continuous high-level expression of IL-2 receptor (IL-2R) (Table 1) (71, 114, 149, 272, 317, 357). Infection with HIV, the etiologic agent of AIDS (14), is marked by a loss of CD4⁺ Th cells, resulting in immune system dysfunction (Table 1). Both infections induce a general state of immune system activation and are associated with neurological disorders. As with other

TABLE 1. Human Th-cell tropic retroviruses and Th-cell diseases

Characteristic	Mechanism in:			
Characteristic	Th-cell cancer	AIDS		
Underlying lesion	Accumulation of Th cells	Loss of Th cells		
Virus	HTLV-1	HIV-1		
Characteristics in host	Persistent intracellular viremia; low viral load	Persistent intracellular and extracellular viremia; high virus load		
Frequency of disease	Infrequent; approximately <1% infected develop ATL	Frequent; approximately >98% infected develop AIDS		
Activation	Increase in IL-2R and IL-2 spontaneous proliferation; non-MHC-restricted responsiveness	Antigen-antibody complexes in germinal centers; infection of APC; aberrant cytokine production by APC and altered cytokine production by Th cells		
Apoptosis	Resistance due to Tax	Increase in frequency and in susceptibility to Tat		
CD3/TCR triggering	Decrease in HTLV-1 expression	Increase in HIV expression		

viruses, these retroviruses have the potential to subvert the cellular transcriptional machinery with specific regard to utilizing Th-cell signals for their own replication. This subversion of the host cell machinery most often results in a dysregulation of normal Th-cell function and growth. While both viral infections result in a generalized increase in Th-cell activation, there is a profound divergence both in the subsequent Th-cell activation-related events and in the ensuing consequences of cell expansion versus cell loss. The alterations in Th-cell populations as a result of HIV-1 and HTLV-1 infection and the resulting dysregulation of normal cellular activation and proliferation are presented schematically in Fig. 2.

HIV Infection

HIV infection can be generally grouped into three clinical phases: (i) the acute infection period, which in some ways presents similarly to a mononucleosis-like syndrome; (ii) an asymptomatic period of variable duration; and (iii) the period of clinical disease during which multiple opportunistic infections and/or neoplasms are manifested. Recent studies have shown that HIV infection in vivo is a dynamic process involving continuous rounds of infection, replication, and cell death (144, 349). HIV predominantly infects CD4⁺ T cells (Th cells) as well as CD4+ cells of the monocytic lineage, which also express MHC class II. The turnover of CD4⁺ T cells during HIV infection is thought to be rapid, with the entire population of peripheral $CD4^+$ \check{T} cells estimated to be replaced on average every 15 days (144). While the greater majority of infected individuals do not present clinical signs of disease for extended periods, virus replication remains high, especially in reservoirs such as the lymph nodes and spleen (74). In patients who die of AIDS-related illnesses, infection is extensive, targeting brain, lung, colon, and liver cells (74). Persistent and high levels of virus replication result in the loss and destruction of normal lymphoid architecture. As the immune system chronically deteriorates, virus and virus-infected cells are less efficiently removed by the host.

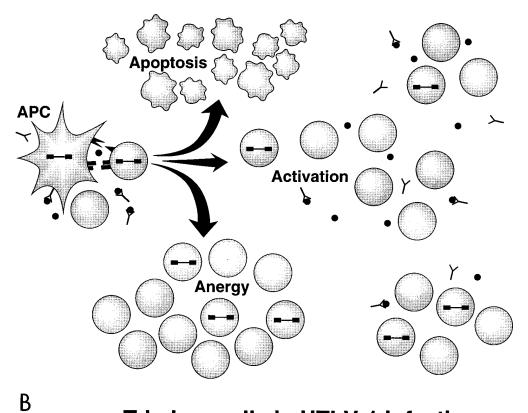
The spread of HIV infection to brain cells may result in an AIDS dementia complex, represented by a variety of mild to severe neurological disorders, including neoplasms, peripheral neuropathies, and myopathies (reviewed in reference 310). Neurotoxins produced by infected cells and induced by viral proteins may contribute to cellular damage, resulting in neu-

rologic disorders. Neurotoxic activity has also been demonstrated for the viral regulatory proteins Tat and Nef in a variety of cell lines and animal models (reviewed in reference 204). Virus replication can be measured in brain tissue (180) and the central nervous system (340) in patients with AIDS dementia disorders. However, there is no evidence to suggest that the development of AIDS dementia is associated with increased viral load. In fact, one study of children with severe AIDS-related encephalopathy indicated low viral replication in brain tissue (340). However, vigorous CTL activity directed toward HIV-1 Env, Gag, Pol, and Nef proteins has been reported to occur in the cerebrospinal fluid of patients with AIDS dementia (157).

Persistence of virus in the host in the presence of ineffective immune system clearance results in a state of chronic immune system activation. Activation of cells in the course of the immune response further favors the spread and establishment of HIV in new target CD4+ Th cells and in macrophages. In addition, virus replication is potentiated both by activation signals and by cytokines such as IL-6 and TNF- α (270, 288). Cell-mediated and humoral immune responses are detectable early in infection. These include virus-specific neutralizing antibodies, antibody-dependent cellular cytotoxicity, CTL and NK cells, and complement-dependent lysis (reviewed in reference 257). In addition, within the CD8⁺ population is a subset of cells which induce an activation-dependent, nonlytic suppression of virus replication (347). Studies of long-term nonprogressors infected with HIV-1 suggest that low viral load is associated with apparently effective neutralizing antibody and CD8⁺ lymphocyte responses (31, 258).

The asymptomatic period associated with HIV-1 infection is characterized by a high-level state of immune system activation, as is evident by the expression of related activation markers (Table 2). These include β_2 -microglobulin, serum and urinary neopterin, soluble IL-2R, soluble CD8 molecules, and soluble TNF- α receptors. The increase in the levels of many of these markers during disease progression is paralleled by the development of immune system dysfunction, which may result in an anergy-like loss of proliferative potential (191, 263, 299, 300) or in cell death by spontaneous or activation-induced apoptosis (4, 12, 115, 122, 227). A deficiency in extracellular cysteines and intracellular glutathione occurs early in HIV infection and has been associated with a decline in CD4⁺ Th cells and with progression to disease (77, 79). While cysteine

A T-helper cells in HIV infection



T-helper cells in HTLV-1 infection

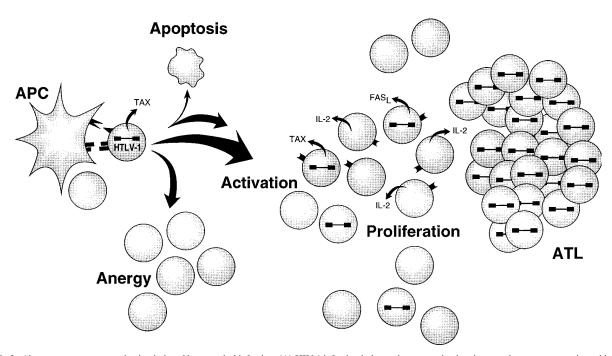


FIG. 2. Aberrant responses to activation induced by retroviral infection. (A) HIV-1 infection induces aberrant activation, increased anergy, apoptosis, and impaired proliferation and loss of Th cells by immune system destruction. (B) Infection with HTLV-1 induces IL-2-independent proliferation of Th cells, which may be protected from apoptosis. Subsequent events lead to transformation, the clonal expansion of Th cells, and the development of T-cell lymphoma.

TABLE 2.	Markers	of activation	in HIV ar	nd HTLV	infection

	Marker expression in ^a :					
Marker	HIV in	fection		HTLV infection		Reference(s)
ASY	ASY^b	AIDS	ASY	HAM/TSP	ATL	
β ₂ -Microglobulin	<u> </u>					64, 85, 187, 195, 253, 370
Neopterin	†	1			↑	64, 97, 121, 195, 250, 290
Soluble CD8	<u>†</u>	,	↑	↑		253, 292, 331
IL-2	<u>†</u>	\downarrow	<u>†</u>	<u>†</u>	\downarrow	296, 325
Soluble IL-2R	<u>†</u>	•	<u>†</u>	<u>†</u>	Ť	126, 148, 251, 296, 303, 309
TGF-β	'		'	<u>†</u>	'	245, 326
TNF-α	↑	1		<u>†</u>		188, 195, 251, 296, 335
TNF-β	'	•		'	↑	96, 326
TNF-αR (RI, RII)	↑				'	113, 163, 164
IFN-α	'	↑				221
IFN-γ	↑	†	↑	↑	↑	245, 326
HLA-DR	†	ļ	'	†	†	108, 171
HLA class II	1		↑	†	'	200
CD38	↑		'	ı		168
CD28	1	Ţ				49
CD45RO	↑	•				265
Soluble CD2	j	↑				291
Immunoglobulins	^v IgA	↑ IgG2		↑ IgA		80, 242, 277, 290, 341
	↑ IgM	↑ IgG4		↑ IgM		45, 211, 337, 341
	↑ IgG1	1 280 .		↑ IgG		277, 341
	↑ IgG3			1 280		213
	↓ IgG2					277
	↓ IgG4					211
Lipoproteins	v 15⊖ ·					307
Cholesterol	<u> </u>					307
Fas (CD95)	<u> </u>	^			^	66, 67, 84, 168, 308

 $^{^{}a}$ ↑, increased; ↓, decreased.

metabolic dysfunction is evident in species highly susceptible to AIDS such as humans with HIV infection and rhesus macaques with simian immunodeficiency virus (SIV) infection, no decreases in cysteine or glutathione levels are evident in HIV-infected chimpanzees or SIV-infected African green monkeys (77).

HTLV Infection

The asymptomatic period of HTLV-1 is associated with low virus expression in peripheral blood T cells and low levels of virus in leukemic cells of ATL patients (95, 107, 149, 174, 316). In vitro culture of ATL cells results in virus production, whereas in vivo culture does not (95). This would suggest that such cells producing high levels of virus in vivo become targets of the immune system and are either removed by CTL, antibody-dependent cellular cytotoxicity, or NK cells or are downregulated in vivo by other cells so that virus is not expressed. In contrast, expression of the virus-encoded trans-activating protein, Tax, has been demonstrated in peripheral blood mononuclear cells (PBMC) of seropositive carriers of HTLV-1 prior to the onset of ATL (325). The inflammatory disease induced in HAM/TSP (106, 252) is associated with higher levels of virus expression, and virus is easily detected in PBMC and spinal fluid (21, 237). The level of HTLV-1 detected in central nervous system tissues from HAM/TSP patients is greater than in brain tissue from ATL patients, indicating that there is an increased viral load in HTLV-1-associated neurological disease (82). The contribution of viral burden to ATL and/or HAM/TSP is a neglected area requiring in-depth study.

The frequency of HTLV-1-specific CTL in HAM/TSP patients is markedly higher than in asymptomatic patients (82).

These CTL recognize target cells expressing the Tax protein (82, 155, 167, 181, 259, 260), and in some patients CTL specific for Gag and Env can be detected (82). Naturally occurring variants of Tax impair its recognition by CTL, and these variants have severely reduced transactivation potential (249). While variation in the principal target epitope may contribute to the persistence of the virus, this could, under some conditions, limit virus expression. Higher frequencies of CTL in HAM/TSP patients may correlate with the high viral load, which is not observed in ATL patients (176). While the frequency of Tax-specific CTL is lower in asymptomatic patients (259), the demonstration of such CTL raises speculation about their role in the pathogenesis of HAM/TSP.

HTLV-1 infection is associated with the constitutive expression of IL-2R (71, 114, 136, 230, 233, 360, 362) and in some cases with the production of IL-2 by infected cells (71, 272, 357). In addition, HTLV-1 carriers demonstrate large numbers of activated T cells and a high degree of spontaneous proliferation of in vitro-cultured T cells (71, 272, 357). The level of immune system activation observed in asymptomatic HTLV-1 patients is not likely to be induced by Tax, because it has been shown previously that Tax cannot activate lymphocytes on its own. Activation can be induced by cell-to-cell contact with HTLV-1-infected cells (172). The maintenance of this state of activation is probably dependent on the ability of Tax and subsequently influenced cellular proteins to up-regulate cellular gene expression. HTLV-1-mediated Th-cell activation, combined with virus-mediated induction of cellular gene expression, may permit the virus to both initiate and maintain the lymphoproliferative process. However, this does not explain the quiescence of the virus in the infected, activated Th cell. Alternatively, the down-regulation of CD3 on leukemic cells

^b ASY, asymptomatic phase.

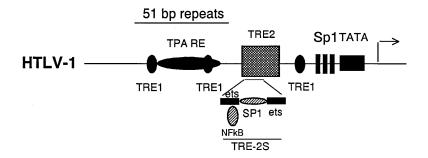




FIG. 3. Schematic representation of the LTR of HTLV-1 and HIV-1. The binding locations of cellular transcription factors are indicated and are further discussed in the text. USF, upstream stimulatory factor; URS, upstream responsive sequence; TCF, T-cell-specific transcription factor. Arrows indicate the start site of transcription.

(152, 330, 363) may suggest that Th-cell activation occurs more continuously in infected cells, perhaps during the asymptomatic period, or may be a reflection of adaptation of the developing tumor cells to immunologic pressures. Infected cells respond indiscriminately to antigen and with no HLA-DR restriction (271). Furthermore, ATL cells have recently been reported to evade NK cell-mediated cytolysis in mice with severe combined immunodeficiency (scid mice), and this evasion appears to be dependent upon low virus expression (89). Th-cell activation and cell proliferation may be a mechanism for the virus infection to be propagated while maintaining a protective intracellular location, thus avoiding detection by immune responses outside of host cells.

EFFECT OF T HELPER CELL ACTIVATION ON RETROVIRAL EXPRESSION

Figure 3 depicts the structure of the long terminal repeat (LTR) regions which mediate replication of HTLV-1 and HIV-1. The striking feature of the LTR regions is how few similar functional promoter elements are shared between HIV-1 and HTLV-1. The HTLV-1 TRE-1 sequences are responsive to activation mediated by the virus-encoded transactivating protein Tax. The response of one TRE-1 to Taxmediated trans-activation is further enhanced by a sequence of four pentanucleotide repeats (TRE-2) located between the second and third TRE-1 sequences (223). A 25-bp sequence (TRE-2S) within TRE-2 is required for cooperative Tax transactivation (324). Mutagenesis studies indicate that the Ets- and NF-κB-binding sites in TRE-2S are dispensable to the cooperative effect (324). Tax does not bind to these enhancer sites but can interact with a number of families of transcription factors to induce gene expression (16, 35, 98, 111, 142, 143, 193, 262, 320, 321, 345, 368). In fact, Tax expression induces phosphorylation and turnover of the inhibitory protein $I\kappa B\alpha$, resulting in constitutive NF-κB activity (186). In addition, Tax activates many cellular genes including IL-2 and its receptor alpha chain (reviewed in references 119, 359, and 360). This deregulation of the expression of cellular genes has been suggested to contribute to alterations in the phenotype of infected cells and the subsequent development of HTLV-1 leukemogenesis via transformation (317, 360). However, the pleiotropic action of Tax is not explained by viral quiescence observed in vivo, unless early T-cell activation genes can support short periods of viral replication and, hence, Tax expression periodically.

The pathways by which Tax exerts its *trans*-activating function appear to be independent of both PKC (278) and cAMP (273) pathways. Up-regulation of transcription by the PKC-stimulating phorbol ester 12-*O*-tetradecanoylphorbol-13-acetate (TPA) is determined by a 60-bp element (TPA RE) which overlaps with one TRE-1 (278). In addition, the integrity of the two 51-bp repeated elements which overlap the first two TRE-1 regions is required for optimal response to phorbol ester (278). Induction by activators of adenylate cyclase (243, 273) is mediated via the octameric cAMP-responsive element located within the TRE-1 sequences and is dependent upon the availability of PKA (162). Thus, HTLV-1 expression may be up-regulated by Tax and by cellular pathways which may be induced in the activated Th cell upon infection.

In comparison with the HTLV LTR, the core elements required for HIV LTR gene expression include the Sp1-binding sites, the TATA box, and the Tat responsive element (TAR element) (Fig. 1) (17, 18, 101, 134, 160, 161). HIV expression is positively regulated by binding of the HIV-encoded Tat trans-activating protein to the TAR element found at the 5' end of all mRNAs (19). The HIV-1 promoter also has several cis-acting regulatory elements in common with cellular gene promoters including NFAT-1, NF-kB, AP-1, and Sp-1. Two NF-κB-binding sites act as a major enhancer of LTR-mediated gene expression and are important for basal LTR activity (18, 150, 241). Deletion of the Sp-1 binding sites results in a marked decrease in Tat-mediated activation (18, 134). Deletion of both NF-κB- and Sp-1-binding sites abolishes Tat-mediated transcription, indicating that Tat may interact with cellular transcription factors to stabilize elongation and to initiate transcription (17, 165, 166, 210). Binding of the nuclear factor of activated T cells (NFAT) is dependent upon Th-cell activation (304). The NFAT-1-binding site falls within a region of the LTR called the negative regulatory element, and deletion of this region results in higher levels of virus expression (212). However, the HIV-1 LTR responds to agents which induce Th-cell activation, including TPA, phytohemagglutinin, and monoclonal antibodies to the cell surface receptors CD3 and CD28 (7, 123, 130, 169, 222, 241, 293, 327). The LTR is also responsive to several cytokines (IL-1, IL-6, and TNF- α) which are up-regulated in the cell during Th cell activation and which may also be provided by accessory cells (11, 78, 153, 175, 184, 225, 253, 344).

We have previously observed, using Jurkat T-cell clones containing integrated LTR-β-galactosidase constructs, marked differences between activation-mediated induction of HIV-1 and HTLV-1 (61). While the HIV-1 LTR was up-regulated by single signals such as CD3 or CD28 pathways, the HTLV-1 LTR was not induced by these pathways. HIV-1 LTR-mediated transcription was increased by TPA, and this response was further enhanced by the calcium (Ca²⁺) ionophore ionomycin. In contrast, the induction of the HTLV-1 LTR by TPA was inhibited by ionomycin (58, 59, 61), suggesting that the triggering of Ca²⁺-dependent pathways in the cell could adversely affect virus expression. Further experiments revealed that the ionomycin-mediated inhibition was not restricted to TPA-induced activation mechanisms, since basal LTR activity and Tax-mediated activation were also compromised by ionomycin. The immunosuppressive drug cyclosporin A suppresses the proliferation of HTLV-1-infected T-cell lines (244). In our experiments, the suppression of Ca²⁺-dependent phosphatase activity by cyclosporin A caused LTR responses in ionomycintreated cells to recover to levels above those obtained in the absence of ionomycin (59), suggesting a complementation of ionomycin and cyclosporin A in HTLV-1 activation. Increased levels of free Ca²⁺ in the presence of cyclosporin A may provide a greater affinity of transcription factors for the LTR and an enhanced LTR induction.

These findings suggest that Th-cell activation, while up-regulatory to HIV-1 LTR function, can be inhibitory to HTLV-1 expression and may contrast two different survival strategies within the human host. In this respect, the presence of proviral DNA and of low levels of viral products, such as Tax, may be sufficient for the expression of IL-2R by the infected T cell. An elevated state of activation resulting in the expression of IL-2R and activation of the IL-2/IL-2R autocrine loop could concurrently provide conditions allowing for the expansion of HTLV-1 provirus-infected cells while suppressing virus expression and cell destruction by the host immune response. This may provide one of several likely events which contribute to the quiescence of the virus during the asymptomatic period of ATL.

T HELPER CELL ACTIVATION AND DISEASE PROGRESSION

HIV and AIDS

Immune dysfunction is a characteristic common to human retrovirus infection, reflecting the central role of Th cells in orchestrating a broad array of immune responses. There are several theories regarding the mechanisms responsible for progression to AIDS, and these are summarized in Table 3. One of many mechanisms suggested to account for the loss of CD4⁺ lymphocytes during progression to AIDS centers on the dysfunctional activation of Th cells. In contrast to HTLV infection, HIV infection is characterized by high virus loads and waves of extracellular viremia due to recurring periods of neutralization and escape. Consequently, there is substantial deposition and trapping of antigen-antibody complexes in the follicular centers of lymph nodes. This saturation of MHC class II-rich regions with virus is likely to substantially impair APC—

TABLE 3. Theories regarding mechanisms mediating disease progression to ATL versus AIDS

ATL	AIDS
Multiple mechanisms singly or combined	Multiple mechanisms singly or combined
IL-2-independent activation resistant to TGF-β regulation of apoptotic removal	Persistent waves of extracellular viremia and high virus load
Persistent activation of Th cells	Impaired APC-Th-cell interac- tion
Reduced expression of β-polymerase and impaired DNA repair	gp120-impaired Th-cell function
•	Anergy
Increased reactive oxygen in- termediates and increased DNA damage	Loss of CD4 renewal capacity
e e	Loss of proliferative responses
Survival of unfit Th cells, accu- mulation of rare mutational events, and eventual ATL	Increased anergy and apoptosis
	Shift in Th-cell population to Th2-predominant response
	Increased oxidative stress and apoptosis

Th-cell interactions (138). Further contributing to Th-cell dysfunction is the loss of APC function of monocytes and dendritic cells in AIDS patients (215, 229). This is supported by the reduced production of IL-12 by macrophages in HIV infection (40) and the ability of IL-12 to restore T-cell responses to recall antigen in HIV-infected individuals (51).

Prior to a significant decline in CD4⁺ cell numbers, immune dysfunction (124, 190, 191, 194, 258, 264, 305, 311), demonstrated by a sequential suppression of activation in response to signalling by antigen, mitogen, and pokeweed mitogen (190, 191, 263, 299, 300) and an increased number of T cells programmed for cell death (227), is evident. These abnormalities in T-cell response are progressive, demonstrated by the loss of proliferative responses to recall antigens and lectins in later stages of infection (226, 305). Th-cell dysfunction may be mediated by the interaction between the CD4 receptor and soluble gp120. In fact, gp120 inhibits the proliferation of PBMC stimulated via the TCR (46, 206) and the expression of IL-2 mRNA in CD4⁺ T cells (254). Proliferation was restored by the addition of exogenous IL-2 (206). A recent report by Schols and Declercq demonstrated that gp120 inhibited CD4⁺ and CD8+ T-cell functions by inducing IL-10 production by monocyte/macrophage cells (294). Elevated levels of IL-10, TNF- α , and IFN- γ have been demonstrated in vivo in individuals infected with HIV-1 (87). The impaired response of normal PBMC (294) and Th1 cells (86) cultured with gp120 was relieved by stimulation through the CD28 receptor. The partial restoration of proliferative responses by CD28 stimulation of Th1 cells also restored IFN- γ and IL-2 production (86). The expression of CD28 is low in HIV-infected individuals (27), and the loss of this recovery route may permit the maintenance of Th-cell dysfunction. Recovery may be further compounded by the fact that IL-10 appears to induce an inhibition of antigen-stimulated proliferation of Th1 cells (91, 92) by downregulating the synthesis of B7 molecules (73).

The induction of Th-cell unresponsiveness is not exclusive to gp120. The HIV Tat protein inhibits T-cell responses to phytohemagglutinin and pokeweed mitogen (343) and to anti-CD3 stimulation (314). Tat directly binds to CD26 to evoke this

inhibition (127, 314), which can be overcome, similarly to gp120-induced unresponsiveness, by exogenous IL-2 or by costimulation via CD28 (314). If Tat and gp120-CD4 interactions are able to induce a state of anergy in T cells, this might result in an eventual loss of T-cell subsets. Additionally, if a shift from Th1 to Th2 occurs, IL-2 levels may not be sufficient to effect recovery.

Progression to disease is marked by an increase in virus expression concurrent with a decrease in the number of CD4+ T cells (125). This may be due to transcriptional activation of the virus by cytokines such as TNF- α and IL-6, whose levels are also elevated at this time (26, 269, 328). However, lower proliferation rates by Th1 cells would be expected to contribute to a lower replicative efficiency, which could not support the observed increase in virus expression associated with progression to AIDS (287). A lower efficiency of Th1 cells to support virus replication could be due to the presence of IFN- γ or to the release of suppressive factors by CD8⁺ T cells. Replication-suppressive factors produced by CD8⁺ T cells include the β -chemokines MIP-1 α , MIP-1 β , and RANTES (56). The entry of HIV-1 into a CD4⁺ cell is dependent upon the coexpression of specific fusion factors. Infection by primary and macrophage-tropic isolates requires the G-protein-coupled seventransmembrane-domain coreceptors, CC CKR3 and CC CKR5 (3, 48, 70, 75, 76), while infection with T-cell-line-adapted isolates and syncytium-inducing primary isolates requires fusin (88), another G-protein-coupled seven-transmembrane-domain coreceptor whose ligand has not yet been identified. Binding of the replication-suppressive chemokines to the coreceptor has been shown to inhibit cell fusion mediated by the HIV envelope glycoproteins (3, 70). However, CD8⁺ suppressive factors also potently suppress HIV LTR-mediated gene expression (42, 62, 63, 216), and this occurs before the onset of RNA transcription (178). In addition, the ability of CD8⁺ T cells to suppress LTR-mediated gene expression does not appear to correlate with improved clinical status (63a). Two distinct CD8+ T-cell suppressor activities have been reported, one which is lost upon disease progression and a second which is maintained at all stages of disease (13). Thus, while virus replication is enhanced during progression to disease, there are mechanisms supporting a down-regulation of virus expression by the preferentially infected Th cell, including suppressive cytokines, chemokines, and a disruption in the normal Th-cell responses to activation. Enhanced virus replication may reflect a change in the phenotype of cells which are infectable and presumably the infection of new cell types brought on by a change in the phenotype of the virus.

It has been suggested that during infection with HIV, the Th-cell population becomes skewed with a loss of Th1/0 cells and a predominance of Th2 cells. This proposed shift from a Th1 to a Th2 profile is associated with decreased cell-mediated immunity (53, 218). The Th1 to Th2 shift hypothesis and its importance in HIV infection remains controversial (118, 218). Taken together with this theory are data which demonstrate preferential replication of HIV-1 in Th2-like cells (218). Elevated levels of soluble CD30 have been detected in HIV-1infected subjects (266). CD30, a member of the TNF/nerve growth factor receptor superfamily, is strongly expressed on activated Th2 clones but not Th1 clones and has been shown to up-regulate HIV-1 expression (68, 286). Strong cell-mediated immunity is detected in HIV⁺ long-term nonprogressors (31, 258), while a type 2 cytokine profile is found in progressors (54, 342). Taken together, the above evidence suggests that HIV can subvert both the cell-mediated and humoral arms of the immune response by causing increased Th-cell dysfunction and decreased Th cell numbers and antigen-specific Th-dependent responses, resulting in skewed cytokine profiles, conditions favoring virus replication, and ultimately Th-cell loss.

Vigorous CTL activity has been reported in HIV-1-infected individuals (reviewed in reference 204), and it has been speculated that this activity may be important in the control of viremia during primary infection (23, 183). The numbers of these HIV-1-specific CTL are reduced in HIV-1-infected subjects with low CD4⁺ T-cell counts (34), and this decrease correlates with disease progression (189). Thus, it appears that a stable pool of CTL is required for the control of HIV-1 infection. A reduced pool of CTL in late-stage disease may be the result of HIV-mediated apoptosis, since CTL specific for Epstein-Barr virus are maintained (34). A reduction in HIV-1-specific CTL activity may contribute to increased viral expression and a reduced clearance of infected cells. In contrast, CTL activity may contribute to the immunopathogenesis of HIV-1 infection through persistent function to inappropriate targets such as Th-APC (54, 138, 204). Insights into the role of CTL in HIV infection have been gleaned through the study of long-term nonprogressors and exposed seronegative individuals (reviewed in reference 54). Indeed, lack of progression is associated with low viral load and with anti-HIV CD8⁺ CTL directed against Gag, Pol, and Env (282). In exposed seronegative individuals, HIV-specific T-cell responses including CTL activity can be detected (54), indicating that limited exposure to the virus may be sufficient to engage a cell-mediated immune response in the absence of HIV-specific antibodies.

HTLV and ATL

On the basis of studies of HTLV-1 transmission, it appears that ATL develops preferentially in individuals infected early by their mothers through breast milk (240). This suggests a link between disease progression and an incubation period spanning many years. It has been suggested that infection with HTLV-1 subtly alters the normal immune system development, such as during ontogeny, and that such events may favor the later development of lymphoma (219). Few infected peripheral blood T cells express HTLV-1 (173), however, infected cells can activate uninfected cells via cell-to-cell contact (353). In this way, a small number of infected cells may maintain a persistent level of immune system activation, possibly precipitating the development of ATL or HAM/TSP. This condition of immune system activation is further supported by the ability of the infected Th cell to maintain the activated state for prolonged periods in an IL-2-independent manner (146, 353). We have postulated that Th-cell activation down-regulates HTLV expression (59, 61). Hollsberg et al. (145) have reported that HTLV-1 mediates T-cell activation through a pathway which is insensitive to the immunosuppressive cytokine TGFβ1. In addition, HTLV-1 infection induces a resistance in previously activated cells to immune suppression by TGF-β1 (145).

While HTLV-1 preferentially infects CD4⁺ Th cells, many cell types may be infected in vitro, including cells of the monocyte-macrophage lineage (72). In addition, Osame et al. (252) reported on the production of a colony-stimulating factor with myelinotoxic activity that is produced in patients with HAM/TSP. Infection of macrophages with HTLV-1 in vivo has not been clearly demonstrated. However, given the low level of virus expression by infected cells in peripheral blood, detection of virus in monocytes has been problematic. It has been suggested that infected macrophages could secrete neurotoxic agents mediating demyelination and inflammation. While this remains to be proven scientifically, it is possible that aberrant activation of Th cells by HTLV-1 will provide the potential for

macrophages to present an altered cytokine profile. Similarly, HIV-infected or gp120-stimulated macrophages produce neurotoxic products as well as inflammatory cytokines, such as TNF- α and IL-1 β , which can further potentiate neurotoxin production (50, 302).

The molecular and cellular events that drive the asymptomatic infected individual to ATL or HAM/TSP remain controversial, however, the low frequency of disease among infected individuals (140) suggests that a rare event or combination of events are required for progression. Some of the current hypotheses postulated to explain the progression from a carrier state to ATL are listed in Table 3. The HTLV-1-encoded Tax protein has been demonstrated to have oncogenic potential (246, 272, 275, 323, 362), and it has been proposed that HTLV-1 infection is a necessary but insufficient step and that secondary or tertiary random, mutational events are required for cellular transformation to ATL. The oncogenic potential could also occur only under specific, possibly rare, cellular conditions when simultaneous Tax expression is present. One mechanism postulated is the development of a prelymphoma state caused by the continuous triggering of T-cell activation (224, 362). However, since progression to disease is uncommon, other mechanisms for the shift to cancer, including previous or subsequent host cell damage, have been proposed (362). The reduced expression of the DNA repair enzyme β-polymerase (158) is a Tax-mediated event which may allow for the accumulations of random, nonlethal, and eventually oncogenic mutational events which could lead to the development of ATL. This model is strengthened by the observation that a variety of different chromosomal abnormalities are frequently found in ATL patients; however, no specific aberration appears to be related to the development of ATL (37, 100, 231). Tax has been shown to activate NF-κB and the HIV-1 LTR by a mechanism which depends upon the production of reactive oxygen intermediates (295; reviewed in reference 356). In addition, mice transgenic for HTLV-1 develop an inflammatory arthropathy which correlates with Tax expression in joints (154). In contrast, Tax also induces the expression of adult T-cell leukemia factor, which is the human homolog of the bacterial coenzyme thioredoxin (147, 356). Thus, while Tax may induce conditions of oxidant stress, which favor gene expression, this condition can be reversed to protect cells from oxidative damage. This type of feedback mechanism may be another way in which virus persists. However, continued rounds of oxidative stress and recovery have the potential to induce irreversible genetic damage to HTLV-infected cells. Given their Tax-related resistance to apoptosis (60), the survival of these cells may allow for the accumulation of mutations, eventually leading to their transformation to ATL cells. As described above, oxidative stress has also been proposed as one of several cellular conditions significant in the progression to AIDS (77, 79).

EFFECT OF HUMAN RETROVIRUS-ENCODED PROTEINS ON APOPTOSIS

The mode of stimulation and the intensity and duration of stimulation of the Th cell determine its subsequent proliferative response. Appropriate engagement of the TCR results in proliferation and IL-2 production. Inappropriate stimulation by engagement of the TCR in the absence of a costimulatory signal may result in a functionally paralyzed or anergic cell or a cell programmed for death by apoptosis (reviewed in reference 306). It has been well documented that one outcome of the in vitro activation of CD4⁺ and CD8⁺ T cells in HIV-1-infected individuals is apoptosis (4, 12, 115, 122, 227, 228). The

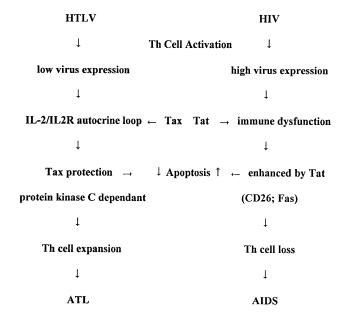


FIG. 4. Infection of Th cells by HTLV or HIV and the consequences of activation on apoptosis.

induction of apoptosis by HIV is multifaceted, being mediated by specific virus-encoded proteins and also by altered cellular pathways, which result from the effects of infection on immune responses. The HIV Tat protein and interactions between the CD4 receptor and gp120 are known mediators of altered T-cell responses; thus, it is not surprising that these mediators have also been identified as apoptotic agents. Apoptosis is induced in T-cell lines by the expression of the HIV env gene (196, 213), and anti-gp120 antibodies can block apoptosis mediated by HIV infection of T cells. Apoptosis induced by cell-to-cell transmission of HIV requires CD4-Env interactions and is not dependent upon new HIV replication (220). Cross-linking of CD4 with gp120 and anti-gp120 or with anti-CD4 antibodies can prime uninfected cells for apoptosis (12, 248). Apoptosis is also induced by cross-linking with gp120 followed by ligation of the TCR, and this sequence of binding events has been suggested as a mechanism of apoptosis in HIV-infected individuals (12, 122). The susceptibility of CD4⁺ T cells to this route of apoptosis depends upon the period between the binding of gp120 and the subsequent encounter with antigen (333). In addition, resistance to gp120-mediated apoptosis is characterized by a down-modulation of CD4 concurrent with a high expression of the anti-apoptosis protein Bcl-2 and is not dependent upon the level of Fas expression by the cells (334). Ligation of CD2 and CD28 in combination with TCR occupancy can rescue CD4⁺ T cells from gp120-mediated apoptosis, and a similar rescue can be provided by IL-4 and IL-2 (334). However, the importance of TCR occupancy can be questioned, since recently activated cells can undergo gp120mediated apoptosis in the absence of TCR ligation (94).

Several mechanisms for gp120-induced apoptosis can be proposed. Binding of gp120 may interfere with normal T-cell regulatory pathways which protect the cell from apoptosis. These pathways include a decrease in p56^{lck} activity (38) or the activation of phosphatidylinositol 3- and 4-kinase activity (276). Alternatively, a disturbed microenvironment in which cytokines normally protective against apoptosis, such as IL-2 and IL-4, are down-regulated by HIV infection could render cells sensitive to apoptosis. In addition, interaction of HIV

gp120 with CD4 can induce the production of a number of cytokines by T cells and macrophages, including TNF- α , IL-6, IL-1, granulocyte-macrophage colony-stimulating factor, and IFN- β (reviewed in reference 256). Finally, apoptosis could reflect the absence of proper accessory cell functions or their costimulatory signals. Underlying all of these proposals is the aberrant activation of cells in the presence of HIV infection.

Recent reports have indicated that infection with HIV-1 results in a greater sensitivity to Fas-mediated apoptosis in human CD4⁺ and CD8⁺ T cells and in macrophages (168, 352). Indeed, infected macrophages can induce Fas-mediated destruction of lymphocytes (10). In addition, HIV-1 Tat has been shown to induce enhanced expression of the Fas receptor, CD95, thus accelerating Fas-mediated apoptosis (352). The ability of Tat to induce apoptosis in various cell types requires nanomolar levels of Tat, while picomolar levels appear to be protective against apoptosis (109, 367). The protective effect of Tat may be mediated by Bcl-2 expression in human cell lines and PBMC (366). However, in contrast to this, infection of Bcl-2-expressing T-cell clones with HIV resulted in an accelerated spreading infection and rapid loss of cell viability (289). Thus, Bcl-2 expression enhanced the ability of T cells to support HIV replication. A similar result was observed in HIVinfected T cells treated with the adenovirus antiapoptosis protein E1B (6). On the other hand, the induction of apoptosis by Tat occurs in the absence of changes in Bcl-2 expression (205) and is associated with the activation of cyclin-dependent kinases (205), which could prevent cells from returning to a quiescent state. A Tat-induced protection from apoptosis could potentially provide conditions required for the lymphoproliferation of cells, leading to neoplasms associated with AIDS (366). In support of this proposal is the finding that Tat acts as a growth factor for cells derived from Kaposi's sarcoma lesions (83).

The contrasting effects of Tat in the studies described above could depend on whether conditions provide for the uptake of Tat, whether Tat is endogenously produced, or whether Tat is acting extracellularly at cell membrane receptors. Tat binds with high affinity to both soluble CD26 and cell surface CD26 on Th cells (127) and, in doing so, inhibits the activity of this receptor (343). CD26 is a surface protease, expressed by CD4⁺ T cells, which mediates recall antigen responses. Although CD26 has been reported to serve as a coreceptor for HIV entry (30), this result could not be confirmed by other laboratories. Additionally, HIV preferentially infects CD4⁺ CD26⁻ T cells (22). There is a selective loss of CD26⁺ cells, including both Fas⁺ and Fas⁻ populations (104), during the progression to AIDS (22). The loss of both these populations suggests that the binding of Tat to CD26 may activate an alternate apoptotic pathway. Further to this, Morimoto et al. (234) have demonstrated that an inhibition of CD26 enzymic activity in CD26⁺ cell lines results in an enhanced sensitivity to apoptosis induced by anti-CD95 antibody.

The Tat protein induces oxidative stress in human T cells (352) which can be abrogated by both inhibitors of Tat and agents which inhibit oxidative stress (81). Ehret et al. examined the ability of Tat to induce oxidative stress and apoptosis in chimpanzee T cells (81). Chimpanzees are susceptible to infection with HIV-1 but are relatively resistant to disease progression (137). While Tat was effectively taken up by chimpanzee T cells, neither oxidative stress nor apoptosis was observed (81). As previously mentioned, unlike HIV-1-infected humans, cysteine metabolic dysfunction is not evident in HIV-infected chimpanzees. The mechanism underlying the resistance of chimpanzee cells to the effects of Tat is not understood but

may provide another avenue to a closer understanding of disease progression in HIV-1-infected individuals.

In addition to Tat, HIV-1 encodes several regulatory proteins, including Rev, which regulates viral mRNA expression, Nef, which induces CD4 internalization, Vpu, which is required for nuclear targeting, and Vpr, which is required for virus release (reviewed in reference 235). While apoptosis has not been demonstrated as an end point, Tat, Rev, Nef, and gp120 possess cytotoxic properties in vitro which may be involved in the pathogenesis of AIDS dementia (112, 209, 214, 351). Vpu, Nef, and Env participate independently in the down-modulation of CD4 in primary cells in vitro (41). The down-modulation is temporal, with Nef acting early and Env and Vpu acting late in the viral life cycle. Further contributing to T-cell dysfunction in HIV infection is the Vpr accessory protein, which has been reported to induce cell cycle arrest in the G_2 phase (15, 267). Vpr expression in the infected cell may protect the cell from apoptosis by preventing entry into the mitotic cycle, further contributing to viral persistence. Thus, apoptosis resulting from HIV-1 infection is a dynamic process, which could be mediated by viral proteins acting independently or in cooperation to undermine the normal function of the Th cell.

In contrast to HIV-1, human T-cell lines infected with HTLV-1 or expressing Tax are protected from Fas-mediated apoptosis (60). This protection is conferred to uninfected activated PBMC by the HTLV-1 Tax protein but not the HIV-1 Tat protein, and it shows a dependence upon basal PKC production in the infected cells (60). In another study, Tax was found to induce apoptosis in Jurkat cells bearing an estrogeninducible fusion protein containing Tax (47); however, apoptosis of these cells required extended Tax expression (9 days). Other reports of the enhancement of apoptosis by Tax used non-T-cell models (99, 355). These contrasting effects of Tax on apoptosis could be, as with Tat, dependent upon the concentration of Tax used. Cell cycle position and regulatory pathways active in the Th cell upon the introduction of Tax could also contribute to the outcome of Tax treatment. This is particularly important to consider, given the pleiotropic effects of Tax on cellular gene activation and suppression.

APOPTOSIS AND DISEASE PROGRESSION

The loss of CD4⁺ T cells is a hallmark of HIV infection. While several mechanisms probably account for this decline, cell death by the normal physiological process of apoptosis is significant in that it can be induced by several pathways (Fig. 4). The effects of HIV on the immune system provide the appropriate conditions for this process. Apoptosis can be mediated by HIV envelope proteins (12, 196, 350), TCR/CD3 interactions (12, 248), expression of Tat (205), cross-linking of CD4 (12, 255), and possibly superantigen (151). The contribution of HIV to the markedly elevated levels of apoptosis is generally believed to occur in HIV-infected lymphoid tissue and PBMC, both in infected and predominantly in uninfected bystander cells (90). Further, a direct correlation between apoptosis and disease progression has recently been demonstrated (116). Apoptosis in lymph nodes of infected individuals is associated with a general state of immune system activation, but earlier reports did not find an association with disease progression or viral load (239). Evidence indicates that pathways mediating apoptosis are up-regulated by HIV infection and may further be associated with disease progression. The role of the Fas pathway in the apoptosis observed in HIV infection has recently been the subject of vigorous study. An increase in Fas-mediated apoptosis in symptomatic HIV-1infected individuals which appears to correlate with an overexpression of Fas antigen during infection has been reported (168). Overexpression of Fas is evident in advanced disease (66, 84, 168, 308); however, the loss of CD4⁺ cells during disease progression is independent of whether the cells are Fas⁺ or Fas⁻ (104). This indicates that HIV-associated apoptosis is not exclusive to the Fas pathway. Importantly, Fas⁺ CD8⁺ T-cell numbers are increased during HIV infection, including the CD8⁺ CD28⁻ subpopulation of these cells (36). A selective targeting of Fas⁺ CD4⁺ Th cells might explain the increase in Fas⁺ CD8⁺ T-cell number.

Th1 cells can be induced to express FasL, whereas Th2 cells express little or no FasL (279). Thus, while cross-linking of gp120-CD4 complexes by antibody would stimulate both Th1 and Th2 cells, FasL would be expressed primarily by Th1 cells with the potential to induce activation-dependent cell death or undergo apoptosis themselves. A clonal expansion of FasLexpressing Th1 cells might lead to a loss of the CD95⁺ Th1 population, thus possibly increasing the proportion of Th2 cells. Naive T cells (CD45RA⁺) express little or no cell surface Fas, but Fas expression can be induced upon activation, while mature (CD45RO⁺) T cells express high levels of Fas (177, 232). Fas-mediated signalling under specific conditions results in an enhanced proliferation of Th cells and increased production of IL-2, IFN-γ, and TNF (2). Not all Fas-expressing cells are susceptible to apoptosis (358); thus, the expression of apoptogen receptors such as Fas alone is not sufficient for apoptosis to occur. Apoptosis may require specific cellular conditions, such as down-regulation of protective genes (i.e., Bcl-2), changes in redox potential, or exhaustion, for cell death to proceed. For instance, one study has demonstrated a high sensitivity of T cells of HIV-1-infected individuals to Fas stimulation by using an antibody unreactive to the apoptotic domain of Fas (308). During progression to AIDS, Th cells are in a high-level state of activation and Fas expression is increased. In the latter stages of the immune response, signalling via Fas may lead to a reduction in the number of T cells via apoptosis. This could result in a change in cytokine profiles as a result of the loss of antigen-specific helper activity. In addition, anergy induced by inappropriate activation may further compound the problem of the apoptotic role of Fas and possibly result in further T-cell loss. Thus, in advanced disease, the Th1 population expressing high levels of Fas could become a target for removal rather than activation. This could be mediated in several ways, including binding of Tat to CD26 or by a Th1to-Th2 shift in cytokine profile. Study of the participation of cytokines in the promotion of activation-induced apoptosis in HIV-1-infected lymphocytes has revealed that type 1 cytokines (IFN- γ , IL-2, and IL-12) are protective against apoptosis (52). In contrast, the Th2 cytokines IL-4 and IL-10 either have no effect or enhance apoptosis (52). Thus, a decline in CD4 counts could represent a shift from a protective Th1 profile to a Th2 cytokine profile, which would permit further apoptosis. The Th1-to-Th2 shift would be further supported by the preference of macrophages to present antigens to Th1 cells. An infected macrophage might deliver an altered activation signal during antigen presentation, which could result in anergy or apoptosis. In this regard, it is important to note that infection of human macrophages results in the de novo expression of FasL and the ability of these cells to induce the Fas-mediated cell death of human PBMC (10).

While CD4⁺ T-cell depletion is central to the development of AIDS, other cell types are also committed to apoptosis throughout the course of infection. CD8⁺ T cells, B cells, and hematopoietic progenitor cells from HIV-1-infected individuals also undergo apoptosis (32, 33, 115, 156, 227, 281). One of

the earliest events following HIV infection is an expansion of the population of activated, memory (CD45RO⁺) CD8⁺ T cells (170, 369). Accompanying the activation and expansion of the CD8 compartment, however, are functional defects in CD8⁺ T-cell responses to activation and recall antigen (123, 339). Associated with the increase in the number of CD8⁺ T cells is the advent of HIV-specific CTL, which are present throughout the asymptomatic period (179, 283, 338). Peripheral CD8⁺ T cells of infected individuals undergo spontaneous and activation-induced apoptosis following a short period of in vitro culture (115, 227). A direct correlation was found between the intensity of spontaneous and anti-CD3-induced apoptosis in both CD4+ and CD8+ T cells from patients and their ex vivo activation state, as evaluated by CD45RO, HLA-DR, and CD38 expression (116). An activation-associated cell death has been observed for the Fas+ CD45RO+ T-cell population in asymptomatic infection (156). The chronic activation state of the immune system induces a down-regulation of Bcl-2 and an up-regulation of Fas in a fraction of CD8+ T cells, which primes these cells for in vitro apoptosis (24). Further to this, in vitro studies indicate that increasing virus load and apoptosis are associated with a shift toward the selective death of CD8⁺ T cells (33). In vivo, a progressive depletion of CD8⁺ T cells and a decline in CD8⁺ T-cell effector function occur upon disease progression (5, 28, 192, 217, 346). Thus, HIV infection induces anergy and apoptosis in the CD8⁺ T-cell compartment. CD8+ T cells expressing Fas and CD28 and demonstrating a Th0 profile (IL-2 and IL-10) have been suggested to be associated with long-term survival. While the susceptibility of this cell population to apoptosis and the potential of these cells for protection from apoptosis by CD28 stimulation remain to be determined, CD8⁺ T cells of patients with progressive disease show reduced Fas expression, loss of CD28 expression, and loss of the production of IL-2 and IL-10 (365). Loss of CD28 expression, combined with lack of IL-2 production, could result in a loss of HLA-1-restricted cytolytic activity and could negatively affect the production of virussuppressive factors (204).

HTLV-1 is mitogenic to resting T cells via the CD2 activation pathway (103), and this effect is mediated by cell-to-cell contact (173). In HTLV-1-infected T-cell lines, CD2 cross-linking induces a cyclosporin A-resistant apoptosis (128). Serum starvation-induced apoptosis of ATL cells is prevented by IL-2 but promoted by glucocorticoid and the activation of PKA (329). Examination of thymic tissue of rabbits infected with a lethal HTLV-1 T-cell line (but not a nonlethal T-cell line) showed evidence of apoptosis (201).

CTL-induced apoptosis has been observed in active spinal cord lesions in patients with HAM/TSP (336). Cells undergoing apoptosis were identified as CD45RO+ T lymphocytes. These authors observed Bcl-2 expression in many of the T cells in the inflammatory lesions but not those susceptible to apoptosis, suggesting that infiltrating T cells could be resistant to apoptosis. This is particularly relevant, since Bcl-2 blocks Taxmediated apoptosis in rat cells (230). In addition, HTLV-1infected individuals with ATL have been reported to undergo a spontaneous remission which was paralleled by a decrease in CD45RO expression by PBMC (318). In another study, the CD45RO cell populations in HTLV-1 carriers and infected individuals with ATL were examined (319). This study found two patterns of CD45RO expression. One was a combination of CD45RO with both high and intermediate fluorescence intensity, but the second pattern, which was exclusively high fluorescence intensity, was associated with disease progression. The intermediate-intensity population expressed far lower levels of Fas antigen than did the high-intensity cells, further

suggesting that intermediate-intensity CD45RO cells may be protective against apoptosis and disease progression (319).

Virus-mediated inhibition of apoptosis has been observed for adenovirus (280), Epstein-Barr virus, (139), and Sindbis virus (203). The dysregulation of cell suicide pathways by virus infection may help to favor the establishment of viral infection of the host by temporarily and partially impairing immune responses. In an HTLV-1-infected person, a limited amount of Tax expression could render infected cells protected from apoptotic elimination during routine immune responses and surveillance. Thus, expansion of the populations of cells expressing HTLV-1 proviral DNA may occur, predisposing these cells to possibly develop into Th-cell cancer. Protection from apoptosis in HTLV-1 infection may be afforded by low-level expression of Tax and the subsequent activation of the IL-2/ IL-2R autocrine loop (Fig. 4). In ATL, protection from apoptosis may eventually allow the more aggressive development of lymphomas over time. Also contributing to the establishment of this type of cancer is the ability of ATL cells to evade immune responses such as NK-cell cytolysis (89). Given the extended asymptomatic period of HTLV-1 infection and the infrequent development of ATL, progression to disease probably depends on the accumulation of a complex series of events including persistent T-cell activation, fluctuating viral quiescence, and lifelong genetic mutations and alterations over

CONCLUSIONS

The specific events which affect Th cells and determine the onset of the progression to ATL and AIDS are still not precisely defined. However, a growing body of evidence supports a central role of T-cell activation in the contrasting pathogenesis of AIDS versus ATL. The differences found in the Th-cell responses to T-cell activation pathways between HTLV-1 and HIV-1 suggest that these two viruses have evolved very different strategies for utilizing and residing in the same host cell type. They both abrogate T-cell activation signals to control their own replication, as well as influencing factors affecting the depletion and expansion of Th-cell populations. However, they differ in the Th-cell activation pathways they abrogate. Those differences must be contrasted in the context of the central biological function that Th cells have and of the loss versus expansion of these cell populations in the pathogenesis of HIV and HTLV infection, respectively. Because progression to ATL occurs infrequently in infected individuals, the events that trigger progression may be rare. In addition, it seems likely that, as with HAM/TSP, higher virus expression may be required for progression to disease. It must also be considered that certain events may need to occur, possibly in a specific order during infection. The observation that T-cell activation limits HTLV expression in vitro may indicate that infrequent events are required during the asymptomatic phase for the stepwise progression to ATL. In contrast, the course of clinical progression to AIDS is more predictable, despite the small population of long-term nonprogressors. This group may be critical to defining the relationship between immune system activation, immune system suppression, and Th-cell loss during HIV infection. A focus on understanding the impact of retroviral infection on Th-cell population dynamics during infection and disease progression is important to further our knowledge of these contrasting human retrovirus-mediated diseases.

ACKNOWLEDGMENTS

K.F.T.C. is supported through a Postdoctoral Fellowship from the Medical Research Council of Canada and the National Health Re-

search and Development Program of Health Canada. The Department of Virology BPRC is supported by the EU Biomedical Programme (grant BMH4-CT95-0206).

REFERENCES

- Abehsira-Amar, O., M. Gibert, M. Joliy, J. Theze, and D. L. Jankovic. 1992. IL-4 plays a dominant role in the differential development of Th0 into Th1 and Th2 cells. J. Immunol. 148:3820–3829.
- Alderson, M. R., R. J. Armitage, E. Maraskovsky, T. W. Tough, E. Roux, K. Schooley, F. Ramsdell, and D. H. Lynch. 1993. Fas transduces activation signals in normal human T-lymphocytes. J. Exp. Med. 178:2231–2235.
- Alkhatib, G., C. Combadiere, C. C. Broder, Y. Feng, P. E. Kennedy, P. M. Murphy, and E. A. Berger. 1996. CC CKR5: A RANTES, MIP-1α, and MIP-1β receptor as a fusion cofactor for macrophage-tropic HIV-1. Science 272:1955–1958.
- Amiesen, J. C., and A. Capron. 1991. Cell dysfunction and depletion in AIDS: the programmed cell death hypothesis. Immunol. Today 12:102–105.
- Anderson, R. E., S. C. Shiboski, R. Royce, N. P. Jewell, W. Lang, and A. Winkelstein, Jr. 1991. CD8+ lymphocytes and progression to AIDS in HIV-infected men: some observations. AIDS 5:213–215.
- Antoni, B. A., P. Sabbatini, A. B. Rabson, and E. White. 1995. Inhibition of apoptosis in human immunodeficiency virus-infected cells enhances virus production and facilitates persistent infection. J. Virol. 69:2384–2392.
- Asjo, B., D. Cefai, P. Debre, Y. Dudoit, and B. Autran. 1993. A novel mode of human immunodeficiency virus type 1 (HIV-1) activation: ligation of CD28 alone induces HIV-1 replication in naturally infected lymphocytes. J. Virol. 67:4395–4398.
- Auwerx, J., and P. Sassone-Corsi. 1991. IP-1: a dominant inhibitor of Fos/Jun whose activity is modulated by phosphorylation. Cell 64:983–993.
- Azuma, M., D. Ito, H. Yagita, K. Okumura, J. H. Phillips, L. Lanier, and C. Samoza. 1993. B70 is a second ligand for CTLA-4 and CD28. Nature (London) 366:76–79.
- Badley, A. D., J. A. McElhinny, P. J. Leibson, D. H. Lynch, M. R. Alderson, and C. V. Paya. 1996. Upregulation of Fas ligand expression by human immunodeficiency virus in human macrophages mediates apoptosis of uninfected T lymphocytes. J. Virol. 70:199–206.
- Baldari, C. T., G. Macchia, A. Massone, and J. L. Telford. 1992. p21^{ras} contributes to HIV-1 activation in T-cells. FEBS Lett. 304:261–264.
- Banda, N. K., J. Bernier, D. K. Kurahara, R. Kurrie, N. Haigwood, R.-P. Sekaly, and T. H. Finkel. 1992. Crosslinking CD4 by human immunodeficiency virus gp120 primes T cells for activation-induced apoptosis. J. Exp. Med. 176:1099–1106.
- Barker, T. D., D. Weissman, J. A. Daucher, K. M. Roche, and A. S. Fauci. 1996. Identification of multiple and distinct CD8⁺ T cell suppressor activities. J. Immunol. 156:4476–4483.
- 14. Barre-Sinousi, F., J. C. Chermann, F. Rey, M. T. Nugeyre, S. Chamarel, T. Gruest, C. Dauguet, C. Axler-Blin, F. Vezin-Brun, C. Rouzioux, W. Rozenbaum, and L. Montagnier. 1983. Isolation of a T-lymphocyte retrovirus from a patient at risk for acquired immune deficiency syndrome (AIDS). Science 220:868–870.
- Bartz, S., M. E. Rogel, and M. Emerman. 1996. Human immunodeficiency virus type 1 cell cycle control: Vpr is cytostatic and mediates G₂ accumulation by a mechanism which differs from DNA damage checkpoint control. J. Virol. 70:2324–2331.
- 16. Beraud, C., S.-C. Sun, P. Ganchi, D. W. Ballard, and W. C. Greene. 1994. Human T-cell leukemia virus type 1 Tax associates with and is negatively regulated by the NF-κB p100 gene product: implications for viral latency. Mol. Cell. Biol. 14:1374–1382.
- Berkhout, B., A. Gatignol, A. B. Rabson, and K.-T. Jeang. 1990. TAR-independent activation of the HIV-1 LTR: evidence that tat requires specific regions of the promoter. Cell 62:757–767.
- Berkhout, B., and K.-T. Jeang. 1992. Functional roles for the TATA promoter and enhancers in basal and Tat induced expression of the human immunodeficiency virus type 1 long terminal repeat. J. Virol. 66:139–149.
- Berkhout, B., R. H. Silverman, and K.-T. Jeang. 1989. Tat trans-activates the human immunodeficiency virus through a nascent RNA target. Cell 59:273–282.
- Betz, M., and B. S. Fox. 1990. Regulation and development of cytochrome c-specific IL-4 producing T cells. J. Immunol. 145:1046–1052.
- Bhagavati, S., G. Ehrlich, R. W. Kula, S. Kwok, J. Sninsky, V. Udani, and B. J. Poiesz. 1988. Detection of human T-cell lymphoma/leukemia virus type I DNA and antigen in spinal fluid and blood of patients with chronic progressive myelopathy. N. Engl. J. Med. 318:1141–1147.
- Blazquez, M. V., J. A. Madueno, R. Gonzalez, R. Jurado, W. W. Bachovchin, J. Pena, and E. Munoz. 1992. Selective decrease of CD26 expression in T cells from HIV-1-infected individuals. J. Immunol. 149:3073–3077.
- Borrow, P., H. Lewicki, B. H. Hahn, G. M. Shaw, and M. B. Oldstone. 1994. Virus-specific CD8⁺ cytotoxic T-lymphocyte activity associated with control of viremia in primary human immunodeficiency virus type 1 infection. J. Virol. 68:6103–6110.
- Boudet, F., H. Lecoeur, and M.-L. Gougeon. 1996. Apoptosis associated with ex vivo down-regulation of bcl-2 and up-regulation of Fas in potential

- cytotoxic CD8⁺ T lymphocytes during HIV infection. J. Immunol. **156**: 2282–2293.
- Boussiotis, V. A., G. J. Freeman, J. G. Gribben, J. Daley, G. Gray, and L. M. Nadler. 1993. Activated human B lymphocytes express three CTLA-4 counterreceptors that costimulate T-cell activation. Proc. Natl. Acad. Sci. USA 90:11059–11063.
- Breen, E. C., A. R. Rezai, K. Nakajima, G. N. Beall, R. T. Mitsuyasu, T. Hirano, T. Kishimoto, and O. Martinez-Maza. 1990. Infection with HIV is associated with elevated IL-6 levels and production. J. Immunol. 144:480– 484.
- Brinchmann, J. E., J. H. Dobloug, B. H. Heger, L. L. Haaheim, M. Sannes, and T. Egeland. 1994. Expression of costimulatory molecule CD28 on T cells in human immunodeficiency virus type 1 infection: functional and clinical correlations. J. Infect. Dis. 169:730–738.
- Brinchmann, J. E., G. Gaudernack, and F. Vartdal. 1990. CD8⁺ T cells inhibit HIV infection in naturally infected CD4⁺ T cells. J. Immunol. 144:2961–2966.
- Brunet, J. F., F. Denizot, M. F. Luciani, M. Roux-Dosseto, M. Suzan, M. G. Mattei, and P. Golstein. 1987. A new member of the immunoglobulin superfamily—CTLA-4. Nature (London) 328:267–270.
- Callebaut, C., B. Krust, E. Jacotot, and A. G. Hovanessian. 1993. T cell activation antigen, CD26, as a cofactor for entry of HIV in CD4+ cells. Science 262:2045–2050.
- Cao, Y., L. Qin, L. Zhang, J. Safrit, and D. D. Ho. 1995. Virologic and immunologic characterization of long-term survivors of human immunodeficiency virus type 1 infection. N. Engl. J. Med. 332:201–208.
- Carbonari, M., M. Cibati, M. Cherchi, D. Sbarigia, A. M. Pesce, L. Dell'anna, A. Modica, and M. Fiorilli. 1994. Detection and characterization of apoptotic peripheral blood lymphocytes in HIV infection and cancer chemotherapy by a novel flow immunocytometric method. Blood 83:1245

 1255
- 33. Carbonari, M., M. Cibati, A. M. Pesce, D. Sbarigia, P. Grossi, G. D'Offizi, G. Luzi, and M. Fiorilli. 1995. Frequency of provirus-bearing CD4⁺ cells in HIV type 1 infection correlates with extent of in vitro apoptosis of CD8⁺ but not of CD4⁺ cells. AIDS Res. Hum. Retroviruses 11:789–794.
- 34. Carmichael, A., X. Jin, P. Sissons, and L. Borysiewicz. 1993. Quantitative analysis of human immunodeficiency virus type 1 (HIV-1)-specific cytotoxic T lymphocyte (CTL) response at different stages of HIV-1 infection: differential CTL responses to HIV-1 and Epstein-Barr virus in late disease. J. Exp. Med. 177:249–256.
- Caron, C., R. Rousset, C. Beraud, V. Moncollin, J.-M. Egly, and P. Jalinot. 1993. Functional and biochemical interaction of the HTLV-1 Tax1 transactivator with TBP. EMBO J. 12:4269–4278.
- 36. Caruso, A., A. Cantalamessa, S. Licenziati, L. Peroni, E. Prati, F. Martinelli, A. D. Canaris, S. Folghera, R. Gorla, A. Balsari, R. Cattaneo, and A. Turano. 1994. Expression of CD28 on CD8(+) and CD4(+) lymphocytes during HIV infection. Scand. J. Immunol. 40:485–490.
- 37. Catovsky, D., M. F. Greaves, M. Rose, D. A. G. Galton, A. W. G. Goolden, D. R. McCluskey, J. M. White, I. Lampert, G. Bourikas, R. Ireland, A. I. Brownwell, J. M. Bridges, W. A. Blattner, and R. C. Gallo. 1982. Adult T-cell lymphoma-leukemia in blacks from the West Indies. Lancet i:639–643
- Cefai, D., M. Ferrer, N. Serpente, T. Idziorek, A. Dautry-Varsat, P. Debre, and G. Bismuth. 1992. Internalization of HIV glycoprotein gp120 is associated with down-modulation of membrane CD4 and p56^{lck} together with impairment of T cell activation. J. Immunol. 149:285–294.
- Chang, T. L., C. M. Shea, S. Urioste, R. C. Thompson, W. H. Boom, and A. K. Abbas. 1990. Heterogeneity of helper/inducer T lymphocytes. III. Responses of IL-2 and IL-4 producing (Th1 and Th2) clones to antigens presented by different accessory cells. J. Immunol. 145:2803–2808.
- Chehimi, J., S. E. Starr, I. Frank, A. D'Andrea, X. Ma, R. R. MacGregor, J. Sennelier, and G. Trinchieri. 1994. Impaired interleukin 12 production in human immunodeficiency virus-infected patients. J. Exp. Med. 179:1361– 1366
- Chen, B. K., R. T. Gandhi, and D. Baltimore. 1996. CD4 down-modulation during infection of human T cells with human immunodeficiency virus type 1 involves independent activities of vpu, env, and nef. J. Virol. 70:6044–6053.
- Chen, C.-H., K. J. Weinhold, J. A. Bartlett, D. P. Bolognesi, and M. Greenberg. 1993. CD8+ T lymphocyte-mediated inhibition of HIV-1 long terminal repeat transcription: a novel antiviral mechanism. AIDS Res. Hum. Retroviruses 9:1079–1086.
- Chen, D., and E. V. Rothenberg. 1994. Interleukin 2 transcription factors as molecular targets of cAMP inhibition: delayed inhibition kinetics and combinatorial transcription roles. J. Exp. Med. 179:931–942.
- Chen, Y., V. K. Kuchroo, J. Inobe, D. A. Hafler, and H. L. Weiner. 1994.
 Regulatory T cell clones induced by oral tolerance: suppression of autoimmune encephalomyelitis. Science 265:1237–1240.
- Chen, Y.-M. A., A. Okayama, T.-H. Lee, N. Tachibana, N. Mueller, and M. Essex. 1991. Sexual transmission of human T-cell leukemia virus type I associated with the presence of anti-Tax antibody. Proc. Natl. Acad. Sci. USA 88:1182–1186.
- 46. Chirmule, N., T. W. McCloskey, R. Hu, V. S. Kalyanaraman, and S. Pahwa.

- 1995. HIV gp120 inhibits T cell activation by interfering with expression of costimulatory molecules CD40 ligand and CD80 (B71). J. Immunol. 155: 917–924.
- Chlichlia, K., G. Moldenhauer, P. T. Daniel, M. Busslinger, L. Gazzolo, V. Schirrmacher, and K. Khazaie. 1993. Immediate effects of reversible HTLV-1 tax function: T-cell activation and apoptosis. Oncogene 10:269–277
- 48. Choe, H., M. Farzan, Y. Sun, N. Sullivan, B. Rollins, P. D. Ponath, L. Wu, C. R. Mackay, G. LaRosa, W. Newman, N. Gerard, C. Gerard, and J. Sodroski. 1996. The β-chemokine receptors CCR3 and CCR5 facilitate infection by primary HIV-1 isolates. Cell 85:1135–1148.
- Choremi-Papadopoulou, H., V. Viglis, P. Gargalianos, T. Kordossis, A. Iniotaki-Theodoraki, and J. Klosmidis. 1994. Downregulation of CD28 surface antigen on CD4⁺ and CD8⁺ T lymphocytes during HIV-1 infection. J. Acquired Immune Defic. Syndr. 7:245–253.
- Chung, I. Y., and E. N. Benveniste. 1990. Tumor necrosis factor-α production by astrocytes. Induction by lipopolysaccharide, IFNγ, and IL-1β. J. Immunol. 144:2999–3007.
- Clerici, M., D. R. Lucey, J. A. Berzofsky, L. A. Pinto, T. A. Wynn, S. P. Blatt, M. J. Dolan, C. W. Hendrix, S. F. Wolf, and G. M. Shearer. 1993. Restoration of HIV-specific cell-mediated immune responses by interleukin-12 in vitro. Science 262:1721–1724.
- 52. Clerici, M., A. Sarin, R. L. Coffman, T. A. Wynn, S. P. Blatt, C. W. Hendrix, S. F. Wolf, G. M. Shearer, and P. A. Henkart. 1994. Type1/type2 cytokine modulation of T-cell programmed cell death as a model for human immunodeficiency virus pathogenesis. Proc. Natl. Acad. Sci. USA 91:11811–11815
- Clerici, M., and G. M. Shearer. 1993. A TH1-TH2 switch is a critical step in the etiology of HIV infection. Immunol. Today 15:312–315.
- Clerici, M., and G. M. Shearer. 1996. Correlates of protection in HIV infection and the progression of HIV infection to AIDS. Immunol. Lett. 51:69–73.
- Clevers, H., B. Alarcon, T. Wileman, and C. Terhorst. 1988. The T-cell receptor/CD3 complex: a dynamic protein ensemble. Annu. Rev. Immunol. 6:629–662.
- Cocchi, F., A. L. DeVico, A. Garzino-Demo, S. K. Arya, R. C. Gallo, and P. Lusso. 1995. Identification of RANTES, MIP-1α, and MIP-1β as the major HIV-suppressive factors produced by CD8⁺ T cells. Science 270:1811–1815.
- Cocks, B. G., C.-C. J. Chang, J. M. Carbllido, H. Yssel, J. E. de Vries, and G. Aversa. 1995. A novel receptor involved in T-cell activation. Nature (London) 376:260–263.
- Copeland, K. F. T., A. G. M. Haaksma, D. Derse, J. Goudsmit, and J. L. Heeney. 1993. Cytochemical analysis of human T-cell leukaemia virus I LTR-regulated beta-galactosidase gene expression using a novel integrated cell system. J. Virol. Methods 45:161–167.
- Copeland, K. F. T., A. G. M. Haaksma, J. Goudsmit, and J. L. Heeney. 1994.
 Calcium-mediated inhibition of phorbol ester and *tax trans*-activation of the human T-cell leukaemia virus-1. J. Gen. Virol. 75:1623–1631.
- Copeland, K. F. T., A. G. M. Haaksma, J. Goudsmit, P. H. Krammer, and J. L. Heeney. 1994. Inhibition of apoptosis in T cells expressing human T cell leukemia virus type I tax. AIDS Res. Hum. Retroviruses 10:1259–1268.
- Copeland, K. F. T., P. J. Hendrikx, A. G. M. Haaksma, S. Fiering, R. van Lier, J. Goudsmit, and J. L. Heeney. 1995. Comparison of the response to T-cell activation by integrated HIV-1 and HTLV-1 LTR-lacZ vectors. Virology 209:633–636.
- Copeland, K. F. T., P. J. McKay, and K. L. Rosenthal. 1995. Suppression of activation of the human immunodeficiency virus long terminal repeat by CD8+ T-cells is not lentivirus specific. AIDS Res. Hum. Retroviruses 11: 321-1326.
- Copeland, K. F. T., P. J. McKay, and K. L. Rosenthal. 1996. Suppression of the human immunodeficiency virus long terminal repeat by CD8+ T-cells is dependent on the NFAT-1 element. AIDS Res. Hum. Retroviruses 12:143– 148.
- 63a.Copeland, K. F. T., et al. Unpublished observations.
- 64. Cuthbert, R. J. G., C. A. Ludlam, J. Tucker, C. M. Steel, D. Beatson, S. Rebus, and J. F. Peutherer. 1990. Five year prospective study of HIV infection in the Edinburgh hemophiliac cohort. Br. Med. J. 301:956–960.
- 65. D'Andrea, A., M. Rengaraju, N. M. Valiante, J. Chehimi, M. Kubin, M. Aste, S. H. Chan, M. Kobayashi, D. Young, E. Nickbarg, R. Chizzonite, S. F. Wolf, and G. Trinchieri. 1992. Production of natural killer cell stimulatory factor interleukin 12 by peripheral blood mononuclear cells. J. Exp. Med. 176:1387–1398.
- 66. Debatin, M. K., A. Fahrig-Farssner, S. Enenkel-Stoodt, W. Kreuz, A. Bender, and P. H. Krammer. 1994. High expression of apo-1 (CD95) on T lymphocytes from human immunodeficiency virus-1-infected children. Blood 83:3101–3103.
- Debatin, K.-M., C. K. Goldman, R. Bamford, T. A. Waldman, and P. H. Krammer. 1990. Monoclonal antibody-mediated apoptosis in adult T-cell leukaemia. Lancet 335:497–500.
- Del Prete, G., M. de Carli, F. Almerigogna, C. K. Daniel, M. M. D'Elios, G. Zancuoghi, F. Vinante, G. Pizzolo, and S. Romagnani. 1995. Preferential

expression of CD30 by human CD4⁺ T cells producing Th2-type cytokines. FASEB J. 9:81–86.

- 69. Del Prete, G. F., M. De Carli, C. Mastromauro, D. Macchia, R. Biagiotti, M. Ricci, and S. Romagnani. 1991. Purified protein derivative of Mycobacterium tuberculosis and excretory-secretory antigen(s) of Toxocara canis expand in vitro human T cells with stable and opposite (type 1 T helper or type 2 T helper) profile of cytokine production. J. Clin. Invest. 88:346–350.
- Deng, H. K., R. Liu, W. Ellmeier, S. Choe, D. Unutmaz, M. Burkhart, P. Di Marzio, S. Marmom, R. E. Sutton, C. M. Hill, C. B. Davis, S. C. Peiper, T. J. Schall, D. R. Littman, and N. R. Landau. 1996. Identification of a major co-receptor for primary isolates of HIV-1. Nature (London) 381:661–666.
- Depper, J. M., W. J. Leonard, M. Kronke, T. A. Waldmann, and W. C. Greene. 1984. Augmented T cell growth factor receptor expression in HTLV-1-infected human leukemic T cells. J. Immunol. 133:1691–1695.
- De Revel, T., A. Mabondzo, G. Gras, B. Delord, P. Roques, F. Boussin, Y. Neveux, M. Bahuau, H. J. A. Fleury, and D. Dormont. 1993. In vitro infection of human macrophages with human T-cell leukemia virus type I. Blood 81:1598–1606.
- Ding, L., P. Linsley, L.-Y. Huang, R. Germain, and E. Shevach. 1993. IL-10
 inhibits macrophage co-stimulatory activity by selectively inhibiting the
 upregulation of B7 expression. J. Immunol. 151:1224–1239.
- Donaldson, Y. K., J. E. Bell, J. W. Ironside, R. P. Brettle, J. R. Robertson, A. Busuttil, and P. Simmonds. 1994. Redistribution of HIV outside the lymphoid system with onset of AIDS. Lancet 343:382–385.
- 75. Dorantz, B. J., J. Rucker, Y. Yi, R. J. Smyth, M. Samson, S. C. Peiper, M. Parmentier, R. G. Collman, and R. W. Doms. 1996. A dual-tropic primary HIV-1 isolate that uses fusin and the β-chemokine receptors CKR-5, CKR-3, and CKR-2b as fusion cofactors. Cell 85:1149–1158.
- 76. Dragic, T, V. Litwin, G. P. Allaway, S. R. Martin, Y. Huang, K. A. Nagashima, C. Cayanan, P. J. Maddon, R. A. Koup, J. P. Moore, and W. A. Paxton. 1996. HIV-1 entry into CD4⁺ cells is mediated by the chemokine receptor CC-CKR-5. Nature (London) 381:667–673.
- 77. Droge, W., K. M. Murthy, C. Stahl-Hennig, S. Hartung, R. Plesker, S. Rouse, E. Peterhans, R. Kinscherf, T. Fischbach, and H.-P. Eck. 1993. Plasma amino acid dysregulation after lentiviral infection. AIDS Res. Hum. Retroviruses 9:807–809.
- 78. Duh, E. J., W. J. Maury, T. M. Folks, A. S. Fauci, and A. B. Rabson. 1989. Tumor necrosis factor alpha activates human immunodeficiency virus type 1 through induction of nuclear factor binding to the NF-kappa B sites in the long terminal repeat. Proc. Natl. Acad. Sci. USA 86:5794–5798.
- Eck, H.-P., H. Mertens, H. Rasokat, G. Fatgenheuer, C. Pohl, M. Schrappe, V. Daniel, H. Naher, D. Petzoldt, P. Drings, and W. Droge. 1992. T4+ cell numbers are correlated with plasma glutamate and cystine levels: association of hyperglutamataemia with immunodeficiency in diseases with different etiologies. Int. Immunol. 4:7–13.
- Edelman, S., and S. Zola-Pazner. 1990. Response of mononuclear cells from HIV-infected patients to B-cell mitogens: correlation with immunological and clinical features of disease progression. AIDS 4:859–864.
- Ehret, A., M. O. Westendorp, Ingrid Herr, K.-M. Debatin, J. L. Heeney, R. Frank, and P. H. Krammer. 1996. Resistance of chimpanzee T cells to human immunodeficiency virus type 1 Tat-enhanced oxidative stress and apoptosis. J. Virol. 70:6502–6507.
- Elovaara, I., S. Koenig, A. Yambusu Brewah, R. M. Woods, T. Lehky, and S. Jacobson. 1993. High human T cell lymphotropic virus type 1 (HTLV-1)-specific precursor cytotoxic T lymphocyte frequencies in patients with HTLV-1-associated neurological disease. J. Exp. Med. 177:1567–1573.
- Ensoli, B., G. Barillari, S. Z. Salahuddin, R. C. Gallo, and F. Wong-Staal.
 1990. Tat protein of HIV-1 stimulates growth of cells derived from Kaposi's sarcoma lesions of AIDS patients. Nature (London) 345:84–86.
- 84. Estaquier, J., T. Idziorek, W. Zou, D. Emilie, C.-M. Farber, J.-M. Bourez, and J. C. Amieson. 1995. Thelper type I/T helper type 2 cytokines and T cell death: preventive effect of interleukin 12 on activation-induced and CD95 (FAS/APO-1)-mediated apoptosis of CD4⁺ T cell from human immunodeficiency virus-infected persons. J. Exp. Med. 182:1759–1767.
- Fahey, J. L., J. M. G. Taylor, R. Deteld, B. Hofmann, R. Melmed, P. Nishanian, and J. V. Giorgi. 1990. The prognostic value of cellular and serologic markers in infection with human immunodeficiency virus type 1. N. Eng. J. Med. 322:166–172.
- Faith, A., H. Yssel, R. E. O'Hehir, and J. R. Lamb. 1996. Reversal of the inhibitory effects of HIV-gp120 on CD4⁺ T cells by stimulation through the CD28 pathway. Clin. Exp. Immunol. 105:225–230.
- Fan, J., H. Z. Bass, and J. L. Fahey. 1993. Elevated IFNτ and decreased IL-2 gene expression are associated with HIV infection. J. Immunol. 151: 5031 5040
- Feng, Y., C. C. Broder, P. E. Kennedy, and E. A. Berger. 1996. HIV-1 entry cofactor: functional cDNA cloning of a seven-transmembrane domain, Gprotein coupled receptor. Science 272:872–877.
- Feuer, G., S. A. Stewart, S. M. Baird, F. Lee, R. Feuer, and I. S. Y. Chen. 1995. Potential role of natural killer cells in controlling tumorigenesis by human T-cell leukemia viruses. J. Virol. 69:1328–1333.
- Finkel, T. H., G. Tudor-Williams, N. K. Banda, M. F. Cotton, T. Curiel, C. Monks, T. W. Baba, R. M. Ruprecht, and A. Kupfer. 1995. Apoptosis occurs

- predominantly in bystander cells and not in productively infected cells of HIV- and SIV-infected lymph nodes. Nat. Med. 1:129–134.
- Fiorentino, D. F., M. W. Bond, and T. R. Mosmann. 1989. Two types of mouse T helper cell. IV. Th2 clones secrete a factor that inhibits cytokine production by Th1 clones. J. Exp. Med. 170:2081–2095.
- Fiorentino, D. F., A. Zlotnik, P. Vieira, T. R. Mosmann, M. Howard, K. W. Moore, and A. Ogarra. 1991. IL-10 acts on the antigen-presenting cell to inhibit cytokine production by Th1 cells. J. Immunol. 146:3444–3451.
- 93. Firestein, G. S., W. D. Roederer, J. A. Laxer, K. S. Townsend, C. T. Weaver, J. T. Hom, J. Linton, B. E. Torbett, and A. L. Glasebrook. 1989. A new murine CD4+ T cell subset with an unrestricted cytokine profile. J. Immunol. 143:518-525.
- Foster, S., P. Beverley, and R. Aspinall. 1995. gp120-induced programmed cell death in recently activated T cells without subsequent ligation of the T cell receptor. Eur. J. Immunol. 25:1778–1782.
- Franchini, G., F. Wong-Staal, and R. C. Gallo. 1984. Human T-cell leukemia virus (HTLV-1) transcripts in fresh and cultured cells of patients with adult T-cell leukemia. Proc. Natl. Acad. Sci. USA 81:6207–6211.
- Fuchs, D., A. Haussen, G. Reibregger, E. R. Werner, G. Werner-Felmayer, M. P. Dierich, and H. Wachter. 1989. Interferon gamma concentrations are increased in sera from individuals infected with human immunodeficiency virus type 1. J. Acquired Immune Defic. Syndr. 2:158–162.
- Fuchs, D., G. M. Shearer, R. N. Boswell, D. R. Lucey, M. Clerici, G. Reibnegger, E. Werner, R. A. Zajaz, and H. Wachter. 1991. Negative correlation between blood cell counts and serum neopterin concentration in patients with HIV-1 infection. AIDS 5:209–213.
- Fujii, M., H. Tsuchiya, T. Chuhjo, T. Akizawa, and M. Seiki. 1992. Interaction of HTLV-1 Tax1 with p67^{SRF} causes the aberrant induction of cellular immediate early genes through CarG boxes. Genes Dev. 6:2066– 2076.
- Fujita, M., and H. Shiku. 1995. Differences in sensitivity to induction of apoptosis among rat fibroblast cells transformed by HTLV-I tax gene or cellular nuclear oncogenes. Oncogene 11:15–20.
- 100. Fukuhara, S., Y. Hinuma, Y. I. Gotoh, and H. Uchino. 1983. Chromosome aberrations in T lymphocytes carrying adult T-cell leukemia-associated antigens (ATLA) from healthy adults. Blood 61:205–207.
- 101. Garcia, J. A., D. Harrich, E. Soutanakis, F. Wu, R. Mitsuyasu, and R. B. Gaynor. 1989. Human immunodeficiency virus type 1 LTR TATA and TAR region sequences required for transcriptional regulation. EMBO J. 8:765–779.
- 102. Gardner, P. 1989. Calcium and T lymphocyte activation. Cell 59:15-20.
- Gazzolo, L., and M. Duc Dodon. 1987. Direct activation of resting T cells by HTLV-1. Nature (London) 326:714–717.
- 104. Gehri, R., S. Hahn, M. Rothen, M. Steuerwald, R. Nuesch, and P. Erb. 1996. The Fas receptor in HIV infection: expression on peripheral blood lymphocytes and role in the depletion of T cells. AIDS 10:9–16.
- 105. Gelfand, E. W., G. B. Mills, R. K. Cheung, J. W. W. Lee, and S. Grinstein. 1987. Transmembrane ion fluxes during activation of human T lymphocytes: Role of Ca²⁺, Na⁺/H⁺ exchange and phospholipid turnover. Immunol. Rev. 95:59–87.
- 106. Gessain, A., F. Barin, and J. C. Vernant. 1985. Antibodies to human T-lymphotropic virus type-I in patients with tropical spastic paraparesis. Lancet ii:407–410.
- 107. Gessain, A., A. Louie, O. Gout, R. C. Gallo, and G. Franchini. 1991. Human T-cell leukemia-lymphoma virus type 1 (HTLV-1) expression in fresh peripheral blood mononuclear cells from patients with tropical spastic paraparesis/HTLV-1-associated myelopathy. J. Virol. 65:1628–1633.
- 108. Gessain, A., F. Saal, M.-L. Giron, J. Lasneret, S. Lagaye, O. Gout, G. de The, F. Sigaux, and J. Peries. 1990. Cell surface phenotype and human T lymphotropic virus type 1 antigen expression in 12 T cell lines derived from peripheral blood and cerebrospinal fluid of West Indian, Guyanese and African patients with tropical spastic paraparesis. J. Gen. Virol. 71:333–341.
- 109. Gibellini, D., A. Caputo, C. Celeghini, A. Bassini, M. La Placa, S. Capitani, and G. Zauli. 1995. Tat-expressing Jurkat cells show an increased resistance to different apoptotic stimuli, including acute human immunodeficiency virus type-1 (HIV-1) infection. Br. J. Haematol. 89:24–33.
- 110. Gimmi, C. D., G. J. Freeman, J. G. Gribben, G. Gray, and L. M. Nadler. 1993. Human T-cell clonal anergy is induced by antigen presentation in the absence of B7 costimulation. Proc. Natl. Acad. Sci. USA 90:6586–6590.
- 111. Gitlin, S. D., J. Dittmer, R. C. Shin, and J. N. Brady. 1993. Transcriptional activation of the human T-lymphotropic virus type I long terminal repeat by functional interaction of Tax1 and Ets1. J. Virol. 67:7307–7316.
- 112. Giulian, D., E. Wendt, K. Vaca, and C. A. Noonan. 1993. The envelope glycoprotein of human immunodeficiency virus type 1 stimulates release of neurotoxins from monocytes. Proc. Natl. Acad. Sci. USA 90:2769–2773.
- 13. Godfried, M. H., T. van der Poll, J. Jansen, J. A. Romijn, J. K. M. Eeftinck-Schattenkerk, E. Endert, S. J. H. van Deventer, and H. P. Sauerwein. 1993. Soluble receptors for tumor necrosis factor: a putative marker of disease progression in HIV infection. AIDS 7:33–43.
- 114. Gootenberg, J. E., F. W. Ruscetti, J. M. Mier, A. Gazdar, and R. C. Gallo. 1981. Human cutaneous T cell lymphoma and leukemia cell lines produce and respond to T cell growth factor. J. Exp. Med. 154:1403–1418.

- Gougeon, M.-L., S. Garcia, J. Heeney, R. Tschopp, H. Lecoeur, D. Guetard, V. Rame, C. Dauguet, and L. Montagnier. 1993. Programmed cell death in AIDS-related HIV and SIV infections. AIDS Res. Hum. Retroviruses 9:553–563.
- 116. Gougeon, M.-L., H. Lecoeur, A. Dulioust, M.-G. Enouf, M. Crouvoisier, C. Goujard, T. Debord, and L. Montagnier. 1996. Programmed cell death in peripheral lymphocytes from HIV-infected persons: increased susceptibility to apoptosis of CD4 and CD8 T cells correlates with lymphocyte activation and with disease progression. J. Immunol. 156:3509–3521.
- 117. Gowda, S. D., B. S. Stein, N. Mohagheghpour, C. J. Benike, and E. G. Engleman. 1989. Evidence that T cell activation is required for HIV-1 entry in CD4+ lymphocytes. J. Immunol. 142:773–780.
- 118. Graziosi, C., G. Pantaleo, K. R. Gantt, J.-P. Fortin, J. F. Demarest, O. J. Cohen, R. P. Sekaly, and A. S. Fauci. 1994. Lack of evidence for the dichotomy of T_H1 and T_H2 predominance in HIV-infected individuals. Science 265:248–252.
- 119. Green, J., C. Begley, D. Wagner, T. Waldmann, and G. Jay. 1989. transactivation of granulocyte-macrophage colony-stimulating factor and the interleukin-2 receptor in transgenic mice carrying the human T-lymphotropic virus type 1 tax gene. Mol. Cell. Biol. 9:4731–4737.
- 120. Gribben, J. G., G. J. Freeman, V. A. Boussiotis, P. Rennert, C. L. Jellis, E. Greenfield, M. Barber, V. A. Restivo, Jr., X. Ke, G. S. Gray, and L. M. Nadler. 1995. CTLA4 mediates antigen-specific apoptosis of human cells. Proc. Natl. Acad. Sci. USA 92:811–815.
- 121. Griffin, D. E., J. C. McArthur, and D. R. Cornblath. 1991. Neopterin and interferon-gamma in serum and cerebrospinal fluid of patients with HIVassociated neurologic disease. Neurology 41:69–74.
- 122. Groux, H., G. Torpier, D. Monte, Y. Mouton, A. Capron, and J. C. Amieson. 1992. Activation-induced death by apoptosis from human immunodeficiency virus-infected asymptomatic individuals. J. Exp. Med. 175:331–338.
- 123. Gruters, R. A., S. A. Otto, B. J. M. Al, A. J. Verhoeven, C. L. Verweij, R. A. W. Van Lier, and F. Miedema. 1991. Non-mitogenic T-cell activation signals are sufficient for induction of human immunodeficiency virus transcription. Eur. J. Immunol 21:167–172.
- 124. Gruters, R. A., F. G. Terpstra, R. De Jong, C. J. M. Van Noesel, R. A. W. Van Lier, and F. Miedema. 1990. Selective loss of T cell functions in different stages of HIV infection. Early loss of anti-CD3-induced proliferation followed by decreased anti-CD3-induced cytotoxic T lymphocyte generation in AIDS-related complex and AIDS. Eur. J. Immunol. 20:1039–1044
- 125. Gupta, P., L. Kinsley, J. Armstrong, M. Ding, M. Cottrill, and C. Rinaldo. 1993. Enhanced expression of human immunodeficiency virus type 1 correlates with development of AIDS. Virology 196:586–595.
- 126. Gupta, S. 1986. Study of activated T cells in man. II. Interleukin 2 receptor and transferrin receptor expression on T cells and production of interleukin 2 in patients with acquired immune deficiency syndrome (AIDS) and AIDS-related complex. Clin. Immunol. Immunopathol. 38:93–100.
- 127. Gutheil, W. G., M. Subramanyam, G. R. Flentke, D. G. Sanford, E. Munoz, B. T. Huber, and W. W. Bachovchin. 1994. Human immunodeficiency virus 1 Tat binds to dipeptidyl aminopeptidase IV (CD26): A possible mechanism for Tat's immunosuppressive activity. Proc. Natl. Acad. Sci. USA 91:6594–6598.
- 128. Guyot, D. J., O. J. Trask, J. M. Andrews, G. C. Newbound, and M. D. Lairmore. 1996. Stimulation of the CD2 receptor pathway induces apoptosis in human T lymphotropic virus type I-infected cell lines. J. Acquired Immune Defic. Syndr. 11:317–325.
- 129. Haanen, J. B. A. G., R. de Waal Malefit, P. C. M. Res, R. R. P. de Vries, and H. Spits. 1991. Selection of a human T helper type 1-like T cell subset by mycobacteria. J. Exp. Med. 174:583–592.
- Harada, S., Y. Koyanagi, H. Nakashima, N. Kobayashi, and N. Yamamoto. 1986. Tumor promoter, TPA, enhances replication of HTLV-III/LAV. Virology 154:249–258.
- Haraguchi, S., R. A. Good, and N. K. Day. 1995. Immunosuppressive retroviral peptides: cAMP and cytokine patterns. Immunol. Today 16:595–603.
- 132. Harding, F. A., J. G. McArthur, J. A. Gross, D. H. Raulet, and J. P. Allison. 1992. CD28-mediated signalling co-stimulates murine T cells and prevents induction of anergy in T-cell clones. Nature (London) 356:607–609.
- 133. Harper, K., C. Balzano, E. Rouvier, M. G. Mattei, M. F. Luciani, and P. Goldstein. 1991. CTLA-4 and CD28 activated lymphocyte molecules are closely related in both mouse and human as to sequence, message expression, gene structure, and chromosomal location. J. Immunol. 147:1037–1044.
- 134. Harrich, D., J. Garcia, F. Wu, R. Mitsuyasu, J. Gonzalez, and R. Gaynor. 1989. Role of Sp1 binding domains in in vivo transcriptional regulation of the human immunodeficiency virus 1 long terminal repeat. J. Virol. 63: 2585_2501
- Hathcock, K. S., G. Laszlo, H. B. Dickler, J. Bradshaw, P. Linsley, and R. J. Hodes. 1993. Identification of an alternative CTLA-4 ligand costimulatory for T cell activation. Science 262:905–907.
- Hattori, T., T. Uchiyama, T. Toibana, K. Takatsuki, and H. Uchino. 1981.
 Surface phenotype of Japanese adult T-cell leukemia cells characterized by monoclonal antibodies. Blood 58:645–647.

- 137. Heeney, J., R. Jonker, W. Koornstra, R. Dubbes, H. Niphuis, A.-M. Di Rienzo, M.-L. Gougeon, and L. Montagnier. 1993. The resistance of HIV-infected chimpanzees to progression to AIDS correlates with absence of HIV-related T-cell dysfunction. J. Med. Primatol. 22:194–200.
- Heeney, J. L. 1995. AIDS: a disease of impaired Th-cell renewal? Immunol. Today 16:515–519.
- 139. Henderson, S., M. Rowe, C. Gregory, D. Croom-Carter, F. Wang, R. Long-necker, E. Kieff, and A. Rickinson. 1991. Induction of bcl-2 expression by Epstein-Barr virus latent membrane protein 1 protects infected B cells from programmed cell death. Cell 65:1107–1115.
- 140. Hinuma, Y., H. Komoda, and T. Chosa. 1982. Antibodies to adult T-cell leukemia-virus-associated antigen (ATLA) in sera from patients with ATL and controls in Japan: a nation-wide sero-epidemiologic study. Int. J. Cancer 29:631–635.
- 141. Hinuma, Y., K. Nagata, M. Hanaoka, M. Nakai, T. Matsumoto, K. Kinoshita, S. Shirakawa, and I. Miyoshi. 1981. Adult T-cell leukemia: antigen in an ATL cell line and detection of antibodies to the antigen in human sera. Proc. Natl. Acad. Sci. USA 78:6476–6480.
- 142. Hirai, H., J. Fujisawa, T. Suzuki, K. Ueda, M. Muramatsu, A. Tsuboi, N. Arai, and M. Yoshida. 1992. Transcriptional activator Tax of HTLV-1 binds to the NFkappa B presursor p105. Oncogene 7:1737–1742.
- 143. Hirai, H., T. Suzuki, J.-I. Fujisawa, J.-I. Inoue, and M. Yoshida. 1994. Tax protein of human T-cell leukemia virus type I binds to the ankyrin motifs of inhibitory factor κB and induces nuclear translocation of transcription factor NF-κB proteins for transcriptional activation. Proc. Natl. Acad. Sci. USA 91:3584–3588.
- 144. Ho, D. D., A. U. Neumann, A. S. Perelson, W. Chen, J. M. Leonard, and M. Markowitz. 1995. Rapid turnover of plasma virions and CD4 lymphocytes in HIV-1 infection. Nature (London) 373:123–126.
- 145. Hollsberg, P., L. J. Ausubel, and D. A. Hafler. 1994. Human T cell lymphotropic virus type I-induced T cell activation. Resistance to TGF-β1-induced suppression. J. Immunol. 153:566–573.
- 146. Hollsberg, P., K. W. Wucherpfennig, L. J. Ausubel, V. Calvo, B. E. Bierer, and D. A. Hafler. 1992. Characterization of HTLV-1 in vivo infected T cell clones. J. Immunol. 148:3256–3263.
- 147. Holmgren, A. 1985. Thioredoxin. Annu. Rev. Biochem. 54:237-271
- 148. Honda, M., K. Kitamura, K. Matsuda, Y. Yokota, N. Yamamoto, R. Mitsuyasu, J.-C. Cherman, and T. Tokunaga. 1989. Soluble IL-2 receptor in AIDS: correlation of its serum level with the classification of HIV-induced diseases and its characterization. J. Immunol. 142:4248–4255.
- 149. Hoshino, H., M. Shimoyama, M. Miwa, and T. Sugimura. 1983. Detection of lymphocytes producing a human retrovirus associated with adult T-cell leukemia by syncytia induction assay. Proc. Natl. Acad. Sci. USA 80:7337– 7341.
- 150. Hoyos, B., D. W. Ballard, E. Bohnlein, M. Siekevitz, and W. C. Greene. 1989. Kappa B specific DNA binding proteins: role in the activation of the interleukin-2 gene. Science 244:457–460.
- 151. Imberti, L., A. Sottini, A. Bettinardi, M. Puoti, and D. Primi. 1991. Selective depletion in HIV infection of T cells that bear specific T cell receptor V_β sequences. Science 254:860–862.
- 152. Inatsuki, A., M. Yasukawa, and Y. Kobayashi. 1989. Functional alteration of herpes simplex virus-specific CD4+ multifunctional T cell clones following infection with human T lymphotropic virus type I. J. Immunol. 143: 1327–1333.
- 153. Israel, N., U. Hazan, J. Alcami, A. Munier, F. Arenzana-Seisdedos, F. Bachelerie, A. Israel, and J.-L. Virelizier. 1989. Tumor necrosis factor stimulates transcription of HIV-1 in human T lymphocytes, independently and synergistically with mitogens. J. Immunol. 143:3956–3960.
- 154. Iwakura, Y., M. Tosu, E. Yoshida, M. Takiguchi, K. Sato, I. Kitajima, K. Nishioka, K. Yamamoto, T. Takeda, M. Hatanaka, H. Yamamoto, and T. Sekiguchi. 1991. Induction of inflammatory arthropathy resembling arthritis in mice transgenic for HTLV-I. Science 253:1026–1028.
- 155. Jacobson, S., H. Shida, D. E. McFarlin, A. S. Fauci, and S. Koenig. 1990. Circulating CD8⁺ cytotoxic T lymphocytes specific for HTLV-I pX in patients with HTLV-I associated neurological disease. Nature (London) 348:245–248.
- 156. Janossy, G., N. Borhtwick, R. Lomnitzer, E. Median, S. Bertel Squire, A. N. Phillips, M. Lipman, M. A. Johnson, C. Lee, and M. Bofill. 1993. Lymphocyte activation in HIV-1 infection. I. Predominant proliferative defects among CD45RO+ cells of the CD4 and CD8 lineages. AIDS 7:613–624.
- 157. Jassoy, C., R. P. Johnson, B. A. Navia, J. Worth, and B. D. Walker. 1992. Detection of a vigorous HIV-1-specific cytotoxic T lymphocyte response in cerebrospinal fluid from infected persons with AIDS dementia complex. J. Immunol. 149:3113–3119.
- 158. Jeang, K. T., S. Wilden, O. Semmes, and S. Wilson. 1990. HTLV-1 transactivator protein, tax, is a trans-repressor of the human β-polymerase gene. Science 247:1082–1084.
- 159. Jenkins, M. K., P. S. Taylor, S. D. Norton, and K. Urdahl. 1991. CD28 delivers a costimulatory signal involved in antigen-specific IL-2 production by human T lymphocytes. J. Immunol. 147:2461–2466.
- 160. Jones, K. A., J. T. Kadonaga, P. A. Luciw, and R. Tjian. 1986. Activation of

- the AIDS retrovirus by the cellular transcription factor Sp1. Science 232: 755–759.
- Jones, K. A., P. A. Luciw, and N. Duchange. 1988. Structural arrangements within the 5'-untranslated leader regions of the HIV-1 and HIV-2 promoters. Genes Dev. 2:1101–1114.
- 162. Kadison, P., H. T. Poteat, K. M. Klein, and D. V. Faller. 1990. Role of protein kinase A in tax transactivation of the human T-cell leukemia virus type 1 long terminal repeat. J. Virol. 64:2141–2148.
- 163. Kalinkovich, A., H. Engelmann, N. Harpaz, R. Burnstein, D. Wallach, and Z. Bentwich. 1992. Elevated serum levels of soluble tumor necrosis factor receptors (sTNF-R) in patients with HIV infection. Clin. Exp. Immunol. 80:351–355
- 164. Kalinkovich, A., G. Livshits, H. Engelmann, N. Harpaz, R. Burnstein, M. Kaminsky, D. Wallach, and Z. Bentwich. 1993. Soluble tumour necrosis factor receptors (sTNF-R) and HIV infection: correlation to CD8⁺ lymphocytes. Clin. Exp. Immunol. 93:350–355.
- 165. Kamine, J., T. Subramanian, and G. Chinnadurai. 1991. Sp1-dependent activation of a synthetic promoter by human immunodeficiency virus type 1 Tat protein. Proc. Natl. Acad. Sci. USA 88:8510–8514.
- 166. Kamine, J., T. Subramanian, and G. Chinnadurai. 1993. Activation of a heterologous promoter by human immunodeficiency virus type 1 Tat requires Sp1 and is distinct from the mode of activation by acidic transcriptional activators. J. Virol. 67:6828–6834.
- 167. Kannagi, M., H. Shida, H. Igarashi, K. Kuruma, H. Murai, Y. Aono, I. Maruyama, M. Osame, T. Hattori, H. Inoko, and S. Harada. 1992. Target epitope in the tax protein of human T-cell leukemia virus type I recognized by class I major histocompatibility complex-restricted cytotoxic T cells. J. Virol. 66:2928–2933.
- 168. Katsikis, P. D., E. S. Wunderlich, C. A. Smith, L. A. Herzenberg, and L. A. Herzenberg. 1995. Fas antigen stimulation induces marked apoptosis of T lymphocytes in human immunodeficiency virus-infected individuals. J. Exp. Med. 181:2029–2036.
- 169. Kaufman, J. D., G. Valandra, G. Rodriquez, G. Bushar, C. Giri, and M. A. Norcross. 1987. Phorbol ester enhances human immunodeficiency virus-promoted gene expression and acts on a repeated 10-base-pair functional enhancer element. Mol. Cell. Biol. 7:3759–3766.
- 170. Kestens, L., G. Vanham, P. Gigase, G. Young, I. Hannet, F. Vanlangendonck, F. Hulstaert, and B. A. Bach. 1992. Expression of activation antigens, HLA-DR and CD38 on CD8 lymphocytes during HIV-1 infection. AIDS 6:793-797.
- 171. Kestens, L., G. Vanham, C. Vereecken, M. Vandenbruane, G. Vercauteren, R. L. Colebunders, and P. L. Gigase. 1994. Selective increase of activation antigens HLA-DR and CD38 on CD4+CD45RO+ T lymphocytes during HIV-1 infection. Clin. Exp. Immunol. 95:436–441.
- 172. Kimata, J. J., T. J. Palker, and L. Ratner. 1993. The mitogenic activity of human T-cell leukemia virus type 1 is T-cell associated and requires the CD2/LFA-3 activation pathway. J. Virol. 67:3134–3141.
- Kimata, J. J., and L. Ratner. 1991. Temporal regulation of viral and cellular gene expression during human T-lymphotropic virus type I-mediated lymphocyte immortalization. J. Virol. 65:4398–4407.
- 174. Kinoshita, T., M. Shimoyama, K. Tobinai, M. Ito, S.-C. Ito, S. Ikeda, K. Tajima, K. Shimotohno, and T. Sugimura. 1989. Detection of mRNA for the *tax₁/rex₁* gene of human T-cell leukemia virus type 1 in fresh peripheral blood mononuclear cells of adult T-cell leukemia patients and viral carriers by using the polymerase chain reaction. Proc. Natl. Acad. Sci. USA 86: 5620–5624.
- 175. Kinter, A. L., G. Poli, W. Maury, T. M. Folks, and A. S. Fauci. 1990. Direct and cytokine-mediated activation of protein kinase C induces human immunodeficiency virus expression in chronically infected promonocytic cells. J. Virol. 64:4306–4312.
- 176. Kira, J.-I., Y. Itoyama, Y. Koyanagi, J. Tateishi, M. Kishikawa, S.-I. Akizuki, I. Kobayashi, N. Toki, K. Sueishi, and H. Sato. 1992. Presence of HTLV-1 proviral DNA in central nervous system of patients with HTLV-1-associated myelopathy. Ann. Neurol. 31:39.
- 177. Klas, C., K.-M. Debatin, R. R. Jonker, and P. H. Krammer. 1993. Activation interferes with the APO-1 pathway in mature human T cells. Int. Immunol. 5:625–630.
- 178. Knuchel, M., D. P. Bednarik, N. Chikkalaand, and A. A. Ansari. 1994. Biphasic in vitro regulation of retroviral replication by CD8⁺ cells from nonhuman primates. J. Acquired Immune Defic. Syndr. 7:438–446.
- 179. Koenig, S., P. Earl, D. Powell, G. Pantaleo, S. Merli, B. Moss, and A. S. Fauci. 1988. Group-specific, major histocompatibility complex class-1 restricted cytotoxic responses to human immunodeficiency virus-1 (HIV-1) envelope proteins by cloned peripheral blood T cells from an HIV-infected individual. Proc. Natl. Acad. Sci. USA 85:8638–8642.
- 180. Koenig, S., H. E. Gendelman, J. M. Orenstein, M. C. Dal Canto, G. H. Pezeshkpour, M. Yungbluth, F. Janotta, A. Aksamit, M. A. Martin, and A. S. Fauci. 1986. Detection of AIDS virus in macrophages in brain tissue from AIDS patients with encephalopathy. Science 233:1089–1093.
- 181. Koenig, S., R. M. Woods, Y. A. Brewah, A. J. Newell, G. M. Jones, E. Boone, J. W. Adelsberger, M. W. Baseler, S. M. Robinson, and S. Jacobson. 1993. Characterisation of MHC class 1 restricted cytotoxic T-cell responses to Tax

- in HTLV-1 infected patients with neurologic disease. J. Immunol. 156: 3874-3883.
- 182. Koulova, L., E. A. Clark, G. Shu, and B. Dupont. 1991. The CD28 ligand B7/BB1 provides a costimulatory signal for alloactivation of CD4⁺ T cells. J. Exp. Med. 173:759–762.
- 183. Koup, R. A., J. T. Safrti, Y. Cao, C. A. Andrews, G. McLeod, W. Borkowsky, C. Farthing, and D. D. Ho. 1994. Temporal association of cellular immune responses with the initial control of viremia in primary human immunodeficiency virus type 1 syndrome. J. Virol. 68:4650–4655.
- 184. Koyanagi, Y., W. A. O'Brien, J. Q. Zhao, D. W. Golde, J. C. Gasson, and I. S. Y. Chen. 1988. Cytokines alter production of HIV-1 from primary mononuclear phagocytes. Science 241:1673–1675.
- 185. Krummel, M. F., and J. P. Allison. 1995. CD28 and CTLA-4 have opposing effects on the response of T cells to stimulation. J. Exp. Med. 182:459–465.
- 186. Lacoste, J., L. Petropoulos, N. Pepin, and J. Hiscott. 1995. Constitutive phosphorylation and turnover of IκBα in human T-cell leukemia virus type I-infected and Tax-expressing cells. J. Virol. 69:564–569.
- 187. Lacy, J. N., M. A. Forbes, M. A. Waugh, E. H. Cooper, and M. H. Hambling. 1987. Serum β₂-microglobulin and human immunodeficiency virus infection. AIDS 1:123–127.
- 188. Lal, R. B., and D. L. Rudolph. 1991. Constitutive production of interleukin-6 and tumor necrosis factor-α from spontaneously proliferating T cells in patients with human T-cell lymphotropic virus type-I/II. Blood 78:571– 574.
- 189. Landay, A. L., C. E. Mackewicz, and J. A. Levy. 1993. An activated CD8+ T cell phenotype correlates with anti-HIV activity and asymptomatic clinical status. Clin. Immunol. Immunopathol. 69:106–116.
- 190. Lane, H. C., J. M. Depper, W. C. Greene, G. Whalen, T. A. Waldman, and A. S. Fauci. 1985. Quantitative analysis of immune function in patients with AIDS: evidence for a selective defect in soluble antigen recognition. N. Engl. J. Med. 313:79–84.
- Lane, H. C., H. Masur, L. C. Edgar, G. Whalen, A. H. Rook, and A. S. Fauci. 1983. Abnormalities of B cell activation and immunoregulation in patients with the acquired immunodeficiency syndrome. N. Engl. J. Med. 309:453– 458.
- 192. Lang, W., M. Perkins, R. E. Anderson, R. Royce, N. Jewell, and W. Winkelstein, Jr. 1993. Patterns of T-lymphocyte changes with human immunode-ficiency virus infection: from seroconversion to the development of AIDS. J. Acquired Immune Defic. Syndr. 2:63–69.
- 193. Lanoix, J., J. Lacoste, N. Pepin, N. Rice, and J. Hiscott. 1994. Overproduction of NFκB2 (lyt-10) and c-Rel: a mechanism for HTLV-1 Tax-mediated trans-activation via the NF-κB signalling pathway. Oncogene 9:841–852.
- 194. Lasky, L. A., G. Nakamura, D. H. Smith, C. Fennie, C. Shimasaki, E. Patzner, P. Berman, T. Gregory, and D. J. Capon. 1987. Delineation of a region of the human immunodeficiency virus type 1 gp120 glycoprotein critical for interaction with the CD4 receptor. Cell 50:975–985.
- 195. Lathey, J. L., S. Kanangat, and B. T. Rouse. 1994. Differential expression of tumor necrosis factor α and interleukin 1β compared with interleukin 6 in monocytes from human immunodeficiency virus-positive individuals measured by polymerase chain reaction. J. Acquired Immune Defic. Syndr. 7:109–115.
- 196. Laurent-Crawford, A. G., B. Krust, Y. Riviére, C. Desgranges, S. Muller, M. Kieny, C. Dauguet, and A. G. Hovanessian. 1993. Membrane expression of HIV envelope glycoproteins triggers apoptosis in CD4 cells. AIDS Res. Hum. Retroviruses 9:761–773.
- 197. Lederer, J. A., J. S. Liou, S. Kim, N. Rice, and A. H. Lichtman. 1996. Regulation of NFκB activation in T helper 1 and T helper 2 cells. J. Immunol. 156:56–63.
- Lederer, J. A., J. S. Liou, M. D. Todd, L. H. Glimcher, and A. H. Lichtman.
 1994. Regulation of cytokine gene expression in T helper cell subsets.
 J. Immunol. 152:77–86.
- 199. Lederer, J. A., V. L. Perez, L. DesRoches, S. M. Kim, A. K. Abbas, and A. H. Lichtman. 1996. Cytokine transcriptional events during helper T cell subset differentiation. J. Exp. Med. 184:397–406.
- Lehky, T. J., E. P. Cowan, L. A. Lampson, and S. Jacobson. 1994. Induction of HLA class I and class II expression in human T-lymphotropic virus type I-infected neuroblastoma cells. J. Virol. 68:1854–1863.
- 201. Leno, M., R. M. Simpson, F. S. Bowers, and T. J. Kindt. 1995. Human T lymphocyte virus 1 from a leukemogenic cell line mediates in vivo and in vitro lymphocyte apoptosis. J. Exp. Med. 181:1575–1580.
- Lenschow, D. J., G. Huei-Ting Su, L. A. Zuckerman, N. Nabavi, C. L. Jellis, G. S. Gray, J. Miller, and J. A. Bluestone. 1993. Expression and functional significance of an additional ligand for CTLA-4. Proc. Natl. Acad. Sci. USA 90:11054–11058.
- Levine, B., Q. Huang, J. Y. Isaacs, J. C. Reed, D. E. Griffith, and J. M. Hardwick. 1993. Conversion of lytic to persistent alphavirus infection by the bcl-2 cellular oncogene. Nature (London) 361:739–742.
- Levy, J. A. 1993. Pathogenesis of human immunodeficiency virus infection. Microbiol. Rev. 57:183–289.
- 205. Li, C. J., D. J. Friedman, C. Wang, V. Metelev, and A. B. Pardee. 1995.

- Induction of apoptosis in uninfected lymphocytes by HIV-1 Tat protein. Science **268**:429–431.
- Liegler, T. J., and D. P. Stites. 1994. HIV-1 gp120 and anti-gp120 induce reversible unresponsiveness in peripheral CD4 T lymphocytes. J. Acquired Immune Defic. Syndr. 7:340–348.
- 207. Linsley, P. S., W. Brady, L. S. Grosmaire, N. K. Damle, and J. A. Ledbetter. 1991. CTLA-4 is a second receptor for the B cell activation antigen B7. J. Exp. Med. 174:561–569.
- Linsley, P. S., J. L. Greene, P. Tan, J. Bradshaw, J. A. Ledbetter, C. Anasetti, and N. K. Damle. 1992. Coexpression and functional cooperation of CTLA-4 and CD28 on activated lymphocytes. J. Exp. Med. 176:1595–1604.
- Lipton, S. A. 1993. Human immunodeficiency virus-infected macrophages, gp120, and N-methyl-p-aspartate receptor-mediated neurotoxicity. Ann. Neurol. 33:227–228.
- Liu, J., N. D. Perkins, R. M. Schmid, and G. J. Nabel. 1992. Specific NFκB subunits act in concert with Tat to stimulate human immunodeficiency virus type 1 transcription. J. Virol. 66:3883–3887.
- Ljunggren, K., P.-A. Broliden, L. Morfeldt-Manson, M. Jondal, and B. Wahren. 1988. IgG subclass response to HIV in relation to antibody-dependent cellular cytotoxicity at different clinical stages. Clin. Exp. Immunol. 73:343–347.
- 212. Lu, Y., N. Touzjian, M. Stenzel, T. Dorfman, J. G. Sodroski, and W. A. Haseltine. 1990. Identification of cis-acting repressive sequences within the negative regulatory element of human immunodeficiency virus type 1. J. Virol. 64:5226–5229.
- 213. Lu, Y.-Y., Y. Koga, K. Tanaka, M. Sasaki, G. Kimura, and K. Nomoto. 1994. Apoptosis induced in CD4+ cells expressing gp160 of human immunodeficiency virus type 1. J. Virol. 68:390–399.
- 214. Mabrouk, K., J. Van Rietschoten, E. Vives, H. Darbon, H. Rochat, and J.-M. Sabatier. 1991. Lethal neurotoxicity in mice of the basic domains of HIV and SIV Rev proteins. FEBS Lett. 289:13-17.
- Macatonia, S. E., R. Lau, S. Patterson, A. J. Pinching, and S. C. Knight. 1990. Dendritic cell infection, depletion and dysfunction in HIV-infected individuals. Immunology 71:38–45.
- Mackewicz, C. E., D. J. Blackbourn, and J. A. Levy. 1995. CD8⁺ T cells suppress human immunodeficiency virus replication by inhibiting viral transcription. Proc. Natl. Acad. Sci. USA 92:2308–2312.
- Mackewicz, C. E., and J. A. Levy. 1992. CD8⁺ cell anti-HIV activity: nonlytic suppression of virus replication. AIDS Res. Hum. Retroviruses 6:1039–1050.
- 218. Maggi, E., M. Mazzetti, A. Ravina, F. Annunziato, M. de Carli, M. P. Piccinni, R. Manetti, M. Carbonari, A. M. Pesce, G. Del Prete, and S. Romagnani. 1994. Ability of HIV to promote a $T_{\rm H}1$ to $T_{\rm H}0$ shift and to replicate preferentially in $T_{\rm H}2$ and $T_{\rm H}0$ cells. Science 265:244–248.
- Maguer-Satta, V., L. Gazzolo, and M.Duc Dodon. 1995. Human immature thymocytes as target cells of the leukemogenic activity of human T-cell leukemia virus type I. Blood 86:1444–1452.
- Maldarelli, F., H. Sato, E. Berthold, J. Orenstein, and M. A. Martin. 1995.
 Rapid induction of apoptosis by cell-to-cell transmission of human immunodeficiency virus type 1. J. Virol. 69:6457–6465.
- 221. Marfella, A., V. Ruocco, A. Capobianco, M. Perna, G. Santelli, G. Frigione, S. K. Kyalwazi, R. D. Mugerwa, D. Serwadda, E. Beth-Giraldo, and G. Giraldo. 1989. Neopterin and alpha-interferon in patients affected by Kaposi's sarcoma from Africa. Eur. J. Cancer Clin. Oncol. 25:1145–1150.
- 222. Markovitz, D. M., M. J. Smith, J. Hilfinger, M. C. Hannibal, B. Petryniak, and G. J. Nabel. 1992. Activation of the human immunodeficiency virus type 2 enhancer is dependent on purine box and κB elements. J. Virol. 66:5479–5484.
- 223. Marriott, S. J., I. Boros, J. Duvall, and J. Brady. 1989. Indirect binding of human T-cell leukemia virus type I tax_I to a responsive element in the viral long terminal repeat. Mol. Cell. Biol. 9:4152–4160.
- 224. Maruyama, M., H. Shibuya, H. Harada, M. Hatekayama, M. Seiki, T. Fujita, J. Inoue, M. Yoshida, and T. Taniguchi. 1987. Evidence for aberrant activation of the interleukin-2 autocrine loop by HTLV-1 encoded p40x and T3/Ti complex triggering. Cell 48:343–350.
- Matsuyama, T., N. Kobayashi, and N. Yamamoto. 1991. Cytokines and HIV infection: is AIDS a tumor necrosis factor disease? AIDS 5:1405–1417.
- 226. Meyaard, L., S. Otto, B. Hooibrink, and F. Miedema. 1994. Quantitative analysis of CD4⁺ T cell function in the course of human immunodeficiency virus infection: decline of both naive and memory alloreactive T cells. J. Clin. Invest. 94:1947–1952.
- 227. Meyaard, L., S. A. Otto, R. R. Jonker, M. J. Mijnster, R. P. M. Keet, and F. Miedema. 1992. Programmed cell death of T-cells in HIV-1 infection. Science 257:217-219.
- 228. Meyaard, L., S. A. Otto, I. P. M. Keet, M. T. L. Roos, and F. Miedema. 1994. Programmed death of T cells in human immunodeficiency virus infection: no correlation with progression to disease. J. Clin. Invest. 93:982–988.
- 229. Miedema, F., A. J. C. Petit, F. G. Terpstra, J. K. M. E. Schattenkerk, F. De Wolf, B. J. M. Al, M. Th. L. Roos, J. M. A. Lange, S. A. Danner, J. Goudsmit, and P. T. A. Schellekens. 1988. Immunological abnormalities in human immunodeficiency virus (HIV)-infected asymptomatic homosexual

- men. HIV affects the immune system before CD4⁺ T helper cell depletion occurs. J. Clin. Invest. **82**:1908–1914.
- Mier, J. W., and R. C. Gallo. 1980. Purification and some characteristics of human T-cell growth factor from phytohemagglutinin-stimulated lymphocyte-conditioned media. Proc. Natl. Acad. Sci. USA 77:6134–6138.
- Miyamoto, K., N. Tomita, A. Ishii, H. Nonaka, T. Kondo, T. Tanaka, and K.-I. Kitajima. 1984. Chromosome abnormalities of leukemia cells in adult patients with T-cell leukemia. JNCI 73:353–362.
- Miyawaki, T., T. Uehara, R. Nibu, T. Tsuji, A. Yachie, S. Yonehara, and N. Taniguchi. 1992. Differential expression of apoptosis-related Fas antigen on lymphocyte subpopulations in human peripheral blood. J. Immunol. 149: 3753–3758.
- 233. Morgan, D. A., F. W. Ruscetti, and R. Gallo. 1976. Selective in vitro growth of T lymphocytes from normal human bone marrow. Science 193:1007– 1008
- 234. Morimoto, C., C. I. Lord, C. Zhang, J. S. Duke-Cohan, N. L. Letvin, and S. F. Schlossman. 1994. Role of CD26/dipeptidyl peptidase IV in human immunodeficiency virus type 1 infection and apoptosis. Proc. Natl. Acad. Sci. USA 91:9960–9964.
- Morrow, C. D., J. Park, and J. K. Wakefield. 1994. Viral gene products and replication of the human immunodeficiency type 1 virus. Am. J. Physiol. 266:C1135–C1156.
- Mosmann, T. R., H. Cherwinski, M. W. Bond, M. A. Giedlin, and R. L. Coffman. 1986. Two types of murine helper T cell clones. I. Definition according to profiles of lymphokine activities and secreted proteins. J. Immunol. 136:2348-2357.
- Mukhopadhyaya, R., and M. Reza Sadaie. 1993. Nucleotide sequence analysis of HTLV-I isolated from cerebrospinal fluid of a patient with TSP/HAM: comparison to other HTLV-I isolates. AIDS Res. Hum. Retroviruses 9:109–114.
- 238. Munoz, E., A. M. Zubiaga, M. Merrow, N. P. Sauter, and B. T. Huber. 1990. Cholera toxin discriminates between T helper 1 and 2 cells in T cell receptor-mediated activation: role of cAMP in T cell proliferation. J. Exp. Med. 172:95–103.
- 239. Muro-Cacho, C. A., G. Pantaleo, and A. Fauci. 1995. Analysis of apoptosis in lymph nodes of HIV-infected persons. Intensity of apoptosis correlates with the general state of activation of the lymphoid tissue and not with the stage of disease or viral burden. J. Immunol. 154:5555–5566.
- 240. Murphy, E. L., B. Hanchard, J. P. Figueroa, W. N. Gibbs, W. S. Lofters, M. Campbell, J. J. Goedert, and W. A. Blattner. 1989. Modeling the risk of adult T-cell leukemia/lymphoma in persons infected with human T-lymphotropic virus type I. Int. J. Cancer 43:250–253.
- Nabel, G., and D. Baltimore. 1987. An inducible factor activates expression of human immunodeficiency virus in T cells. Nature (London) 326:711–713.
- 242. Nakamura, M., Y. Itoyama, M. Kuroki, S. Nakano, S. Kondoh, S. Nagafuchi, J. Kira, I. Ichinose, K. Mitsugi, K. Anzai, H. Mori, M. Fukui, S. Okamura, and Y. Niho. 1992. Increase in peripheral B lymphocytes committed to the production of monoreactive and high affinity antibodies to HTLV-1 in patients with HAM/TSP. J. Neuroimmunol. 37:35–45.
- 243. Nakamura, M., M. Niki, K. Ohtani, and K. Sugamura. 1989. Differential activation of the 21-base-pair enhancer element of human T-cell leukemia virus type I by its own trans-activator and cyclic AMP. Nucleic Acids Res. 17:5207–5221.
- 244. Natazuka, T., T. Umemiya-Okada, T. Matsui, T. Saida, and Y. Nakao. 1993. FK506 and cyclosporin A regulate proliferation and proto-oncogene expression in HTLV-1-associated myelopathy/tropical-spastic-paraparesis-derived T cells. Int. J. Cancer 54:348–354.
- 245. Navikas, V., J. Link, B. Wahren, CH. Persson, and H. Link. 1994. Increased levels of interferon-gamma (IFNγ), IL-4 and transforming growth factor-beta (TGF-β) mRNA expressing blood mononuclear cells in human HIV infection. Clin. Exp. Immunol. 96:59–63.
- Nerenberg, M., S. Hinrichs, K. Reynolds, G. Khoury, and G. Jay. 1987. The tat gene of HTLV-1 induces mesenchymal tumors in transgenic mice. Science 237:1324–1329.
- 247. Neumann, M., T. Grieshammer, S. Chuvpilo, B. Kneitz, M. Lohoff, A. Schimpl, B. R. Franza, and E. Serfling. 1995. RelA/p65 is a molecular target for the immunosuppressive action of protein kinase A. EMBO J. 14:1991–2004.
- Newell, M. K., L. J. Haughn, C. R. Maroun, and M. H. Julius. 1990. Death
 of mature T cells by separate ligation of CD4 and the T-cell receptor for
 antigen. Nature (London) 347:286–289.
- 249. Niewiesk, S., S. Daenke, C. E. Parker, G. Taylor, J. Weber, S. Nightingale, and C. R. M. Bangham. 1995. Naturally occurring variants of human T-cell leukemia virus type I Tax protein impair its recognition by cytotoxic T lymphocytes and the transactivation function of Tax. J. Virol. 69:2649–2653.
- Nomoto, M. Y. Utatsu, Y. Soejima, and M. Osame. 1991. Neopterin in cerebrospinal fluid: a useful marker for diagnosis of HTLV-I-associated myelopathy/tropical spastic paraparesis. Neurology 41:457.
- 251. Noronha, I. L., V. Daniel, K. Schimpf, and G. Opelz. 1992. Soluble IL-2 receptor and tumour necrosis factor-α in plasma of haemophilia patients infected with HIV. Clin. Exp. Immunol. 87:287–292.
- 252. Osame, M., K. Usuku, S. Izumo, N. Ijichi, H. Amitini, A. Igata, M. Matsu-

- moto, and M. Tara. 1986. HTLV-1 associated myelopathy, a new clinical entity. Lancet i:1031–1032.
- Osmond, D. H., S. Shiboski, P. Bacchetti, E. E. Winger, and A. R. Moss. 1991. Immune activation markers and AIDS prognosis. AIDS 5:505–511.
- 254. Oyaizu, N., N. Chirmule, V. S. Kalyanaraman, W. W. Hall, R. A. Good, and S. Pahwa. 1990. Human immunodeficiency virus type 1 envelope glycoprotein gp120 produces immune defects in CD4+ T lymphocytes by inhibiting interleukin 2 mRNA. Proc. Natl. Acad. Sci. USA 87:2379–2383.
- 255. Oyaizu, N., T. W. McCloskey, M. Coronesi, N. Chirmule, V. S. Kalyanaraman, and S. Pawha. 1993. Accelerated apoptosis in peripheral blood mononuclear cells (PBMCs) from human immunodeficiency virus type-1 infected patients and in CD4 cross-linked PBMCs from normal individuals. Blood 82:3392–3400.
- Oyaizu, N., and S. Pahwa. 1995. Role of apoptosis in HIV disease pathogenesis. J. Clin. Immunol. 15:217–231.
- Pantaleo, G., and A. S. Fauci. 1995. New concepts in the immunopathogenesis of HIV infection. Annu. Rev. Immunol. 13:487–512.
- 258. Pantaleo, G., S. Menzo, M. Vaccarezza, C. Graziosi, O. J. Cohen, J. F. Demarest, D. Montegiori, J. M. Orenstein, C. Fox, L. K. Schrager, J. B. Margolick, S. Buchbinder, J. V. Giorgi, and A. S. Fauci. 1995. Studies in subjects with long-term nonprogressive human immunodeficiency virus infection. N. Engl. J. Med. 332:209-216.
- Parker, C. E., S. Daenke, S. Nightingale, and C. R. M. Bangham. 1992. Activated, HTLV-1-specific cytotoxic T-lymphocytes are found in healthy seropositives as well as patients with tropical spastic paraparesis. Virology 188:628-636.
- 260. Parker, C. E., S. Nightingale, G. P. Taylor, J. Weber, and C. R. M. Bangham. 1994. Circulating anti-Tax cytotoxic T lymphocytes from human T-cell leukemia virus type I-infected people, with and without tropical spastic paraparesis, recognize multiple epitopes simultaneously. J. Virol. 68:2860–2868
- 261. Parronchi, P., D. Macchia, M.-P. Piccinni, P. Biswas, C. Simonelli, E. Maggi, M. Ricci, A. A. Ansari, and S. Romagnani. 1991. Allergen- and bacterial antigen-specific T-cell clones established from atopic donors show a different profile of cytokine production. Proc. Natl. Acad. Sci. USA 88:4538-43.
- Perini, G., S. Wagner, and M. R. Green. 1995. Recognition of bZIP proteins by the human T-cell leukaemia virus transactivator Tax. Nature (London) 376:602–605.
- Pinching, A. J. 1988. Factors affecting the natural history of human immunodeficiency virus infection. Immunol. Rev. 1:23–38.
- 264. Pinching, A. J., and K. E. Nye. 1990. Defective signal transduction—a common pathway for cellular dysfunction in HIV infection? Immunol. Today 11:256–259.
- Pinto, L., M. J. Covas, and R. M. M. Victorino. 1991. Loss of CD45RA and gain of CD45RO after *in vitro* activation of lymphocytes from HIV-infected patients. Immunology 73:147–150.
- 266. Pizzolo, G., F. Vinante, L. Morosato, G. Nadali, M. Chilosi, G. Gandini, A. Sinicco, R. Raiteri, G. Semenzato, H. Stein, and G. Perona. 1994. High serum level of the soluble form of CD30 molecule in the early phase of HIV-1 infection as an independent predictor of progression to AIDS. AIDS 8:741–745.
- 267. Planelles, V., J. B. M. Jowett, Q.-X. Li, Y. Xie, B. Hahn, and I. S. Y. Chen. 1996. Vpr-induced cell cycle arrest is conserved among primate lentiviruses. J. Virol. 70:2516–2524.
- 268. Poiesz, B.J., F. W. Ruscetti, A. F. Gazdar, P. A. Bunn, J. D. Minna, and R. C. Gallo. 1980. Detection and isolation of type C retrovirus particles from fresh and cultured lymphocytes of a patient with cutaneous T-cell lymphoma. Proc. Natl. Acad. Sci. USA 77:7415–7419.
- 269. Poli, G., P. Bressler, A. Kinter, E. Duh, W. L. Timmer, A. Rabson, J. S. Justement, S. Stanley, and A. S. Fauci. 1990. IL-6 induces human immunodeficiency virus expression in infected monocytic cells alone and in synergy with TNF by transcriptional and post-transcriptional mechanisms. J. Exp. Med. 172:151–158.
- Poli, G., and A. S. Fauci. 1992. The effect of cytokines and pharmacologic agents on chronic HIV infection. AIDS Res. Hum. Retroviruses 8:191–197.
- 271. Popovic, M., N. Flomenberg, D. J. Volkman, D. Mann, A. S. Fauci, B. Dupont, and R. C. Gallo. 1984. Alteration of T-cell functions by infection with HTLV-I or HTLV-II. Science 226:459–462.
- 272. Popovic, M., G. Lange-Wantzin, P. S. Sarin, D. Mann, and R. C. Gallo. 1983. Transformation of human umbilical cord blood T cells by human T cell leukemia/lymphoma virus. Proc. Natl. Acad. Sci. USA 80:5402–5406.
- 273. Poteat, H., P. Kadison, K. McGuire, L. Park, R. E. Park, J. G. Sodroski, and W. A. Haseltine. 1989. Response of the human T-cell leukemia virus type I long terminal repeat to cyclic AMP. J. Virol. 63:1604–1611.
- 274. Powrie, F., and R. L. Coffman. 1993. Cytokine regulation of T-cell function: potential for therapeutic intervention. Immunol. Today 14:270–274.
- 275. Pozzati, R., J. Vogel, and G. Jay. 1990. The human T-lymphotropic virus type I tax gene can cooperate with the ras oncogene to induce neoplastic transformation of cells. Mol. Cell. Biol. 10:413–417.
- 276. Prasad, K. V., R. Kapeller, O. Janssen, H. Repke, J. S. Ducke-Cohan, L. C. Cantley, and C. E. Rudd. 1993. Phosphatidylinositol (PI) 3-kinase and PI

- 4-kinase binding to the CD4-p56^{lck} complex: the p56^{lck} SH3 domain binds to PI 3-kinase but not PI 4-kinase. Mol. Cell. Biol. **13:**7708–7717.
- 277. Procaccia, S., A. Lazzarin, A. Colucci, A. Gasparini, P. Forcellini, D. Lanzanova, C. U. Foppa, R. Novati, and C. Zanussi. 1987. IgM, IgG and IgA rheumatoid factors and circulating immune complexes in patients with AIDS and AIDS-related complex with serological abnormalities. Clin. Exp. Immunol. 67:236–244.
- 278. Radonovich, M., and K.-T. Jeang. 1989. Activation of the human T-cell leukemia virus type I long terminal repeat by 12-O-tetradecanoylphorbol-13-acetate and by tax (p40^x) occurs through similar but functionally distinct target sequences. J. Virol. 63:2987–2994.
- 279. Ramsdell, F., M. S. Seaman, R. E. Miller, K. S. Picha, M. K. Kennedy, and D. H. Lynch. 1994. Differential ability of Th1 and Th2 T cells to express Fas ligand and to undergo activation-induced cell death. Int. Immunol. 6:1545– 1552
- 280. Rao, L., M. Debbas, P. Sabbatini, D. Hockenbery, S. Korsmeyer, and E. White. 1992. The adenovirus E1A proteins induce apoptosis, which is inhibited by the R1B 19-kDa and Bcl-2 proteins. Proc. Natl. Acad. Sci. USA 89:7742–7746.
- 281. Re, M. C., G. Zauli, D. Gibellini, G. Furlini, E. Ramazzotti, P. Monari, S. Ranieri, S. Capitani, and M. La Placa. 1993. Uninfected haematopoietic progenitor (CD34+) cells purified from the bone marrow of AIDS patients are committed to apoptotic cell death in culture. AIDS 7:1049–1055.
- 282. Rinaldo, C., X.-L. Huang, Z. Fan, M. Ding, L. Beltz, A. Logar, D. Panicali, G. Mazzara, J. Liebmann, M. Cottrill, and P. Gupta. 1995. High levels of anti-human immunodeficiency virus type 1 (HIV-1) memory cytotoxic Tlymphocyte activation and low viral load are associated with lack of disease in HIV-1-infected long-term nonprogressors. J. Virol. 69:5838–5842.
- Riviere, Y. 1994. Virus-specific cytotoxic T lymphocyte responses in patients infected with human immunodeficiency virus. Cell. Mol. Biol. 40(Suppl. 1):45–48.
- Romagnani, S. 1992. Human TH1 and TH2 subsets: regulation of differentiation and role in protection and immunopathology. Int. Arch. Allergy Immunol. 98:279–285.
- 285. Romagnani, S. 1994. Lymphokine production by human T cells in disease states. Annu. Rev. Immunol. 12:227–257.
- 286. Romagnani, S., F. Annunziato, R. Manetti, F. Almerigogna, R. Biagiotti, M. G. Giudizi, A. Ravina, V. Gianno, L. Tomaevic, and E. Maggi. 1996. Role for CD30 in HIV expression. Immunol. Lett. 51:83–88.
- 287. Romagnani, S., G. Del Prete, R. Manetti, A. Ravina, F. Annunziato, M. de Carli, M. Mazzetti, M.-P. Piccinni, M. M. D'Elios, P. Parronchi, S. Sampognaro, and E. Maggi. 1994. Role of T_H1/T_H2 cytokines in HIV infection. Immunol. Rev. 140:73–92.
- Rosenberg, Z., and A. Fauci. 1990. Immunopathogenic mechanisms of HIV infection: cytokine induction of HIV expression. Immunol. Today 11:176–180
- 289. Sandstrom, P. A., D. Pardi, C. S. Goldsmith, D. Chengying, A. M. Diamond, and T. M. Folks. 1996. bcl-2 expression facilitates human immunodeficiency virus type 1-mediated cytopathic effects during acute spreading infections. J. Virol. 70:4617–4622.
- 290. Santelli, G., G. Melillo, A. Marfella, M. Napolitano, V. D'Alessio, R. A. Satriano, E. Beth-Giraldo, G. Giraldo, M. Perna, and G. Castello. 1988. Urinary neopterin and immunological features in patients with Kaposi's sarcoma. Eur. J. Cancer Clin. Oncol. 24:1391–1396.
- 291. Schlesinger, M., O. Afik, O. Loves, S. Maayan, and R. Rabinowitz. 1990. Changes in the level of soluble CD2 molecules in the serum of HIV-infected individuals. Cancer Detect. Prev. 14:347–351.
- 292. Schlesinger, M., F. N. Chiu, M. Badamchian, J. D. Jiang, J. P. Roboz, A. L. Goldstein, and J. G. Bekesi. 1994. A distinctive form of soluble CD8 is secreted by stimulated CD8⁺ cells in HIV-1-infected and high-risk individuals. Clin. Immunol. Immunopathol. 73:252–260.
- 293. Schmidt, A., L. Hennighausen, and U. Siebenlist. 1990. Inducible nuclear factor binding to the κB elements of the human immunodeficiency virus enhancer in T cells can be blocked by cyclosporin A in a signal-dependent manner. J. Virol. 64:4037–4041.
- 294. Schols, D., and E. Declercq. 1996. Human immunodeficiency virus type 1 gp120 induces anergy in human peripheral blood lymphocytes by inducing interleukin-10 production. J. Virol. 70:4953–4960.
- 295. Schreck, R., R. Grassman, B. Fleckenstein, and P. A. Baeuerle. 1992. Antioxidants selectively suppress activation of NFκB by human T-cell leukemia virus type I tax protein. J. Virol. 66:6288–6293.
- 296. Scott-Algara, D., F. Vullier, M. Marasescu, J. de Saint-Martin, and G. Didhiero. 1991. Serum levels of IL-2, IL-1α, TNFα and soluble receptor of IL-2 in HIV-infected patients. AIDS Res. Hum. Retroviruses 7:381–386.
- 297. Seder, R. A., and W. E. Paul. 1994. Acquisition of lymphokine-producing phenotype by CD4⁺ T cells. Annu. Rev. Immunol. 12:635–673.
- 298. Seder, R. A., W. E. Paul, M. M. Davis, D. S. Fazekas, and B. Groth. 1992. The presence of interleukin 4 during in vitro priming determines the lymphokine producing potential of CD4⁺ T cells from T cell receptor transgenic mice. J. Exp. Med. 176:1091–1098.
- 299. Seligman, M., L. Chess, J. L. Fahey, A. S. Fauci, P. J. Lachman, J. L'Age-Ster, J. Ngu, A. J. Pinching, F. S. Rosen, T. J. Spira, and J. W. Wybran.

- 1984. AIDS—an immunologic reevaluation. N. Engl. J. Med. 311:1286–1292.
- 300. Seligman, M., A. J. Pinching, F. S. Rosen, J. L. Fahey, R. M. Khaitov, D. Klatzmann, S. Koenig, N. Luo, J. Ngu, G. Reithmuller, and T. J. Spira. 1987. Immunology of human immunodeficiency virus infection and the acquired immunodeficiency syndrome. An update. Ann. Intern. Med. 107: 234, 242.
- 301. Sellins, K. S., and J. J. Cohen. 1987. Gene induction by γ -irradiation leads to DNA fragmentation in lymphocytes. J. Immunol. 139:3199–3206.
- 302. Selmaj, K. W., M. Farooq, W. T. Norton, C. S. Raine, and C. F. Brosnan. 1990. Proliferation of astrocytes in vitro in response to cytokines. A primary role for tumor necrosis factor. J. Immunol. 144:129–135.
- 303. Sethi, K. K., and H. Naher. 1986. Elevated titers of cell-free interleukin-2 receptor in serum and cerebrospinal fluid specimens of patients with acquired immunodeficiency syndrome. Immunol. Lett. 13:179–184.
- 304. Shaw, J.-P., P. J. Ut, D. B. Durand, J. J. Toole, E. A. Emanuel, and G. Crabtree. 1988. Identification of a putative regulator of early T-cell activation genes. Science 241:202–205.
- 305. Shearer, G. M., D. C. Bernstein, K. S. K. Tung, C. S. Via, R. Redfield, S. Z. Salahuddin, and R. C. Gallo. 1986. A model for the selective loss of MHC self-restricted T-cell immune responses during the development of acquired immune deficiency syndrome (AIDS). J. Immunol. 137:2514–2521.
- Sheppard, H. W., and M. S. Ascher. 1992. The relationship between AIDS and immunologic tolerance. J. Acquired Immune Defic. Syndr. 5:143–147.
- Shor-Posner, G., A. Basit, Y. Lu, C. Cabrejos, J. Chang, M. Fletcher, E. Mantero-Atienza, and M. K. Baum. 1993. Hypercholesterolemia is associated with immune dysfunction in early human immunodeficiency virus-1 infection. Am. J. Med. 94:515–519.
- 308. Silvestris, F., P. Cafforio, M. A. Frassanito, M. Tucci, A. Romito, S. Nagata, and F. Dammacco. 1996. Overexpression of Fas antigen on T cells in advanced HIV-1 infection: differential ligation constantly induces apoptosis. AIDS 10:131–141.
- 309. Simmonds, P., D. Beatson, R. J. G. Cuthbert, H. Watson, B. Reynolds, J. F. Peutherer, J. V. Parry, C. A. Ludlam, and C. M. Steel. 1991. Determinants of HIV disease progression: six year longitudinal study in the Edinburgh haemophilia/HIV cohort. Lancet 338:1159–1163.
- Simpson, D. M., and M. Tagliati. 1994. Neurologic manifestations of HIV infection. Ann. Intern. Med. 121:769–785.
- 311. Smith, D. H., R. A. Byrn, S. A. Marsters, T. Gregory, J. E. Groopman, and D. J. Capon. 1987. Blocking of HIV-1 infectivity by a soluble, secreted form of the CD4 antigen. Science 238:1704–1707.
- Stevenson, M., T. L. Stanwick, M. P. Dempsey, and C. A. Lamonica. 1990.
 HIV-1 replication is controlled at the level of T cell activation and proviral integration. EMBO J. 9:1551–1560.
- 313. Street, N. E., J. H. Schmacher, T. A. T. Fong, H. Bass, D. F. Fiorentino, J. A. Leverah, and T. R. Mosmann. 1990. Heterogeneity of mouse helper T cells: evidence from bulk cultures and limiting dilution cloning for precursors of Th1 and Th2 cells. J. Immunol. 144:1629–1639.
- 314. Subramanyam, M., W. G. Gutheil, W. W. Bachovchin, and B. T. Huber. 1993. Mechanism of HIV-1 Tat induced inhibition of antigen-specific T cell responsiveness. J. Immunol. 150:2544–2553.
- Suda, T., and S. Nagata. 1994. Purification and characterization of the Fas-ligand that induces apoptosis. J. Exp. Med. 179:873–879.
- 316. Sugamura, K., M. Fujii, M. Kannagi, M. Sakitani, M. Takeuchi, and Y. Hinuma. 1984. Cell surface phenotypes and expression of viral antigens of various human cell lines carrying human T-cell leukemia virus. Int. J. Cancer 34:221–228.
- 317. Sugamura, K., and Y. Hinuma. 1993. Human retroviruses: HTLV-I and HTLV-II, p. 399–435. *In J. A. Levy* (ed.), The retroviridae, vol. 2. Plenum Press, New York.
- Suzuki, M., H. Uno, A. Kiyomizu, Y. Kubuki, K. Yamashita, K. Maeda, H. Matsuoka, K. Tsuda, and H. Tsubouchi. 1995. Observation of T-cell surface antigens in the clinical course of adult T-cell leukemia: case report of a spontaneous remission. Acta Haematol. 93:40–45.
- 319. Suzuki, M., H. Uno, K. Yamashita, T. Toyama, Y. Kubuki, K. Maeda, H. Matsuoka, S. Ohtaki, and H. Tsubouchi. 1996. Clinical significance of CD45RO expression on peripheral blood mononuclear cells in HTLV-1-infected individuals. Br. J. Haematol. 92:401–409.
- 320. Suzuki, T., J.-I. Fujisawa, M. Toita, and M. Yoshida. 1993. Transactivator Tax of human T-cell leukemia virus type I (HTLV-I) interacts with cAMPresponsive element (CRE) binding and CRE modulator proteins that bind to the 21-base-pair enhancer of HTLV-I. Proc. Natl. Acad. Sci. USA 90: 610, 614.
- 321. Suzuki, T., H. Hirai, J.-I. Fujisawa, T. Fujita, and M. Yoshida. 1993. A trans-activator Tax of human T-cell leukemia virus type I binds to NF-κB p50 and serum response factor (SRF) and associates with enhancer DNAs of the NF-κB site and CarG box. Oncogene 8:2391–2397.
- 322. Swain, S. L., G. Huston, S. Tonkonogy, and A. Weinberg. 1991. Transforming growth factor-beta and IL-4 cause helper T cell precursors to develop into distinct effector helper cells that differ in lymphokine secretion pattern and cell surface phenotype. J. Immunol. 147:2991–3000.
- 323. Tanaka, A., C. Takahashi, S. Yamaoka, T. Nosaka, M. Maki, and M.

- **Hatanaka.** 1990. Oncogenic transformation by the *tax* gene of human T-cell leukemia virus type 1 *in vitro*. Proc. Natl. Acad. Sci. USA **87**:1071–1075.
- 324. Tanimura, A., H. Teshima, J.-I. Fujisawa, and M. Yoshida. 1993. A new regulatory element that augments the tax-dependent enhancer of human T-cell leukemia virus type I and cloning of cDNAs encoding its binding proteins. J. Virol. 67:5375–5382.
- 325. Tendler, C. L., S. J. Greenberg, W. A. Blattner, A. Manns, E. Murphy, T. Fleisher, B. Hanchard, O. Morgan, J. D. Burton, D. L. Nelson, and T. A. Waldman. 1990. Transactivation of interleukin 2 and its receptor induces immune activation in human T-cell lymphotropic virus I-associated myelopathy: pathogenic implications and a rationale for immunotherapy. Proc. Natl. Acad. Sci. USA 87:5219–5222.
- 326. Tendler, C. L., S. J. Greenberg, J. D. Burton, D. Danielpour, S.-J. Kim, W. A. Blattner, A. Manns, and T. A. Waldman. 1991. Cytokine induction in HTLV-1 associated myelopathy and adult T-cell leukemia: alternate molecular mechanisms underlying retroviral pathogenesis. J. Cell. Biochem. 46:302–311.
- 327. Tong-Starksen, S. E., P. A. Luciw, and B. M. Peterlin. 1989. Signaling through T lymphocyte surface proteins, TCR/CD3 and CD28, activates the HIV-1 long terminal repeat. J. Immunol. 142:702–707.
- 328. Trentin, L., S. Garbisa, R. Zambello, C. Agostini, C. Caenazzo, C. Di Francesco, A. Cipriani, E. Francavilla, and G. Semenzato. 1992. Spontaneous production of interleukin-6 by alveolar macrophages from human immunodeficiency virus type-1-infected patients. J. Infect. Dis. 166:731–737.
- Tsuda, H., R.-W. Huang, and K. Takatsuki. 1993. Interleukin-2 prevents programmed cell death in adult T-cell leukemia cells. Jpn. J. Cancer Res. 84:431–437.
- Tsuda, H., and K. Takatsuki. 1984. Specific decrease in T3 antigen density in adult T-cell leukemia cells I. Flow microfluorometric analysis. Br. J. Cancer 50:843–845.
- 331. Tsukada, N., M. Matsuda, K. Miyagi, and N. Yanagisawa. 1991. Soluble CD4 and CD8 in the peripheral blood of patients with multiple sclerosis and HTLV-1-associated myelopathy. J. Neuroimmunol. 35:285–293.
- 332. Tsuruta, L., H.-J. Lee, E. S. Masuda, N. Koyano-Nakagawa, N. Arai, K. Arai, and T. Yokota. 1995. Cyclic AMP inhibits expression of the IL-2 gene through the nuclear factor of activated T cells (NF-AT) site, and transfection of NF-AT cDNAs abrogates the sensitivity of EL-4 cells to cyclic AMP. J. Immunol. 154:5255–5264.
- 333. Tuosto, L., M. S. G. Montani, S. Lorenzeti, E. Cundari, S. Moretti, G. Lombardi, and E. Piccolella. 1995. Differential susceptibility to monomeric HIV gp120-mediated apoptosis in antigen-activated CD4⁺ T cell populations. Eur. J. Immunol. 25:2907–2916.
- 334. Tuosto, L., C. Piazza, S. Moretti, A. Modesti, R. Greenlaw, R. Lechler, G. Lombardi, and E. Piccolella. 1995. Ligation of either CD2 or CD28 rescues CD4⁺ T cells from HIV-gp120-induced apoptosis. Eur. J. Immunol. 25: 2917–2922.
- 335. Umehara, F., S. Izumo, A. T. Ronquillo, K. Matsumuro, E. Sato, and M. Osame. 1994. Cytokine expression in the spinal cord lesions in HTLV-I-associated myelopathy. J. Neuropathol. Exp. Neurol. 53:72–77.
- 336. Umehara, F., A. Nakamura, S. Izumo, R. Kubota, S. Ijichi, N. Kashio, K.-I. Hashimoto, K. Usuku, E. Sato, and M. Osame. 1994. Apoptosis of T lymphocytes in the spinal cord lesions in HTLV-1-associated myelopathy: a possible mechanism to control viral infection in the central nervous system. J. Neuropathol. Exp. Neurol. 53:617–624.
- 337. Van de Perre, P., P. Lepage, A. Simonon, C. Desgranges, D. G. Hitimana, A. Bazubagira, C. van Goethem, A. Kleinschmidt, F. Bex, K. Broliden, V. Erfle, F. Gotch, and A. Burny. 1992. Biological markers associated with prolonged survival in African children maternally infected by the human immunodeficiency virus type 1. AIDS Res. Hum. Retroviruses 8:435–442.
- 338. Vanham, G., L. Kestens, G. Penne, C. Goilav, P. Gigase, R. Colebunders, M. Vandenbruaene, J. Goeman, G. Van Der Groen, and J. L. Ceuppens. 1991. Subset markers of CD8(+) cells and their relation to enhanced cytotoxic T-cell activity during human immunodeficiency virus infection. J. Clin. Immunol. 11:345–356.
- 339. Van Noesel, C. J., R. A. Gruters, F. G. Terpstra, P. T. A. Schellekens, R. A. W. van Lier, and F. Miedema. 1990. Functional and phenotypic evidence for a selective loss of memory T cells in asymptomatic human immunodeficiency virus-infected men. J. Clin. Invest. 86:293–299.
- 340. Vazeux, R., C. Lacroix-Ciaudo, S. Blanche, M.-C. Cumont, D. Henin, F. Gray, L. Boccon-Gibod, and M. Tardieu. 1992. Low levels of human immunodeficiency virus replication in the brain tissue of children with severe acquired immunodeficiency syndrome encephalopathy. Am. J. Pathol. 140: 137–144.
- Verdier, M., J. Bonis, G. Leonard, M. Dumas, and F. Denis. 1993. HTLV-1 antibody class and subclass distribution in African TSP and control populations. J. Neuroimmunol. 42:117–120.
- 342. Vigano, A., N. Principi, M. L. Villa, C. Riva, L. Crupi, D. Trabattoni, G. M. Shearer, and M. Clerici. 1995. Immunologic characterization of children vertically infected with human immunodeficiency virus, with slow or rapid disease. J. Pediatr. 126:368–374.
- 343. Viscidi, R. P., K. Mayur, H. M. Lederman, and A. D. Frankel. 1989.

Inhibition of antigen-induced lymphocyte proliferation by Tat protein from HIV-1. Science **246**:1606–1608.

- 344. Vyakarnam, A., J. McKeating, A. Meager, and P. C. Beverly. 1990. Tumor necrosis factor (alpha, beta) induced by HIV-1 in peripheral blood mononuclear cells potentiate virus replication. AIDS 4:21–27.
- 345. Wagner, S., and M. R. Green. 1993. HTLV-I Tax protein stimulation of DNA binding of bZIP proteins by enhancing dimerization. Science 262: 395–399.
- Walker, B. D., and F. Plata. 1990. Cytotoxic T-lymphocytes against HIV. AIDS 4:177–184.
- 347. Walker, C. M., D. J. Moody, D. P. Stites, and J. A. Levy. 1986. CD8+ lymphocytes can control HIV infection in vitro by suppressing virus replication. Science 234:1563–1566.
- Weaver, C. T., C. M. Hawrylowicz, and E. R. Unanue. 1988. T helper cell subsets require the expression of distinct costimulatory signals by antigenpresenting cells. Proc. Natl. Acad. Sci. USA 85:8181–8185.
- 349. Wei, X., S. K. Ghosh, M. E. Taylor, V. A. Johnson, E. A. Emini, P. Deutsch, J. D. Lifson, S. Bonhoeffer, M. A. Nowak, B. H. Hahn, M. S. Saag, and G. Shaw. 1995. Viral dynamics in human immunodeficiency virus type 1 infection. Nature (London) 373:117–122.
- 350. Weinhold, K. J., H. K. Leyerly, S. D. Stanley, A. A. Austin, T. J. Matthews, and D. P. Bolognesi. 1989. HIV-1 gp120-mediated immune suppression and lymphocyte suppression and lymphocyte destruction in the absence of viral infection. J. Immunol. 142:3091–3097.
- 351. Werner, T., S. Ferroni, T. Saermark, R. Brack-Werner, R. B. Banati, R. Mager, L. Steinaa, G. W. Kreutzberg, and V. Erfle. 1991. HIV-1 Nef protein exhibits structural and functional similarity to scorpion peptides interacting with K⁺ channels. AIDS 5:1301–1308.
- 352. Westendorp, M. O., R. Frank, C. Ochsenbauer, K. Stricker, J. Dhein, H. Walczak, K.-M. Debatin, and P. H. Krammer. 1995. Sensitization of T cells to CD95-mediated apoptosis by HIV-1 Tat and gp120. Nature (London) 375:497–500.
- 353. Wucherpfennig, K., P. Hollsberg, J. H. Richardson, D. Benjamin, and D. A. Hafler. 1992. T cell activation by autologous human T cell leukemia virus type I-infected T cell clones. Proc. Natl. Acad. Sci. USA 89:2110–2114.
- Wyllie, A. H. 1980. Glucocorticoid-induced thymocyte apoptosis is associated with endogenous endonuclease activation. Nature (London) 284:555

 556.
- 355. Yamada, T., S. Yamaoka, T. Goto, M. Nakai, Y. Tsujimoto, and M. Hatanaka. 1994. The human T-cell leukemia virus type I Tax protein induces apoptosis which is blocked by the Bcl-2 protein. J. Virol. 68:3374–3379.
- 356. Yodoi, J., and T. Uchiyama. 1992. Diseases associated with HTLV-1: virus, IL-2 receptor dysregulation and redox regulation. Immunol. Today 13:405–411
- Yodoi, J., Y. Uchiyama, and M. Maeda. 1983. T-cell growth factor receptor in adult T-cell leukemia. Blood 62:509–511.

- 358. Yonehara, S., A. Ishii, and M. Yonehara. 1989. A cell-killing monoclonal antibody (anti-Fas) to a cell surface antigen co-downregulated with the receptor of tumor necrosis factor. J. Exp. Med. 169:1747–1755.
- 359. Yoshida, J. 1994. Host-HTLV type I interaction at the molecular level. AIDS Res. Hum. Retroviruses 10:1193–1197.
- 360. Yoshida, M., J. Inoue, J. Fujisawa, and M. Seiki. 1989. Molecular mechanisms of regulation of HTLV-1 gene expression and its association with leukemogenesis. Genome 31:662–667.
- 361. Yoshida, M., I. Miyoshi, and Y. Hinuma. 1982. Isolation and characterization of retrovirus from cell lines of human and adult T-cell leukemia and its implication in the disease. Proc. Natl. Acad. Sci. USA 79:2031–2035.
- Yoshida, M., and M. Seiki. 1987. Recent advances in the molecular biology of HTLV-1: transactivation of viral and cellular genes. Annu. Rev. Immunol. 5:541–559.
- 363. Yssel, H., R. D. W. Malefyt, M. Duc Dodon, D. Blanchard, L. Gazzolo, J. E. de Vries, and H. Spits. 1989. Human T cell leukemia/lymphoma virus type I infection of a CD4+ proliferative/cytotoxic T cell clone progresses in at least two distinct phases based on changes in function and phenotype of the infected cells. J. Immunol. 142:2279–2289.
- 364. Zack, J. A., S. J. Arrigo, S. R. Weitsman, A. S. Go, A. Haislip, and I. S. Y. Chen. 1990. HIV-1 entry into quiescent primary lymphocytes: molecular analysis reveals a labile, latent viral structure. Cell 61:213–222.
- 365. Zanussi, S., C. Simonelli, M. D'Andrea, C. Caffau, M. Clerici, U. Tirelli, and P. de Paoli. 1996. CD8⁺ lymphocyte phenotype and cytokine production in long-term non-progressor and in progressor patients with HIV-1 infection. Clin. Exp. Immunol. 105:220–224.
- 366. Zauli, G., D. Gibellini, A. Caputo, A. Bassini, M. Negrini, M. Monne, M. Mazzoni, and S. Capitani. 1995. The human immunodeficiency virus type-1 Tat protein upregulates bcl-2 gene expression in Jurkat T-cell lines and primary peripheral blood mononuclear cells. Blood 86:3823–3834.
- 367. Zauli, G., D. Gibellini, D. Milani, P. Borgatti, M. La Placa, and S. Capitani. 1993. Human immunodeficiency virus type 1 tat protein protects lymphoid, epithelial and neuronal cell lines from death by apoptosis. Cancer Res. 53:4481–4485.
- 368. Zhao, L. J., and C. Z. Giam. 1992. Human T-cell lymphotropic virus type I (HTLV-I) transcriptional activator, Tax, enhances CREB binding to HTLV-I 21-base-pair repeats by protein-protein interaction. Proc. Natl. Acad. Sci. USA 89:7070–7074.
- 369. Ziegler-Heitbrock, H. W. L., D. Stachel, T. Schlunk, L. Gurtler, W. Schramm, M. Froschl, J. R. Bogner, and G. Riethmuller. 1988. Class II (DR) antigen expression on CD8⁺ lymphocyte subsets in acquired immune deficiency syndrome (AIDS). J. Clin. Immunol. 8:1–6.
- 370. **Zola-Pazner, S., D. William, W. El-Sadr, M. Marmor, and R. Stahl.** 1984. Quantitation of β_2 -microglobulin and other immune characteristics in a progressive study of men at risk for acquired immunodeficiency syndrome. JAMA 22:2951–2955.