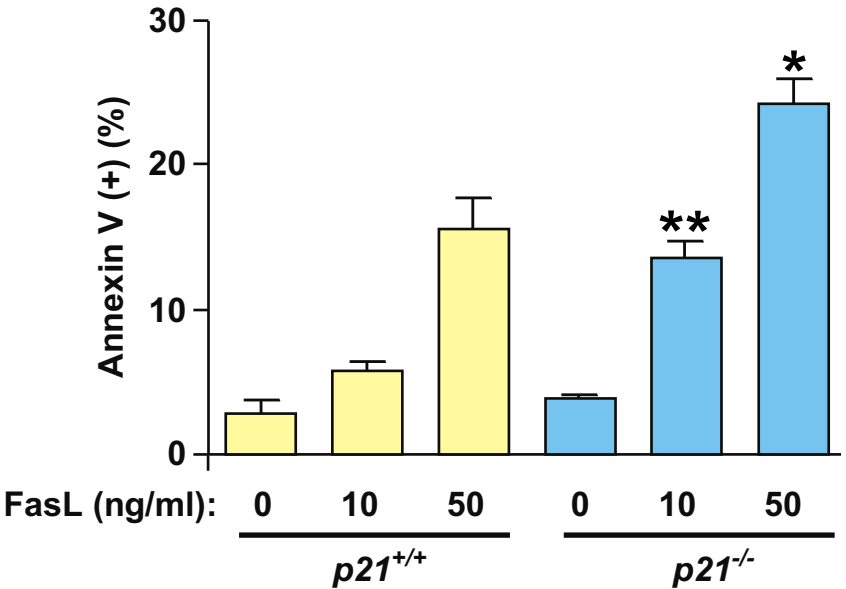
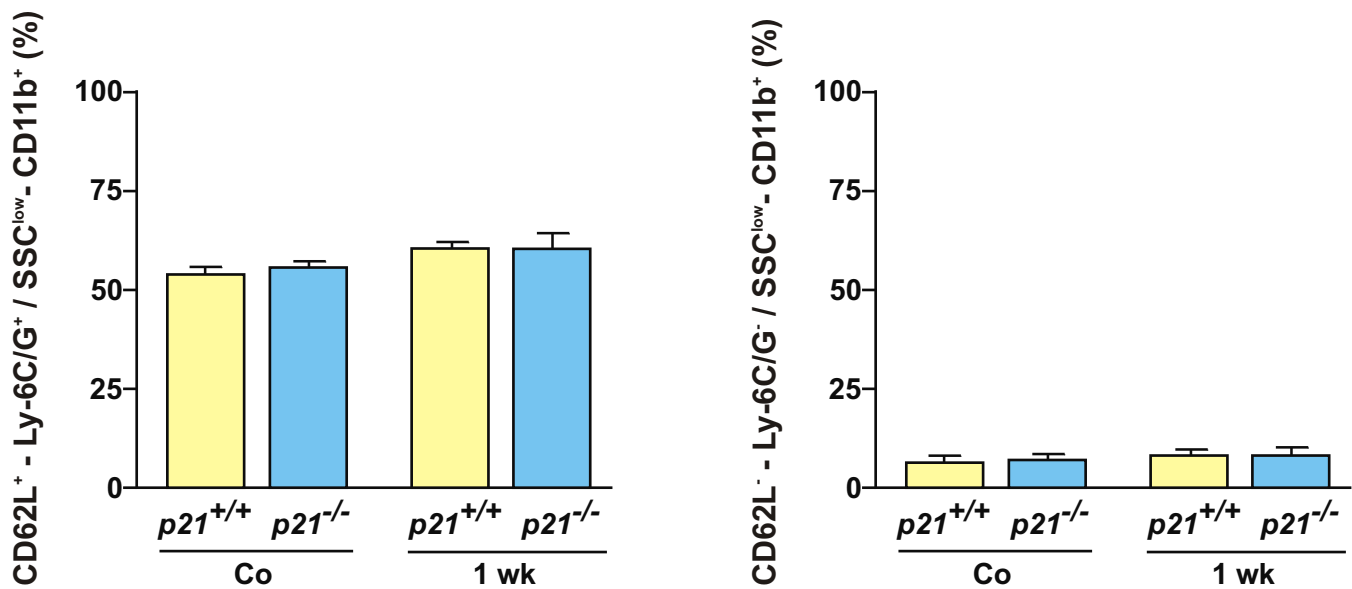


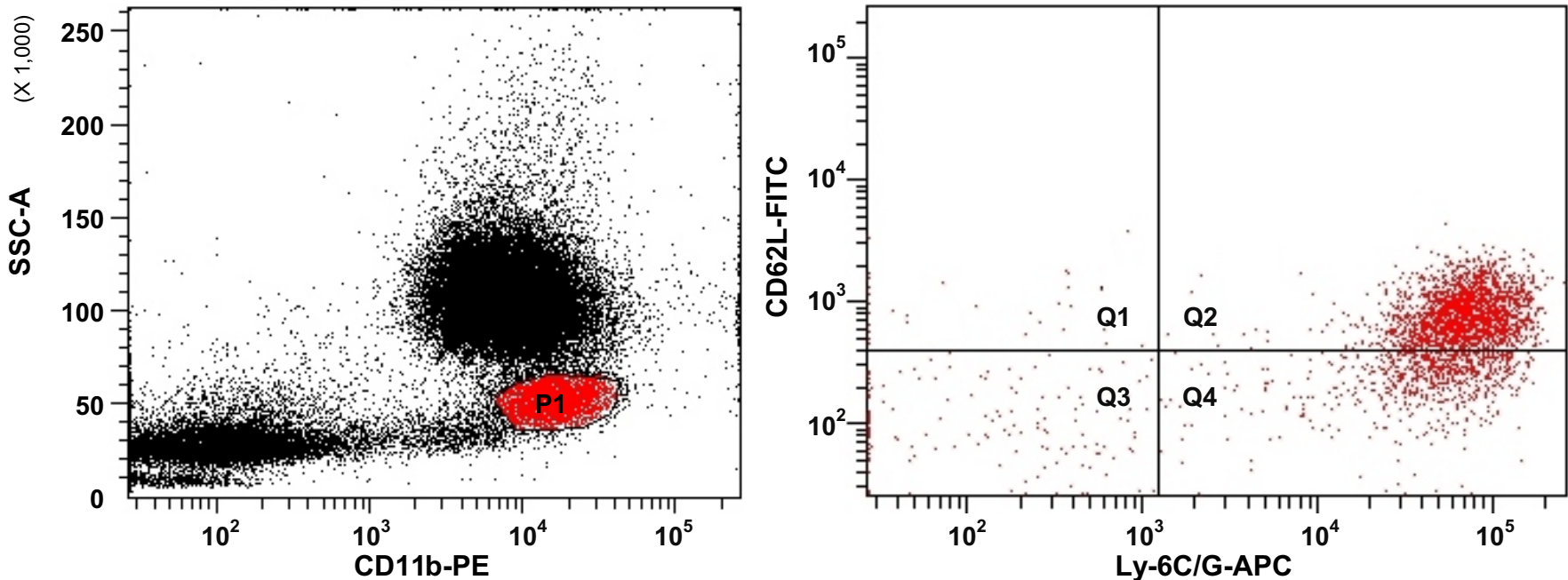
Supplemental Fig 1



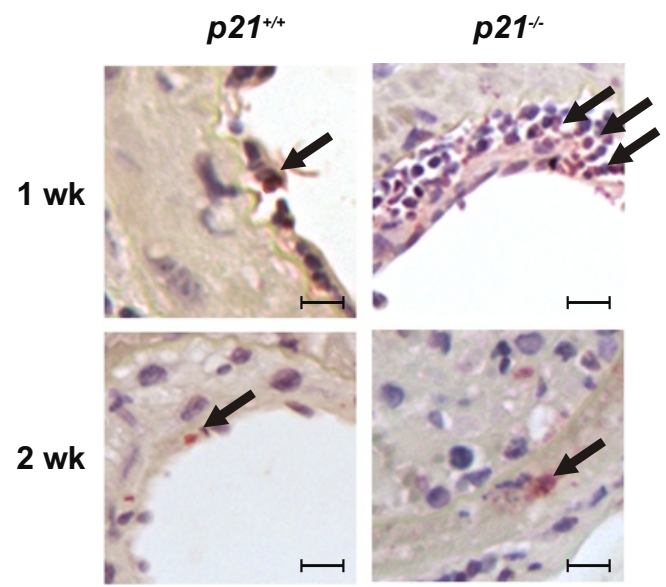
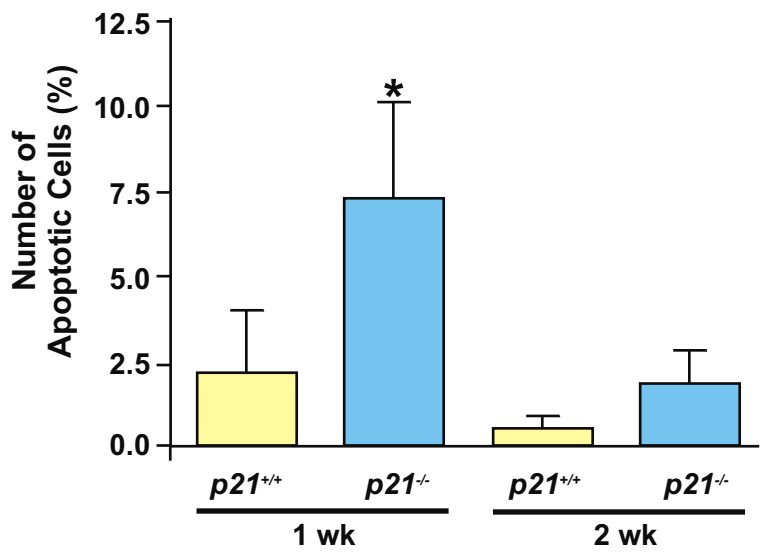
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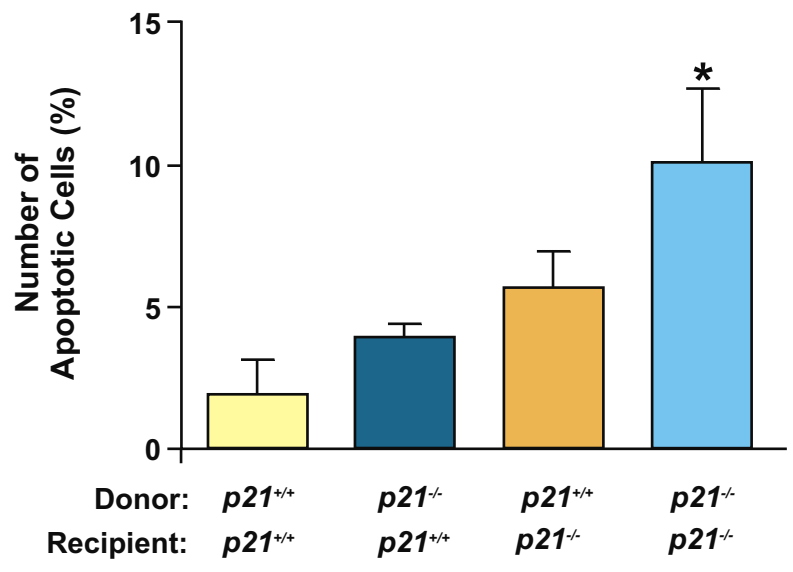
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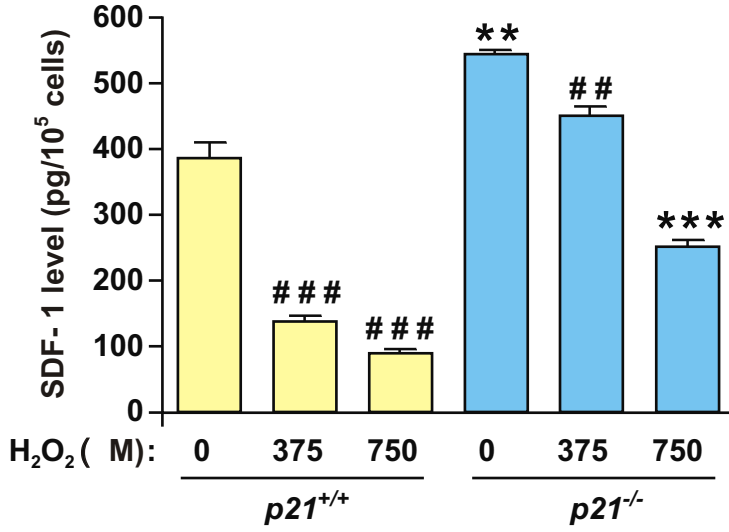
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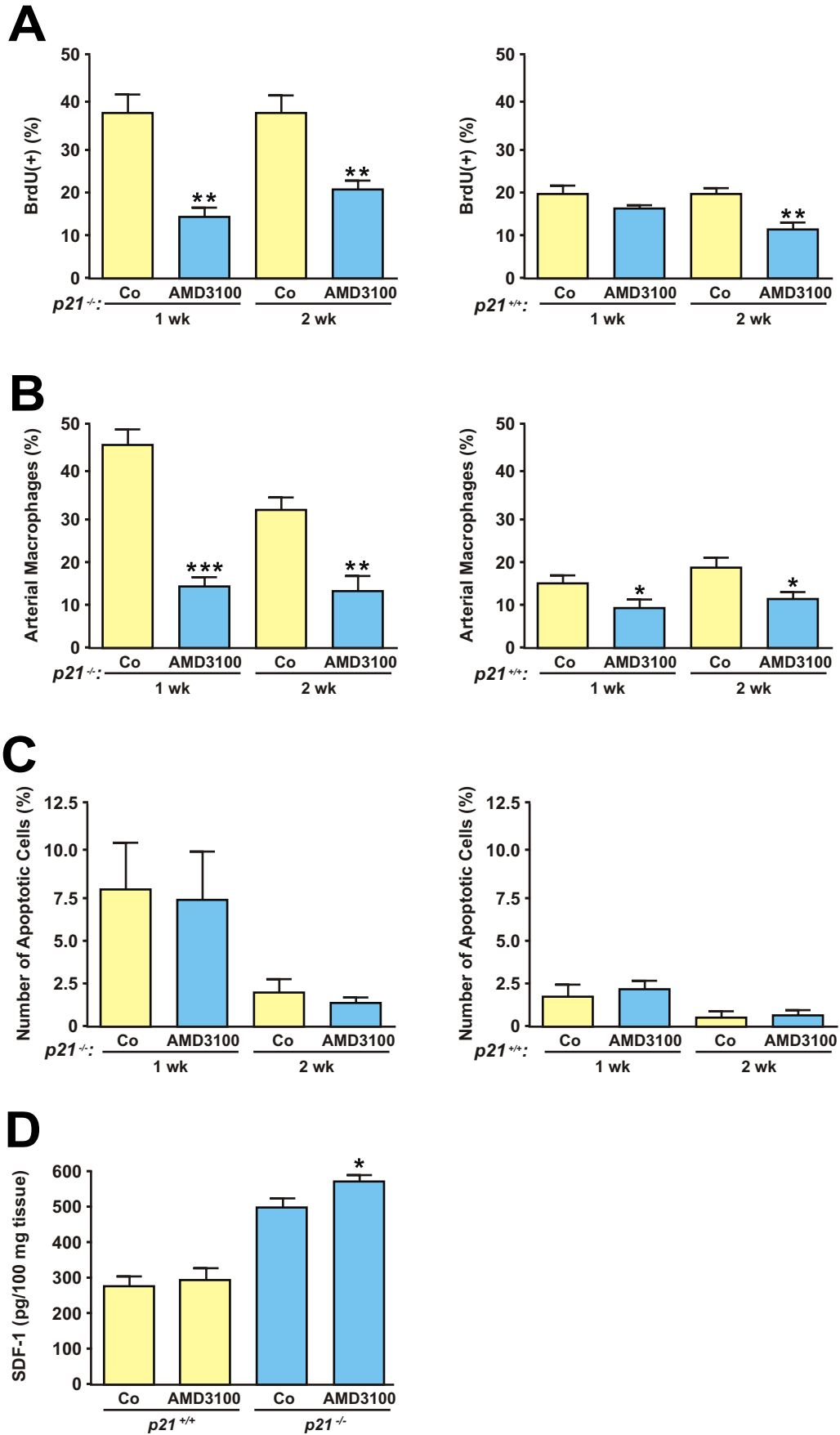


B



Supplemental Fig 4





Supplemental Figure legends:

Supplemental Figure 1: Accelerated apoptosis in $p21^{-/-}$ VSMCs in vitro. $p21^{-/-}$ VSMCs in culture (blue bars) displayed increased apoptosis compared to $p21^{+/+}$ VSMCs (yellow bars) with in vitro administration of 0, 10 or 50 ng / ml of FasL ($n=3$; * $P < 0.05$, ** $P < 0.01$ for comparison of $p21^{-/-}$ to $p21^{+/+}$ at identical FasL doses).

Supplemental Figure 2: Immunophenotyping of peripheral blood before and after wire vascular injury in $p21^{+/+}$ and $p21^{-/-}$ mice. (A) Peripheral blood leukocytes were isolated and stained with CD11b-PE, Ly-6C/G-APC and CD62L-FITC and the percentages of Ly-6C/G⁺/CD62L⁺ and Ly-6C/G⁻/CD62L⁻ monocytes were determined ($n=3$). (B) Representative dot blots. Left: acquisition of the side scatter-A (SSC-A) vs. CD11b-PE. P1 represents the gated CD11b-PE positive SSC-A low cells. Right: acquisition of Ly-6C/G-APC and CD62L-FITC cells present in the CD11b-PE population. No differences were observed between $p21^{+/+}$ and $p21^{-/-}$ mice.

Supplemental Figure 3: Cell apoptosis in injured arteries from $p21^{+/+}$ and $p21^{-/-}$ mice. (A) Left panel: percentage of apoptotic cells in arteries from $p21^{+/+}$ and $p21^{-/-}$ mice 1 and 2 weeks after vascular injury ($n=5$; * $P < 0.05$ $p21^{+/+}$ vs. $p21^{-/-}$ 1 wk). Right panel: TUNEL staining of the arteries following vascular injury in $p21^{-/-}$ and $p21^{+/+}$ mice 1 and 2 weeks after injury. Arrows indicate apoptotic cells. Scale bar = 7 μ m. (B) One week after femoral artery wire injury, the number of TUNEL positive cells was decreased in arterial tissue from $p21^{+/+}$ mice receiving $p21^{+/+}$ BM compared to $p21^{-/-}$ mice receiving

$p21^{-/-}$ BM ($n=5$; $*P < 0.05$). No other between group comparisons reached statistical significance.

Supplement Figure 4: SDF-1 levels are decreased in $p21^{-/-}$ and $p21^{+/+}$ VSMCs upon oxidative stress. Increased SDF-1 levels were observed in $p21^{-/-}$ compared to $p21^{+/+}$ VSMCs prior to oxidative stress. The induction of oxidative stress, with 375 or 750 μM of H_2O_2 , caused a reduction in SDF-1 levels in both $p21^{-/-}$ (blue bars) and $p21^{+/+}$ (yellow bars) VSMCs ($n=3$; $**P < 0.01$ vs. $p21^{+/+}$ 0 μM , $^{##}P < 0.01$ vs. $p21^{-/-}$ 0 μM , $^{***}P < 0.001$ vs. $p21^{-/-}$ 0 μM , $^{###}P < 0.001$ vs. $p21^{+/+}$ 0 μM).

Supplement Figure 5: SDF-1 inhibition induced by AMD3100 administration prevents excessive proliferation during vascular wound repair. (A) AMD3100 treatment decreased cellular proliferation as assessed by BrdU incorporation at 1 and 2 weeks after vascular injury in $p21^{-/-}$ arteries and at 2 weeks after vascular injury in $p21^{+/+}$ arteries (left: $n=4$; $**P < 0.01$ vs. $p21^{-/-}$ 1 wk; $**P < 0.01$ vs. $p21^{-/-}$ 2 wk), (right: $n=4$; $**P < 0.01$ vs. $p21^{+/+}$ 2 wk). (B) AMD3100 treatment reduced the number of local arterial macrophages in $p21^{-/-}$ and $p21^{+/+}$ arteries at 1 and 2 weeks after vascular injury (left: $n=5$; $^{***}P < 0.001$ vs. $p21^{-/-}$ 1 wk; $**P < 0.01$ vs. $p21^{-/-}$ 2 wk), (right: $n=10$; $*P < 0.05$ vs. $p21^{+/+}$ 1 wk; $*P < 0.05$ vs. $p21^{+/+}$ 2 wk). (C) The number of local apoptotic TUNEL positive cells after vascular injury was unchanged after treatment with AMD3100. (D) One week after vascular injury, $p21^{-/-}$ mice treated with AMD3100 displayed marginally increased local vascular SDF-1 levels compared to control $p21^{-/-}$

mice ($n=5$; $*P < 0.05$ vs. Co $p21^{-/-}$). AMD3100 administration did not influence SDF-1 tissue levels in $p21^{+/+}$ mice.

Supplemental Table 1: Peripheral blood cell analysis of $p21^{+/+}$ and $p21^{-/-}$ mice before and after wire injury. Blood was collected from non-injured mice (Co) and 1 week following wire injury and analyzed using a Cell-dyne 3500 hematology analyzer ($n=3$).

	$p21^{+/+}$		$p21^{-/-}$	
	Co	1 wk	Co	1 wk
Monocytes/mm ³	191.6 ± 47.7	169.2 ± 45.7	233.8 ± 55.7	294 ± 94.7
Monocytes (%)	4.8 ± 1.3	4.2 ± 1.2	5.9 ± 1.6	5.2 ± 1.5
Neutrophils/mm ³	422.4 ± 108.9	1362.0 ± 250.0	664.6 ± 73.4	1656.0 ± 202.3
Neutrophils (%)	10.4 ± 2.8	35.6 ± 6.7	16.3 ± 1.8	42.1 ± 5.9
Lymphocytes/mm ³	2538.0 ± 348.8	3164.0 ± 130.9	2462.0 ± 277.2	2173.0 ± 243.3
Lymphocytes (%)	61.8 ± 9.1	59.3 ± 6.9	76.8 ± 3.3	52.4 ± 5.6

Supplemental Methods

Primer Sequences:

mp21^{Cip1}: forward 5'-GAC CTG GGA GGG GAC AAG AG-3'
reverse 5'- TTC TCT TGC AGA AGA CCA ATC-3'

mp53: forward 5'-TGA GGT TCG TGT TTG TGC CTG-3'
reverse 5'-GGT AGC TGG AGT GAG CCC TGC-3'

mSDF-1: forward 5'-ACA CTC CAA ACT GTG CCC TTC AGA-3'
reverse 5'-ATG CTG GCA AAC CTT AGC ATG ACC-3'

18S: forward 5'-TTT CGG AAC TGA GGC CAT GA-3'
reverse 5'-GCA AAT GCT TTC GCT CTG GTC-3'

ChIP mSDF-1 promoter STAT3 binding site:

forward 5'-ACC TGT TTG GTC TCT TTG CTC GGT-3'
reverse 5'-CTG TCA AAG GCA CAA GCC GTG AAA-3'

β -actin genomic control (ChIP):

forward 5'-TTT CCC TGA GCA GCT TGT CA-3'
reverse 5'-CTG GGC CGT TAG CTA GTG TC-3'