HYDROGEN PEROXIDE PROMOTES Aβ PRODUCTION THROUGH JNK-DEPENDENT ACTIVATION OF γ-SECRETASE

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SUPPLEMAENTAL DATA

SUPPLEMENTARY MATERIALS AND METHODS

Plasmids - Expression vectors encoding GST-c-Jun-(1-79), HA-JNKK2-JNK1, HA-JNKK2(KM)-JNK1 have been described previously (1,2). To construct GST-PS1 bacterial expression vectors, PS1 cDNA was amplified by PCR from the pcDNA3.1-PS1 plasmid (3). The PCR product was digested with BamHI and EcoRI enzymes, and cloned into pGEX-4T-1 vectors (Amersham Pharmacia).

Subcellular Fractionation - SH-SY5Y/APP695myc cells were harvested and homogenized in cell lysis buffer containing 50 mM Tris-HCl (pH 7.4), 150 mM NaCl, 1 mM EDTA, 0.25 mM PMSF and cocktails of protease inhibitors. The homogenate was centrifuged for 10 min at $1,000 \times g$ at 4°C to prepare a post-nuclear supernatant fraction. Membranes were pelleted from the PNS by centrifugation for 30 min at $13,000 \times g$. The resulting supernatant was designated as cell cytoplasm. All fractions were subjected to Western blot.

Purification of Recombinant Proteins - GST-PS1 and GST-c-Jun-(1-79) proteins were purified using glutathione-agarose according to the manufacturer's procedure (Amersham Pharmacia).

Protein Kinase Assay - HEK293T cells were transfected with expression vectors encoding HA-JNKK2-JNK1, HA-JNKK2(KM)-JNK1 (1 μ g each) for 24 h as indicated. Cells were lysed and HA-JNKK2-JNK1 proteins were immunoprecipitated by anti-HA antibody (1:1,000, Sigma). The activity of the JNK immunocomplex was measured by kinase assays with GST-c-Jun-(1-79) and GST-PS1 as substrates. The reaction was incubated at 30°C for 1 h in 30 μ l of kinase buffer in the presence of 10 μ M ATP/10 μ Ci [γ -³²P]ATP (10 Ci/mmol). The reactions were terminated with 4× Laemmli sample buffers. The proteins were resolved by 13% SDS-PAGE, followed by autoradiography.

SUPPLEMENTARY FIGURE LEGENDS

FIGURE S1. The characteristics of APP processing products. A. HEK293T cells were transiently transfected with expression vectors encoding AICD57myc (lane 2), AICD59myc (lane 3), SP-C99myc (lane 4) as positive controls to recognize the bands. pcDNA3.1 vectors were transfected as negative controls (lanes 1 and 6). SH-SY5Y/APP695myc cells were subjected to subcellular fractionation followed by Western blot (lanes 7, 8 and 9). APP695myc and C83myc in cytoplasm was a result of membrane contamination from incomplete subcellular fractionation separation. B. HEK293T cells were transiently transfected with expression vectors encoding SP-C99myc (lane 2) and APP695myc (lanes 3, 4, 5 and 6). After 24 h, cells were pretreated with 10 μ M DAPT (lanes 5 and 6) or DMSO (lanes 3 and 4) for 3 h, and treated with 1 mM H₂O₂ for another hour. Cells were harvested and analyzed by immunoblotting with APP c-terminal antibody (anti-APP676-695, 1:4,000, Sigma). C. HEK293T cells were transiently transfected with expression vectors encoding APP695 (lanes 2 and 3). After 24 h, cells were transiently transfected with expression vectors encoding APP695 (lanes 2 and 3). After 24 h, cells were transiently model of the expression vectors encoding APP695 (lanes 2 and 3). After 24 h, cells were transiently transfected with expression vectors encoding APP695 (lanes 2 and 3). After 24 h, cells were transiently transfected with expression vectors encoding APP695 (lanes 2 and 3). After 24 h, cells were transiently transfected with expression vectors encoding APP695 (lanes 2 and 3). After 24 h, cells were transiently transfected with 10 mmunoblotting with anti-APP676-695.

FIGURE S2. Activated-JNK directly phosphorylates GST-PS1 in vitro. HEK293T cells were transfected with expression vectors encoding HA-JNKK2-JNK1, HA-JNKK2(KM)-JNK1. Cells were

lysed and the JNK activity of the immunocomplex was measured by kinase assays with GST-c-Jun-(1-79) and GST-PS1 as substrates. An aliquot of each sample was analyzed for the expression of HA-JNKK2-JNK1 and HA-JNKK2(KM)-JNK1 by immunoblotting using anti-HA antibody (1:10,000). KA, kinase assay. IB, immunoblotting. IP, immunoprecipitation. CS, Coomassie blue staining.

FIGURE S3. Effects of cell signal inhibitors on H_2O_2 -triggered cell death. SH-SY5Y/APP695myc cells were serum-starved for 12 h and pretreated with U0126 (5 μ M) (c and d), Wortmannin (20 nM) (e and f), SP600125 (20 μ M) (g and h) or the control DMSO (a and b) for 3 h. With exposure to 1 mM H_2O_2 for 1 h, morphological images of cells were taken with Olympus BX50 microscopy to determine the effects of cell signal inhibitors on H_2O_2 -triggered cell death.

REFERENCES

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FIGURE S1



IB: α-myc



A

l



С



FIGURE S2



FIGURE S3

