

**Supplemental Table 1. AbrB does not repress *sinI***

Genotype <sup>a</sup>	<i>PsinI-lacZ</i> activity (MU) <sup>b</sup>
Wild type	5.8 ± 0.4
<i>epsH</i>	7 ± 2
<i>abrB epsH</i>	3.8 ± 0.1

a. The following *PsinI-lacZ* containing strains were used FC149, FC321, FC346.

b. β-Galactosidase activity, calculated in Miller Units (MU), was measured from late-exponential phase cells grown in MSgg and is the average of at least three replicates.

**Table S2. Strains<sup>a</sup>**

Strain	Genotype	
3610	Undomesticated wild strain	
FC13	$\Delta$ <i>epsH::tet; amyE::P<sub>eps</sub>-lacZ</i> (cat)	Kearns et al., 2005
FC14	$\Delta$ <i>sinR::spec; ΔepsH::tet; amyE::P<sub>eps</sub>-lacZ</i> (cat)	Kearns et al., 2005
FC26	$\Delta$ <i>slr::tet</i>	
FC37	$\Delta$ <i>slr::tet; ΔsinR::spec</i>	
FC134	$\Delta$ <i>epsH::tet; amyE::P<sub>yqxM</sub>-lacZ</i> (cat)	Chu et al., 2006
FC135	$\Delta$ <i>sinR::spec; ΔepsH::tet; amyE::P<sub>yqxM</sub>-lacZ</i> (cat)	Chu et al., 2006
FC149	<i>amyE::P<sub>sinI</sub>-lacZ</i> (cat)	
FC171	$\Delta$ <i>sipW::tet</i>	Chu et al., 2006
FC280	$\Delta$ <i>abrB::kan</i>	
FC315	$\Delta$ <i>yoaW::tet</i>	
FC321	$\Delta$ <i>epsH::tet; amyE::P<sub>sinI</sub>-lacZ</i> (cat)	
FC346	$\Delta$ <i>epsH::tet; ΔabrB::kan; amyE::P<sub>sinI</sub>-lacZ</i> (cat)	
FC413	$\Delta$ <i>spo0A::spec; amyE::P<sub>eps</sub>-lacZ</i> (cat)	
FC414	$\Delta$ <i>spo0A::spec; amyE::P<sub>yqxM</sub>-lacZ</i> (cat)	
FC415	$\Delta$ <i>epsH::tet; ΔabrB::kan; amyE::P<sub>eps</sub>-lacZ</i> (cat)	
FC416	$\Delta$ <i>epsH::tet; ΔabrB::kan; amyE::P<sub>yqxM</sub>-lacZ</i> (cat)	
FC444	$\Delta$ <i>epsH::tet; Δspo0A::spec; ΔabrB::kan; amyE::P<sub>eps</sub>-lacZ</i> (cat)	
FC445	$\Delta$ <i>epsH::tet; Δspo0A::spec; ΔabrB::kan; amyE::P<sub>yqxM</sub>-lacZ</i> (cat)	
FC480	$\Delta$ <i>epsH::tet; ΔsinR::spec; ΔabrB::kan; amyE::P<sub>eps</sub>-lacZ</i> (cat)	
FC481	$\Delta$ <i>epsH::tet; ΔsinR::spec; ΔabrB::kan; amyE::P<sub>yqxM</sub>-lacZ</i> (cat)	
FC488	<i>amyE::-112 P<sub>yqxM</sub>-lacZ</i> (cat)	
FC495	<i>amyE::-64 P<sub>yqxM</sub>-lacZ</i> (cat)	
FC503	$\Delta$ <i>slr::tet; ΔabrB::kan</i>	
FC504	$\Delta$ <i>slr::tet; ΔabrB::kan; amyE::P<sub>yqxM</sub>-lacZ</i> (spec)	
FC513	$\Delta$ <i>slr::tet; ΔabrB::kan; amyE::P<sub>eps</sub>-lacZ</i> (cat)	

FC518	<i>amyE</i> ::P <sub>slr</sub> - <i>lacZ</i> (cat)	
FC519	Δ <i>epsH</i> ::tet; <i>amyE</i> ::P <sub>slr</sub> - <i>lacZ</i> (cat)	
FC520	Δ <i>epsH</i> ::tet; Δ <i>sinR</i> ::spec; <i>amyE</i> ::P <sub>slr</sub> - <i>lacZ</i> (cat)	
FC522	Δ <i>epsH</i> ::tet; Δ <i>abrB</i> ::kan; <i>amyE</i> ::P <sub>slr</sub> - <i>lacZ</i> (cat)	
FC525	Δ <i>slr</i> :: <i>mls</i>	
FC526	Δ <i>epsH</i> ::tet; Δ <i>slr</i> :: <i>mls</i> ; <i>amyE</i> ::P <sub>eps</sub> - <i>lacZ</i> (cat)	
FC527	Δ <i>epsH</i> ::tet; Δ <i>sinR</i> ::spec; Δ <i>slr</i> :: <i>mls</i> ; <i>amyE</i> ::P <sub>eps</sub> - <i>lacZ</i> (cat)	
FC529	Δ <i>epsH</i> ::tet; Δ <i>sinR</i> ::spec; Δ <i>slr</i> :: <i>mls</i> ; <i>amyE</i> ::P <sub>yqxM</sub> - <i>lacZ</i> (cat)	
FC536	Δ <i>slr</i> :: <i>mls</i> ; <i>amyE</i> ::P <sub>slr</sub> - <i>slr</i> (cat)	
FC591	<i>amyE</i> ::P <sub>yqxM</sub> - <i>lacZ</i> (cat)	Chu et al., 2006
FC593	Δ <i>slr</i> :: <i>mls</i> ; <i>amyE</i> ::P <sub>yqxM</sub> - <i>lacZ</i> (cat)	
FC600	Δ <i>slr</i> :: <i>mls</i> ; <i>yqxM</i> Ω P <sub>IPTG</sub> - <i>yqxM-sipW-tasA</i> (spec)	
ALM16	<i>amyE</i> ::-100 P <sub>yqxM</sub> - <i>lacZ</i> (cat)	
ALM35	<i>amyE</i> ::5' site mutation P <sub>yqxM</sub> - <i>lacZ</i> (cat)	
ALM36	<i>amyE</i> ::3' site mutation P <sub>yqxM</sub> - <i>lacZ</i> (cat)	
ALM37	<i>amyE</i> ::heptad repeat mutation P <sub>yqxM</sub> - <i>lacZ</i> (cat)	
DS1674	Δ <i>epsH</i> ::tet; Δ <i>sinR</i> ::kan	

a. All strains are derivatives of 3610

**Table S3.** Primers<sup>a</sup>

Primer	Sequence (5'→3')
-499sinI	tggcGAATTCAggtttcaactgacgtctcaaataatgt
-2sinI	gccAGGATCCAggttctccctctaaaatacttgttta
Slr1F	tttgcatttatgtatcgagaaaatgtca
Slr2R	caattcgccctatagtggatgtcgatcggtacaaacggataattttcca
Slr2_tetR	gaacaacctgcaccattgtcaagaacgggtacaaacggataattttcca
Slr3F	ccagctttgtcccttagtgagggtctgtccatgaagtcaaattcctt
Slr3_tetF	ttgatccttttataacaggaaattctgtctgtccatgaagtcaaattcctt
Slr4R	aattaatagcggaataccggaaagca
slrcomp_F	cgcGGATCCgtattcatagccttcagcc
slrcomp_R	ccgGAATTCAacgttagcccttaaccgtca
yoaW1_F	agaacaaggcataagaatagaatca
yoaWP2_R	caattcgccctatagtggatgtcgatggaaatcacatcaacatctttcaa
yoaWP3_F	ccagctttgtcccttagtgagggtttcagttgtcagccaga
yoaW4_R	ttgcgatgaaagttaggaattcatga
Pslr-F1	gtcGAATTCTagacaatcgcatataatttttg
Pslr-R1	gtcGGATCCctagaaattctcttattctgtcg
+1slr	gttcCATATGattggaagaattatcggttgc
+339slr	aggcCTCGAGgtaaaggcaggttcaggctgtt
-302yqxM	tggcGAATTCatagacaatcacatgtttgatca
-26yqxM	gccAGGATCCatcttacccctgtaaaacactgtaa
-175yqxM	gttcGAATTCTgagcaatactgagcaagactttgt

-160yqxM	gttcGAATTCAgcaagactttaatatgtga
-126yqxM	gttcGAATTCAaacgaacaaaatgagcgattcggt
-270yqxM	tggcGAATTCTgtcaccccttcttgttattattac
+14yqxM	gccaaGGATCCaaacaatcgaaacataatcttacctct
-21yqxM_R	gccaaGGATCCtgtaacttgatgatcgttctc
-244yqxM_F	gttcGAATTCCaccaaataataatggatgcatattaa
ALM1	gttcCTCGAG agactttgataatgtatgaaaacattct
ALM2	gttcCTCGAGgtatgaaagtgttttcaatttttagaaattttggga
ALM3	gttcCTCGAGgtattgtcattttcaatttttagaaatt
ALM4	gttcCTCGAGatactgagcaagactttgataatgtat
ALM5	gttcCTCGAGtttttcaatttttagaaattttggga
ECH318	ggGGAATTCatatgttatgaaatctactggattgtat
ECH336	accgCTCGAGtcatttaaggtttgaagctgggtttggaa
ECH341	aaactcactggcagcgatatcg
ECH342	tctatttgtttaagagccatatctaag
ECH343	cggttatacaaggattcatcgagc
ECH344	caatttgttgcgcgttcacttcat
-246epsA	gttcGAATTCTaatatgaaattctcttattccgt
+4epsA	gccaaGGATCCtcatgtattcatgccttcagcctt
JSK362	ggcAAGCTTacaataaggagaactactatgagtaaaggagaagaactttcac
JSK445	ggcGGATCCttattgtatgtcatccatgcat

<sup>a</sup>Capital letters designate restriction enzyme sites added to the primers

### Supplementary Figure legends

**Figure S1. Effects of AbrB-regulated genes on biofilm formation.** ‘Colony’ column depicts 16 X images of colonies grown on MSgg semi-solid agar medium for 3 days at 22°C. Scale bar is 1 mm. ‘Pellicle’ column depicts top-down images of 6-well microtitre plates in which cells have been grown statically in MSgg medium for 3 days at 22°C. Scale bar is 1 cm. The indicated wild type and mutant strains were as follows: wild type (3610), *sipW* (FC171), and *yoaW* (FC315).

**Figure S2: Antibodies generated to SinR cross-react with Slr.** Western blot using α-SinR as primary antibody. Whole cell lysates of the strains were loaded as follows: wild

type (3610); *slr* (FC26); *sinR slr* (FC37); *sinR* (DS1674). Lanes were loaded and normalized to 0.1 OD600 cell equivalents.

**Figure S3. A *sinR* mutation, but not an *abrB* mutation, is partially epistatic to the effects of an *slr* mutation.** A) ‘Colony’ column depicts 16 X images of colonies grown on MSgg semi-solid agar medium for 3 days at 22°C. Scale bar is 1 mm. ‘Pellicle’ column shows top-down images of 6-well microtitre plates in which cells have been grown statically in MSgg medium for 3 days at 22°C. Scale bar is 1 cm. The indicated wild type and mutant strains were as follows: wild type (3610), *sinR slr* (FC37), *abrB slr* (FC503). B,C) The indicated strains were grown at 30°C in MSgg and β-galactosidase was measured in Miller Units. B) β-galactosidase expression of  $P_{eps}$ -*lacZ* in wild type (♦)(FC13), *slr* (■) (FC526), *sinR* (▲)(FC14), *abrB* (□)(FC415), *sinR slr* (X)(FC527), *abrB slr* (○)(FC513). C) β-galactosidase expression of  $P_{yqxM}$ -*lacZ* in wild type (♦) (FC134), *slr* (■) (FC528), *sinR* (▲)(FC135), *abrB* (□)(FC416), *sinR slr* (X)(FC529), *abrB slr* (○)(FC504).

**Figure S4. Slr is a DNA-binding protein.** EMSA in which radiolabeled DNA’s (denoted below each panel) were mixed with purified N-terminal Slr at the indicated concentrations. Sizes of the DNA fragments containing *gfp* and  $P_{yqxM}$ , were 741 and 284 base pairs, respectively. The horizontal arrow indicates the position of free probe.

Figure S1

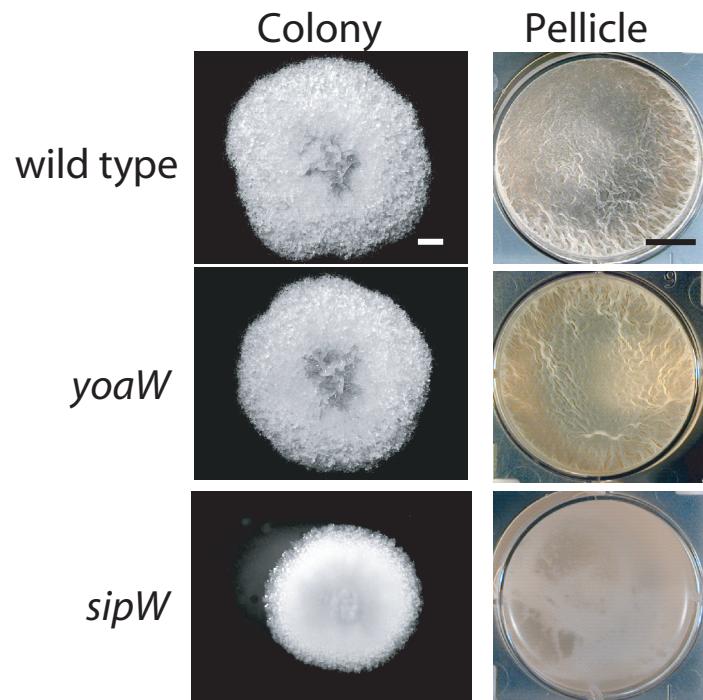
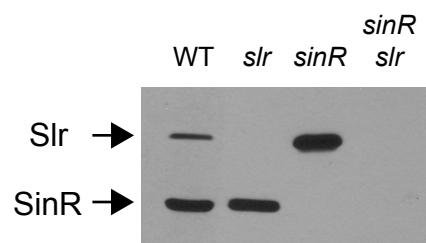


Figure S2



**Figure S3**

A

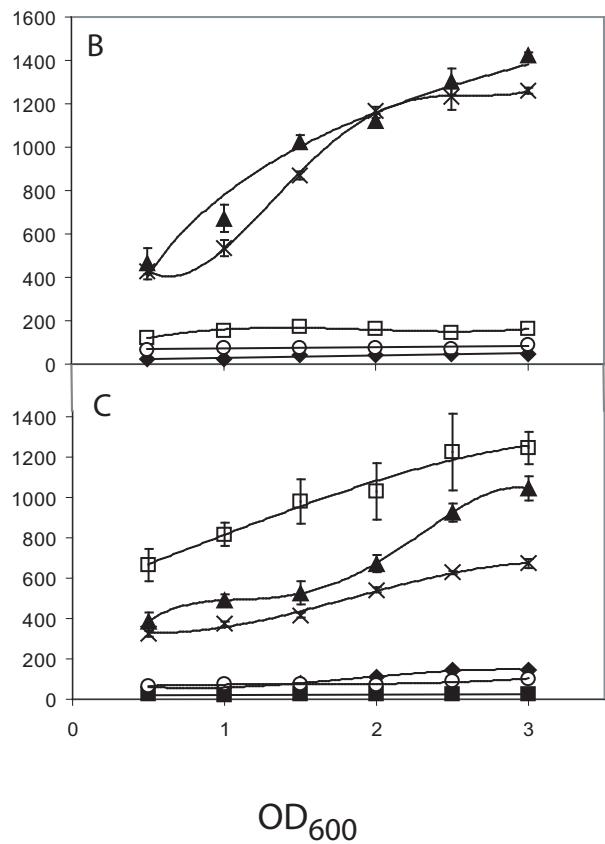
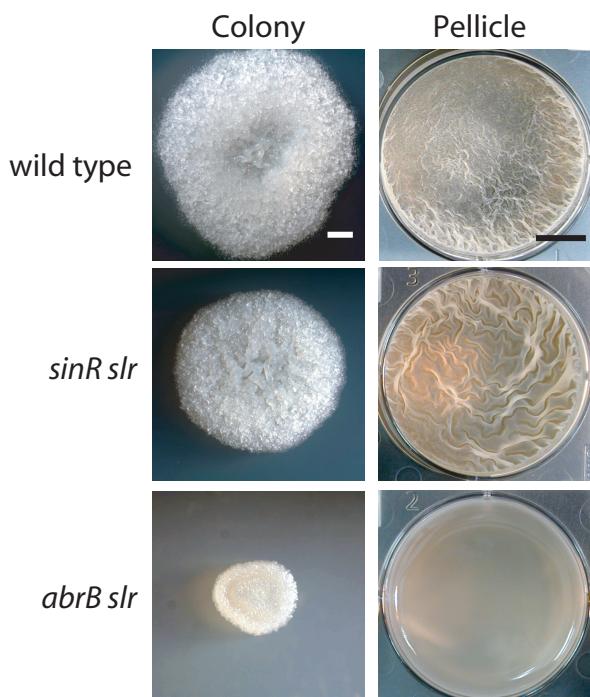


Figure S4

