Supporting Information

Stach et al. 10.1073/pnas.0710196105

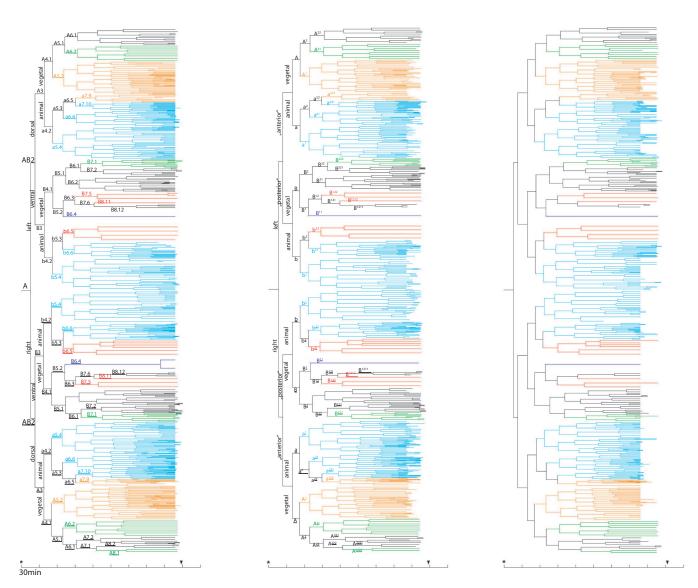


Fig. S1. Tree representations and fate maps of the development of three individuals of the tunicate chordate *Oikopleura dioica*. Colors indicated different tissues identifiable at hatching (see Fig. 1 for details). Left tree with nomenclature according to Conklin (1) and Nishida (2). Middle tree with nomenclature according to Delsman (3). Asterisks indicate times of fertilization, arrowheads mark times of hatching. Distance between two dashes on *x* axis corresponds to 30 min developmental time at 15°C.

- 1. Conklin EG (1905) Organization and cell lineage of the ascidian egg. J Acad Nat Sci Philadelphia Series 2, 13:1–119, plates I-XII.
- 2. Nishida H (1987) Cell lineage analysis in ascidian embryos by intracellular injection of a tracer enzyme. III. Up to the tissue restricted stage. Dev Biol 121:526–541.
- 3. Delsman HC (1910) Beiträge zur Entwicklungsgeschichte von Oikopleura dioica. Verhandelingen uit het Rijksinstituut voor het Onderzoek der Zee 3:1–24.

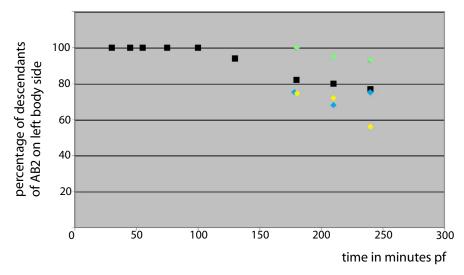


Fig. S2. Percentage of descendent cells of AB2 that remain on the left body side. To facilitate counting, tail cells were not considered. Black squares, total cell number; green, endodermal cells; blue, epidermis cells; yellow, nervous system cells.

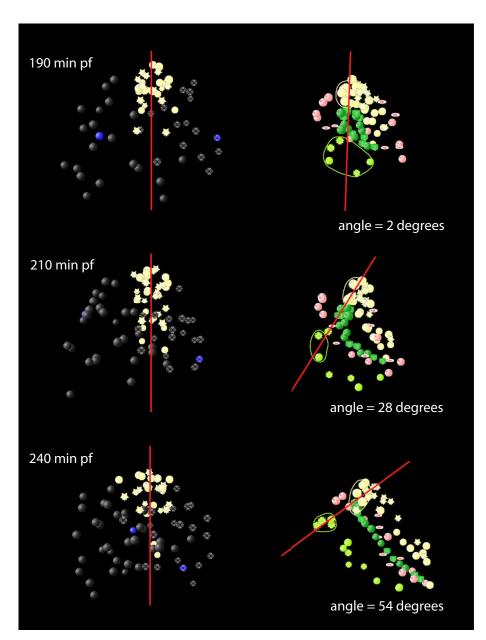


Fig. S3. Tail rotation observed during the development of *Oikopleura dioica* detected by using the 3D representation of the positions of the nuclei of each cell implemented in the 4D microscopy software SIMI°BioCell. Spheres derive from the right side (<u>AB2</u>). Black, endodermal and meso-/endodermal cells in the trunk; blue, prospective germ cells, green, (prospective) notochordal cells; light green, (prospective) endodermal strand cells; yellow, (prospective) nervous system cells (epidermis cells omitted for clarity). (*Left*) Anterior view of trunk, with axis of bilateral symmetry indicated by red line. (*Right*) Anterior view of tail in same angle of view, with anatomical axis of bilateral symmetry indicated by red line.

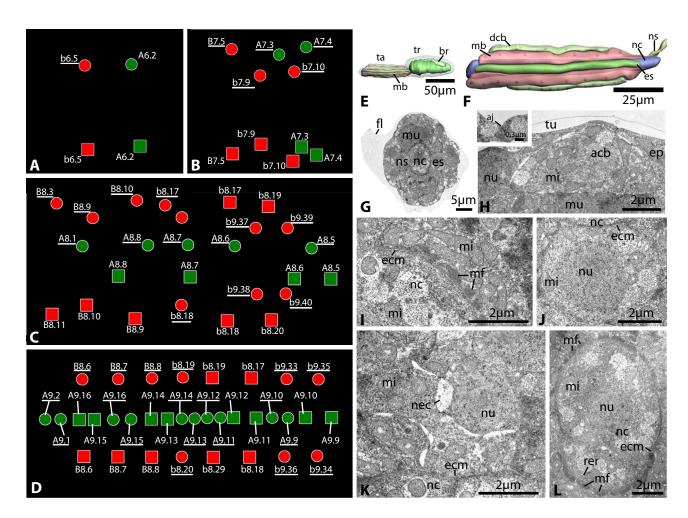


Fig. S4. (A–D) Schematic representations to depict the ontogeny of the tail muscle cells (red) and the notochord (green). Cells from the left side are represented as squares, cells from the right side as circles; view is from the anatomic dorsal side. (A) Seventy-two minutes postfertilization (PF). (B) One hundred four minutes PF. (C) One hundred sixty-eight minutes PF. (D) Two hundred twenty minutes PF. (Right) Anatomy of a newly hatched larva of the tunicate chordate Oikopleura dioica. (E and F) 3D representations of the larva based on serial transmission electron micrographs. (G–L) Transmission electron micrographs of cross sections through the tail. acb, additional cellular band, aj, apical junction, br, brain, dcb, additional dorsal cellular band, ecm, extracellular matrix, ep, epidermis cell, es, endodermal strand, fl, finlet, mb, band of muscle cells; mf, myofilaments; mi, mitochondrium; mu, muscle cell; nc, notochord; nec, neural canal; ns, nervous system; nu, nucleus; rer; rough endoplasmatic reticulum, ta, tail; tr, trunk; tu, tunic.

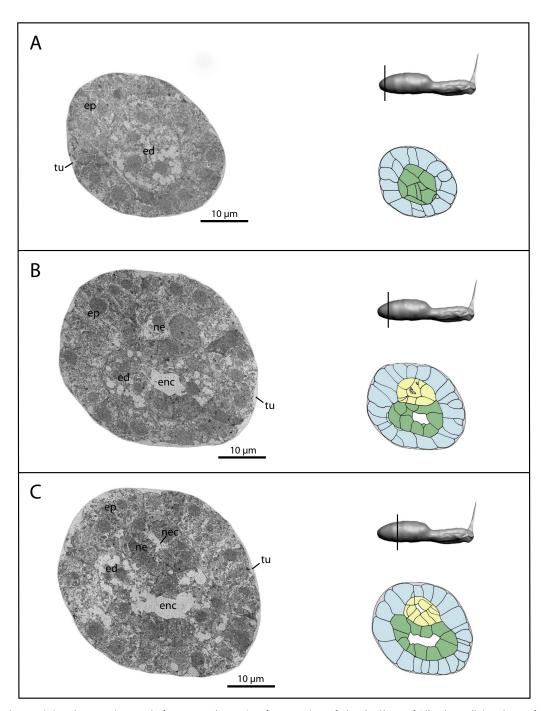


Fig. 55. Selected transmission electron micrographs from a complete series of cross sections of a hatched larva of Oikopleura dioica. Planes of sections indicated by black line in schematic in Upper Right; schematic representation of cross section in Bottom Right. Colors correspond to fates as in Fig. 1. Abbreviations: a, anterior; ax, axon; br, brain; brc, brain; ell; c1/2/3/...; cjl, cell junction; cm, cell membrane; d, dorsal; dd, dorsal cell; dmb, dorsal muscle band; dmc, dorsal muscle cell; ecm, extracellular matrix; ed - endodermal cell; eds, endodermal strand; ems, undifferentiated meso-/endoderm; enc, endodermal cavity; ep, epidermis; epc, epidermis cell; er, endoplasmatic reticulum; fl, finlet; gr, granules; l, left; mi, mitochondrion; my, myofibrillae; nc, notochord; ne, nervous system; nec, cavity in the nervous system; nm, nuclear membrane; no, nucleolus; nt, neural tube; nu, nucleus; nu 1, 2, 3, etc., nucleus 1, 2, 3, etc.; nup, nuclear pore; p,posterior; r,right; ta,tail; tr,trunk; tu, tunic; tu 1/2/3, tunic layer 1/2/3; v, ventral; vcb, ventral cellular band; ve, vesicle; vmb, ventral muscle band; vmc, ventral muscle cell.

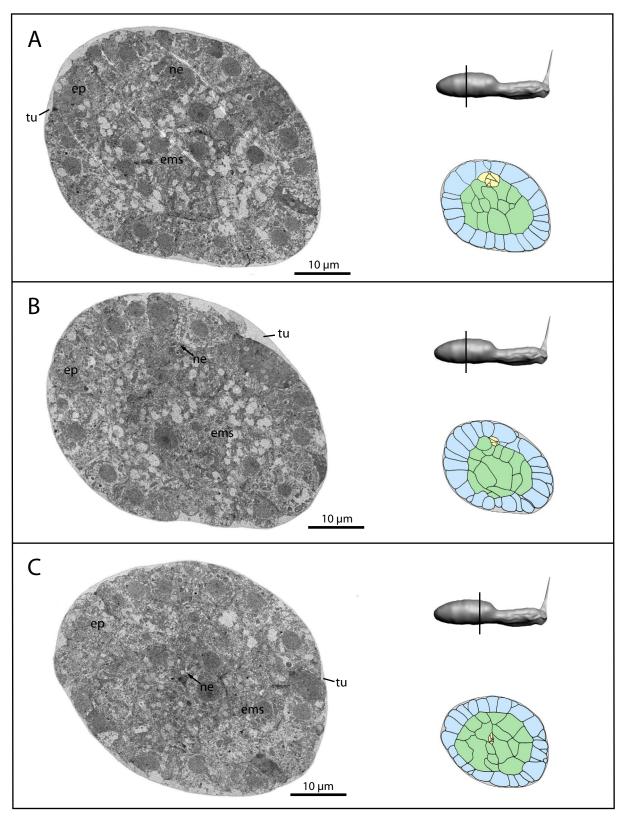


Fig. S6. Selected transmission electron micrograph from a complete series of cross sections of a hatched larva of Oikopleura dioica. For details and abbreviations, see Fig. S5 legend.

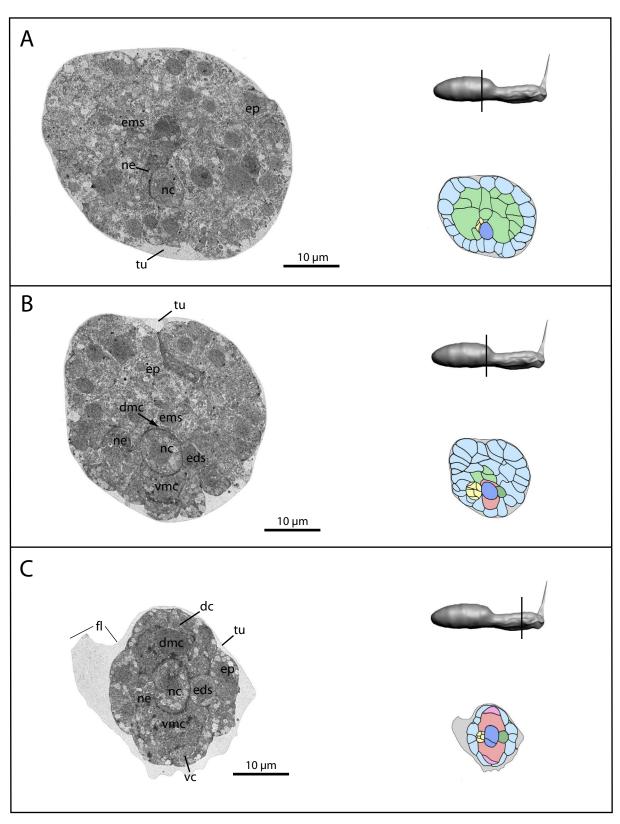


Fig. S7. Selected transmission electron micrograph from a complete series of cross sections of a hatched larva of Oikopleura dioica. For details and abbreviations, see Fig. S5 legend.

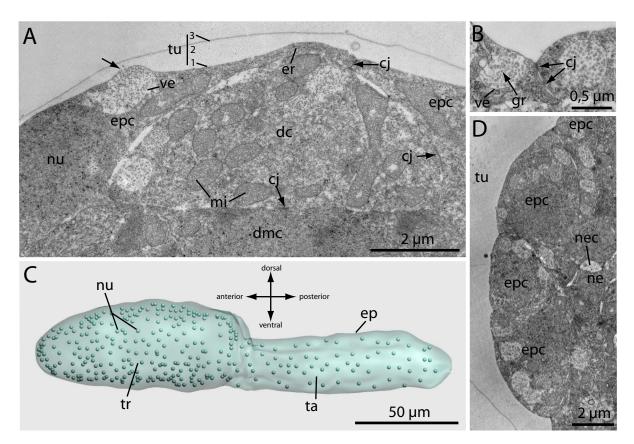


Fig. S8. Microscopic anatomy of epidermis in the hatched larva of *Oikopleura dioica*. 3D reconstruction and transmission electron micrographs at higher magnification. (*A*) Upper part of tail. (*B*) Junction between two neighboring epidermis cells. (*C*) 3D representation of epidermis of entire larva with nuclei of epidermis indicated by spheres. (*D*) Left part of tail with three cells that are probably prospective fin forming epidermis cells. For abbreviations, see Fig. S5 legend.



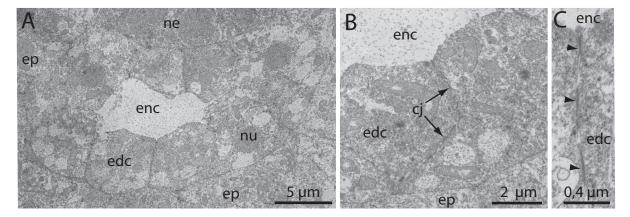
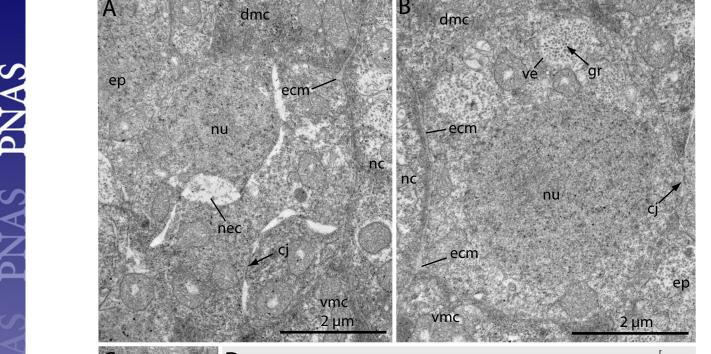


Fig. S9. Transmission electron micrographs of anterior endoderm in the hatched larva of Oikopleura dioica. For abbreviations, see Fig. S5 legend.



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Fig. S10. Microscopic anatomy of the tail in the hatched larva of Oikopleura dioica. 3D reconstruction and transmission electron micrographs at higher magnification. (A) Nerve cord. (B) Endodermal strand. (C) Extracellular matrix surrounding the notochord at higher magnification. (D) 3D representation of the tail in dorsal aspect. For abbreviations, see Fig. S5 legend.

dmc

0,5 µm

ecm-

ems

 $30 \, \mu m$

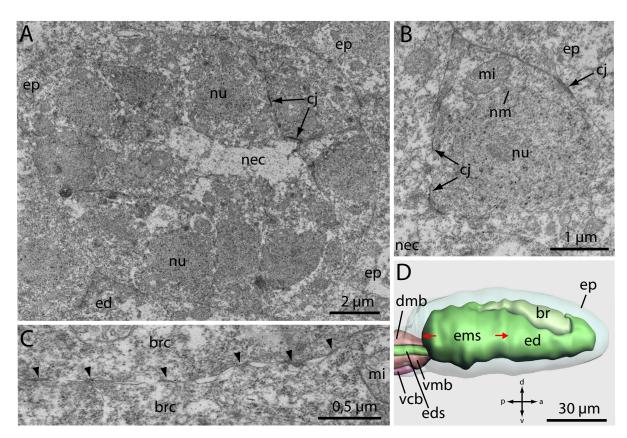


Fig. S11. Microscopic anatomy of the nervous system in the trunk of a hatched larva of Oikopleura dioica. 3D reconstruction and transmission electron micrographs at higher magnification. (A) Nervous system in anterior trunk. (B) Posterior brain region at the transition to the nerve connecting the brain with the nerve cord in the tail. (C) Cell junctions between nerve cells in anterior brain. (D), 3D representation of trunk. For abbreviations, see Fig. S5 legend.

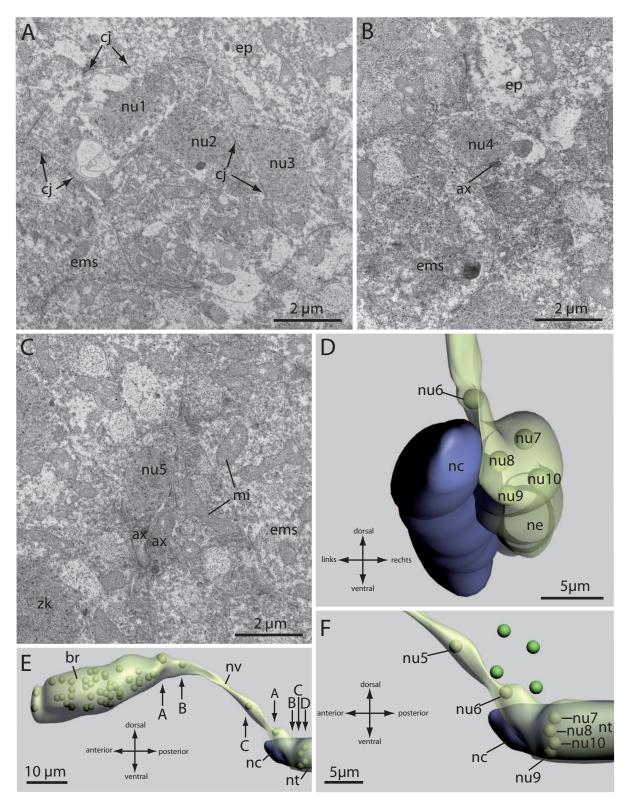
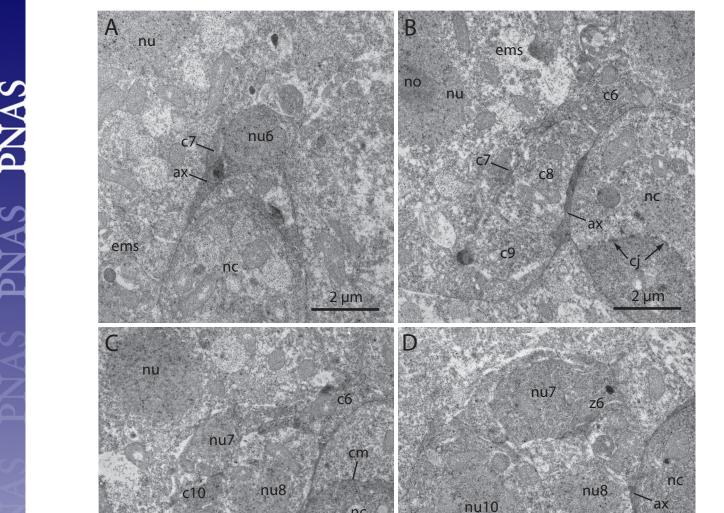


Fig. S12. Microscopic anatomy of the nervous system of a hatched larva of $Oikopleura\ dioica$. 3D reconstruction and transmission electron micrographs at higher magnification. (A–C) Consecutive sections from anterior to posterior; planes of section are indicated on the schematic 3D representation in E by the upward pointing arrows. (D–F) 3D representation of nervous system in different aspect. It could not be decided whether the four green nuclei in F belong to the nervous system or to the undifferentiated meso-endodermal cell mass. For abbreviations, see Fig. S5 legend.



nc

Fig. S13. Transmission electron micrographs of the nervous system of a hatched larva of Oikopleura dioica. (A-D) Consecutive sections from anterior to posterior; planes of section are indicated on the schematic 3D representation in Fig. S4G by the downward pointing arrows. For abbreviations, see Fig. S5 legend.

ems

nu9

ems

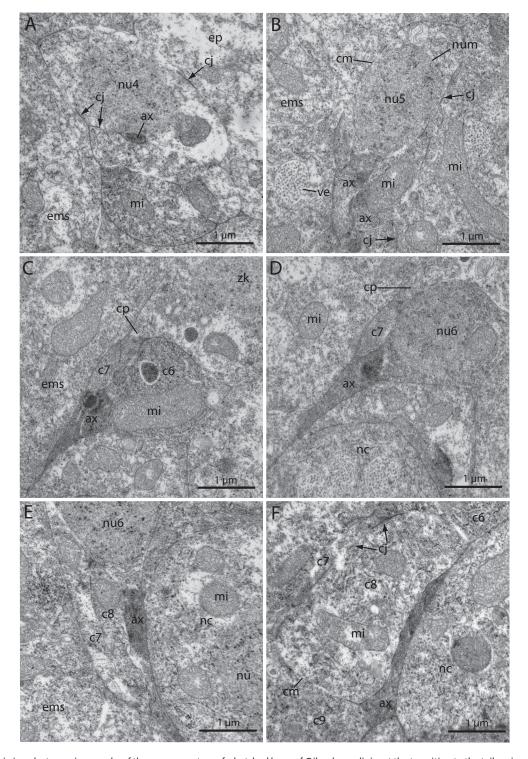


Fig. S14. Transmission electron micrographs of the nervous system of a hatched larva of *Oikopleura dioica* at the transition to the tail region. (A–F) Consecutive sections from anterior to posterior. Numbering of cells according to Fig. S4 G and H. For abbreviations, see Fig. S5 legend.

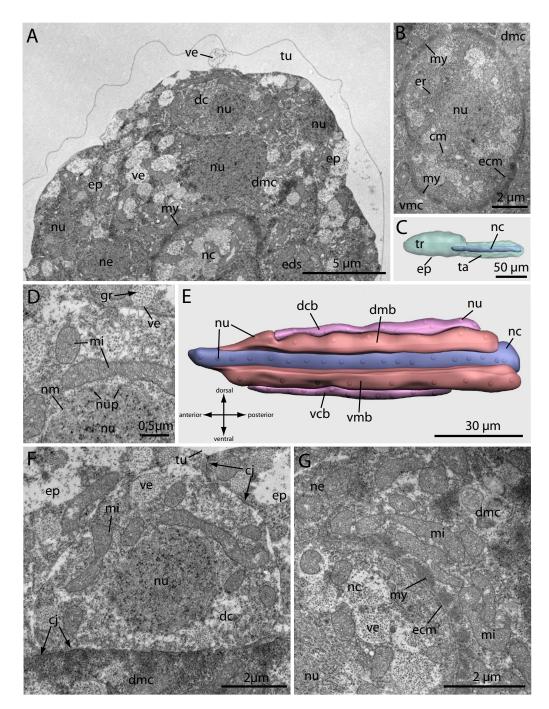


Fig. S15. Microscopic anatomy of the tail in the hatched larva of Oikopleura dioica. 3D reconstruction and transmission electron micrographs at higher magnification. (A) Cross section through tail. (B) Notochord. (C) 3D representation of complete larva in lateral aspect. (D) Higher magnification nuclear region of of muscle cell. (E) 3D representation of the tail, without epidermis and nerve tube in lateral aspect. (F) Higher magnification of a cross section through a cell situated dorsal to the dorsal muscle cell, yet beneath the epidermis. (G) Higher magnification of myofibrillae in the dorsal muscle cell close to the notochord. For abbreviations, see Fig. S5 legend.

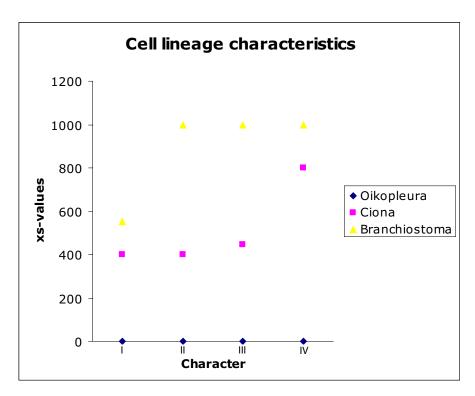
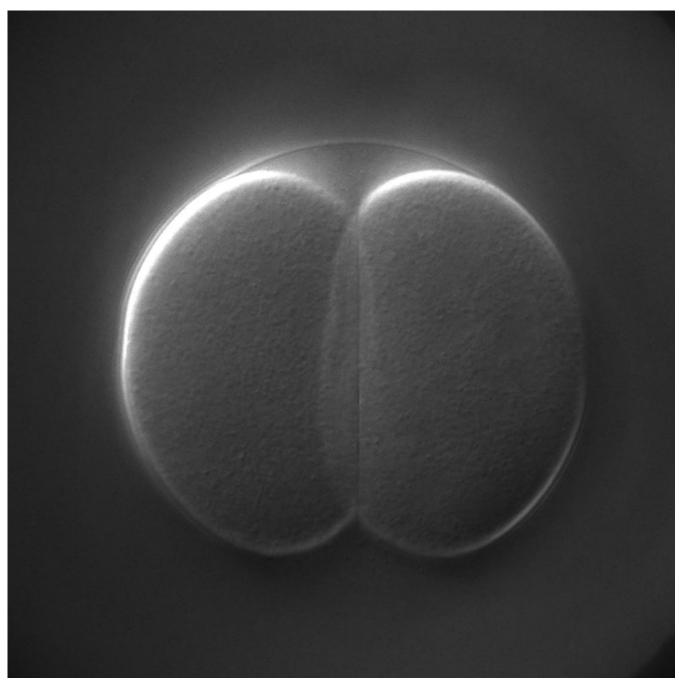
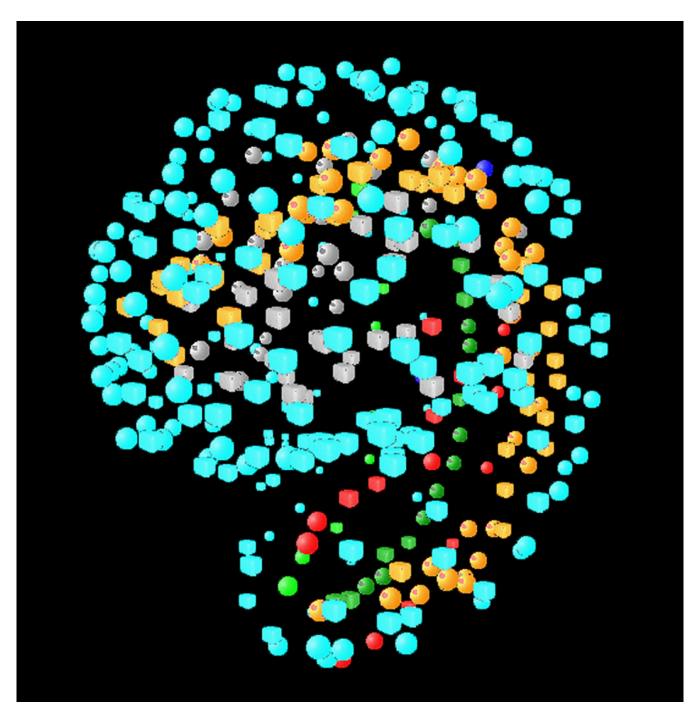


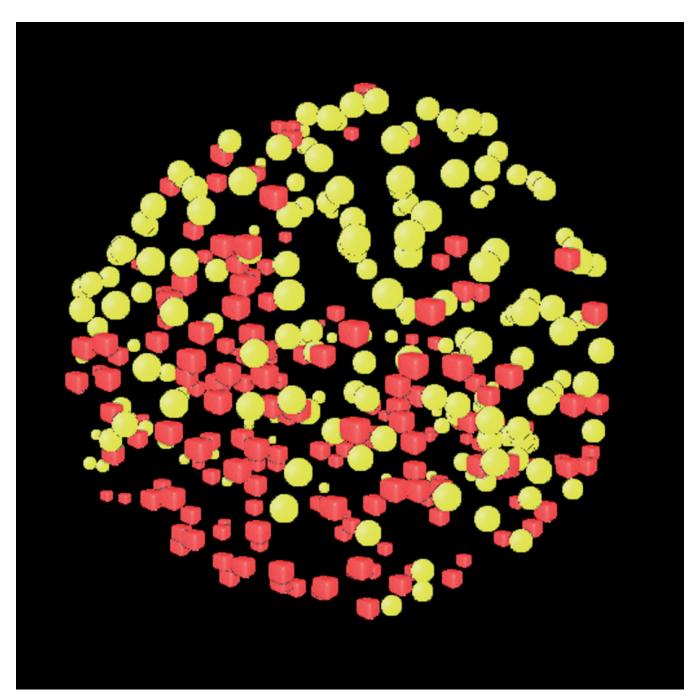
Fig. S16. Comparison of *x*_s values derived from Table S1 in the approach specified by Wiens [Wiens JJ (2001) Character analysis in morphological phylogenetics: Problems and Solutions. *Syst Biol* 50:689–699]. Note that no values belonging *to Oikopleura dioica* are intermediate between the cephalochordate *Branchiostoma* and the ascidian *Ciona* as would be expected, if *O. dioica* occupied a phylogenetic position at the base of the tunicate clade. Characters are: I, absolute time of occurrence of first cleavage; II, cell cycle number when blastula is formed; III, cell cycle number when gastrulation begins; IV, cell cycle number when neurulation begins.



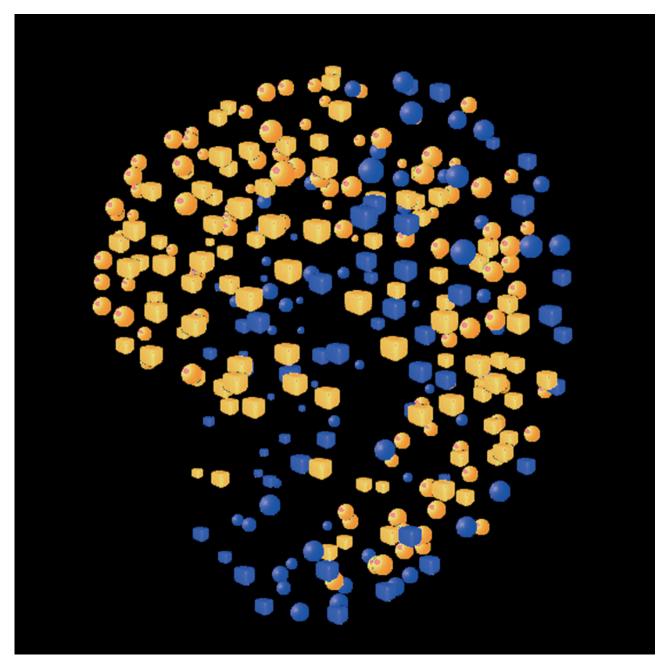
Movie \$1. Time-compressed movie of the development of Oikopleura dioica in light microscopic aspect.



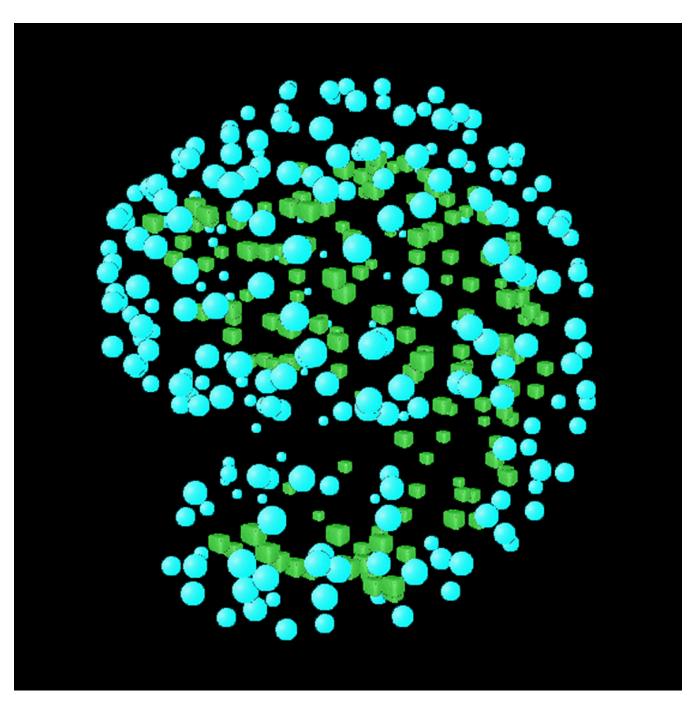
Movie S2. Development of Oikopleura dioica using 3D representations of the positions of the nuclei of each cell implemented in the 4D microscopy software SIMI°BioCell. Colors code for tissue fates as seen in Fig. 1. Aspect before hatching as in Fig. 3.



Movie 53. Movie of the development of *Oikopleura dioica*, using 3D representations of the positions of the nuclei of each cell implemented in the 4D microscopy software SIMI°BioCell. Virtual cell marking: red cubes, left sided cells derived from AB2; yellow spheres, right sided cells derived from AB2. Aspect before hatching as in Fig. 3, third column.



Movie 54. Movie of the development of *Oikopleura dioica*, using 3D representations of the positions of the nuclei of each cell implemented in the 4D microscopy software SIMI°BioCell. Virtual cell marking: blue, ventral cells (traditional "anterior"); orange, dorsal cells (traditional "posterior"); cubes, left-sided cells derived from AB2; spheres, right-sided cells derived from AB2. Aspect before hatching as in Fig. 3, fourth column.



Movie S5. Movie of the development of *Oikopleura dioica*, using 3D representations of the positions of the nuclei of each cell implemented in the 4D microscopy software SIMI°BioCell. Virtual cell marking: light blue, animal cells; light green, vegetal cells; cubes, left-sided cells derived from AB2; spheres, right-sided cells derived from *AB2*. Aspect before hatching as in Fig. 3, far right.

Table S1. Cell lineage characters for phylogenetic analysis and as estimates of heterochrony in chordate evolution.

						Fate restriction		Fate restriction	
	First cleavage,				Neurulation,	of tail	Fate restriction	of nervous	Fate restriction
Animal	Ref.	min	Blastula	Gastrulation	h	musculature	of notochord	system	of epidermis
Oikopleura	This study	20	3	5	6	6	6	5	5
Ciona	1–5	60	5	7	10	6	7	6	5
Halocynthia	6–8	120	5–6	7–8	9–10	7	7	7	5
Branchiostoma	9–15	75	8	9–10	≈11	na	na	na	na
Lampetra	_	180-240	na	na	na	na	na	na	na
Xenopus	16, 17	90	na	na	24	na	na	na	na
Ambystoma	18	200	11	≈16	na				
Echinus	19, 20	60-70	10	≈12	ne	ne	ne	na	na
Sphaerechinus	20		9		ne	ne	ne	na	na
Patiriella	21	40-60	6–9	30 h-35 h	ne	ne	ne	na	na
Saccoglossus	22, 23	100-120	>8	13 h–24 h	44–95	ne	ne	na	na

Figures given without units are cleavage numbers approximately in the middle of the respective event. Fate restrictions are given as the earliest determination of a clonal line dedicated to a single tissue type. na, data not available; ne, not applicable.

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Zygote	2-cells	4-cells	8-cells	16-cells	32-cells	62-cells	122-cells
	AB2	A3	A4.1 A	A5.1 A ²	A6.1 A^{22} A6.2 A^{2I}		
				A5.2 A ¹	A6.3 A^{12} A6.4 A^{11}		
			a4.2 a B4.1 B	a5.3 a ²	a6.5 a^{22}	$a7.9$ a^{222} $a7.10$ a^{221}	
					$a6.6$ a^{21}		
				a5.4 a' B5.1 B ²	a6.7 a ¹² a6.8		
					<i>a</i> ¹¹	B7.1	
					$B^{0.1}$	$B7.1$ B^{222} $B7.2$ B^{221}	
		В3			B6.2 B ²¹	В	
				B5.2 B ¹	B6.3 B ¹²	B7.5 B ¹²²	
A						B7.6 B ¹²¹	B8.11 B ¹²¹² B8.12 B ¹²¹¹
					B6.4 B ¹¹		
			b4.2 <i>b</i>	b5.3 b ²	b6.5 b ²²		
					b6.6 b ²¹		
				b5.4 b ¹	b6.7 b6.8		
					b^{II}		

Cells of the left side of the embryo are tabulated up to the stage where all tissue restrictions identified in the present study occured. Nomenclature on the right side is the same with denominations (Conklin) or superscripts (Delsman) underlined, see also Fig. 51. Colors indicate tissue restrictions (see Fig. 1).

^{1.} Conklin EG (1905) Organization and cell lineage of the ascidian egg. J Acad Nat Sci Philadelphia. 13(2):1–119, plates I-XII.

^{2.} Delsman HC (1910) Beiträge zur Entwicklungsgeschichte von Oikopleura dioica. Verhandelingen uit het Rijksinstituut voor het Onderzoek der Zee 3:1–24.