Order-Disorder-Order Transitions Mediate the Activation of Cholera Toxin - Supplementary Data

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CTA1:MNDDKLYRADSRPPDEIKQSGGLMPRGQSEYFDRGTQMNINLYDHARGTQTGFVRHDDGY
MNDDKLYRADSRPPDEIKQSGGLMPRGQSEYFDRGTQMNINLYDHARGTQTGFVRHDDGYCTA1-T4:VSTSISLRSAHLVGQTILSGHSTYYIYVIATAPNMFNVNDVLGAYSPHPDEQEVSALGGI
VSTSISLRSAHLVGQTILSGHSTYYIYVIATAPNMFNVNDVLGAYSPHPDEQEVSALGGI
VSTSISLRSAHLVGQTILSGHSTYYIYVIATAPNMFNVNDVLGAYSPHPDEQEVSALGGICTA1:PYSQIYGWYRVHFGVLDEQLHRNRGYRDRYYSNLDIAPAADGYGLAGFPPEHRAWREEPW
PYSQIYGWYRVHFGVLDEQLHRNRGYRDRYYSNLDIAPAADGYGLAGF
PYSQIYGWYRVHFGVLDEQLHRNRGYRDRYSNLDIAPAADGYGLAGF

CTA1: IHHAPPGCGNAPR

Supplementary Figure 1: a) Free CTA1 Constructs used for C-terminal truncation studies. CTA1 full (N1-R192), CTA1-T2 (N1-F167) and CTA1-T4 (N1-G144). The six C-terminal residues that are deleted in CTA1 (N1-G186) are boxed.



Supplementary Figure 2: Preparative gel filtration FPLC chromatography traces of refolded CTA1 (N1-G186) by solubilization in a) 6 M Guanidine-HCI and b) 8 M Urea denaturation solutions. From standard molecular weight markers, the expected retention volume of the 21.1 kDa monomer peak was about 69 mL for both samples. Sample volume was 2 mL. Note the presence of a shoulder peak in the urea sample at 58.4 mL. Guanidine refolding was preferred due to the absence of a residual dimeric species, which was present in the urea refolded samples (shoulder peak at 58.4 mL).



Supplementary Figure 3: Both soluble expressed NhisCTA1 and inclusion body refolded CTA1 show similar intensity variations of both the random coil resonances and dispersed resonances as evident from the expansion of the amide region within their 1D-¹H NMR spectrum. a) 6 μ M soluble expressed CTA1 (1-192) (E110:112D) obtained from an extended incubation of the CTA1:ARF6-GTP complex (O'Neal *et al. Science* 2005) in 10 mM EDTA; the CTA1 domain was then purified by dilution and Ni-NTA affinity chromatography and exchanged into 10mM dTris, pH 7.2, 200 mM NaCl and 6% v/v ²H₂O; number of scans = 8192. (Note some solvent peaks remain) b) ~100 μ M inclusion body (refolded) CTA1 (CTA1 1-186) in the same NMR buffer; number of scans = 32.



Supplementary Figure 4: ¹H/¹⁵N HSQC spectral comparison of CTA1 and C-terminal truncation mutants. a) CTA1 (N1-G186), b) CTA1-T2 and c) CTA1-T4. Spectra were acquired at 288 K on the Bruker Avance 800 MHz.



Supplementary Figure 5: Sequence specific assignment of CTA1-T2. a) CTA1-T2 assigned connectivity strip-plot from T82-I88. The HN(CO)CACB and HNCACB were used where Ca (yellow peaks) and CB (red peaks) were connected to display a sequential walk. Figure was produced by SPARKY. b) Assigned 1 H/ 15 N HSQC spectrum of 250 μ M CTA1-T2 acquired on Bruker Avance 800 MHz (8 scans at 288 K). BMRB accession number 15162.

Supplementary Table 1: Residues which show intense peak volumes in the ¹ H/ ¹⁵ N HSQC spectra (>20X increase over that of the downfield shifted amides).				
D2	A 4 E	L77	G133	A156
D3	A45	H80	D136	A159
K17	R46	S81	E137	D160
G34	G47	T82	0138	G161
T35	T48	102		V160
Q36	G51	V 100	L139	¥ 102
N38	F52	L101	R141	L164
N40	\/53	G102	S151	A165
1140	V 33	A103	L153	G166
L41	K54	Y104	D154	F167



Supplementary Figure 6: Structural mapping of residues which show intense peak volumes in the NMR Spectra: a) Ribbon diagram of the enzymatic domain from the CTA1:ARF6-GTP crystal structure (pdb# 2A5F) showing residues N1-F167 that correspond to the termini of CTA1-T2, while b) is the same diagram, rotated 180°. The boundaries of the secondary structural elements were based on the CSI data of CTA1-T2 (Figure 5a, main text), with structural region indicated by color: CTA1₁ (N1-N142) is shown in yellow, while CTA1₂ (residues R143-G161) is shown in black and CTA1₃ (Y162-F167, the C-terminus of CTA1-T2) is shown in light green. Amide nitrogen atoms of residues with intense peak volumes (Supplementary Table 1) are shown in blue CPK. Figure was generated in MolMol.