## **Supporting Information**

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## SI Text

**Strains, Plasmids, and Culture Conditions.** All *V. cholerae* strains used in this study were derived from E1 Tor C6706 and were propagated in LB media containing appropriate antibiotics at 37°C, unless otherwise noted. AKI medium was used to induce virulence gene expression (1).

The *tcpA* promoter was fused to the *sh ble* gene, which had been amplified by PCR from pEM7/Zeo (Invitrogen), and cloned into the plasmid pCVD442 (2). The resulting construct was integrated into the *lacZ* locus within the chromosomes of various C6706-derivatived strains. The *aphA-lacZ* transcriptional reporter was constructed by cloning the *aphA* promoter se-

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quence into a plasmid containing a promoterless *lacZ* and a pBBR322 origin (3). The construction details of the *hapR-lacZ* and *hapR-K*<sub>m</sub>' transcriptional fusions were described (4, 5). To construct FlgM-His<sub>6</sub> and FliA-His<sub>6</sub>, *flgM* and *fliA* coding sequences were amplified with C-terminal His<sub>6</sub> tags by PCR and cloned into the plasmid pBAD24, which contains an arabinose-inducible promoter (3). Chromosomal in-frame deletions were constructed by cloning the two flanking regions of the target gene into the suicide vector pWM91, which also contains a *sacB* counterselectable gene. For each mutant, selection for double-cross-over homologous recombination as described (6) resulted in the desired in-frame deletion.

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**Fig. S1.** Repression of *hapR* facilitates activation of a cascade of transcription factors that regulate the expression of virulence genes. HapR represses *aphA*, which encodes the transcription factor that up-regulates *tcpP* expression. The gene *tcpP* encodes the transcription factor TcpP, which in turn up-regulates *tcpP* expression. ToxT regulates the expression of multiple virulence genes, including *tcpA*, which encodes the major TCP pilin. Also shown in the figure is LuxO, which is phosphorylated under conditions of low cell density. In its phosphorylated form, LuxO activates the expression of several small RNAs that repress *hapR* expression (7, 8). Thus, under conditions of low cell density, *hapR* expression is repressed, and virulence genes are transcribed.



**Fig. 52.** The flagellar rod protein FlgD affects expression of virulence genes via quorum sensing. (*A*). The production of virulence factors in the *luxO* mutant is restored by a *flgD* mutation. Strains were grown in AKI medium (1), which induces virulence genes. TcpA production was assayed by Western blot analysis (9), and cholera toxin (CT) production was assayed by ELISA (10). All samples were normalized to contain 10<sup>9</sup> bacterial cells. (*B*). Expression of *aphA* in the *luxO* mutant is restored by mutation of *flgD*. Strains containing the *aphA-lacZ* transcriptional reporter plasmid were grown to midlog in AKI medium.  $\beta$ -Galactosidase activity was measured and reported as described (11). (*C*). Mutation of *flgD* attenuates regulation by quorum sensing. Strains harboring pBB1 from *V*. harveyi were diluted into fresh LB media and grown at 30°C. Luminescence and cell density were measured at various time points. Relative luminescence units (RLU) were calculated as light units/OD<sub>600</sub> ml<sup>-1</sup>. (*D*). Mutation of *flgD* reduces expression of *hapR*. Strains containing the *hapR-lacZ* chromosomal transcriptional fusion were experiments ± standard deviations.





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Fig. S4. Motility and mouse colonization of flagellar mutants. (A) Wild type and flagellar mutants were spotted on LB containing 0.3% agar and incubated at 37°C for 6 h. (B). Competitive index (CI) of *fliA hapR* and *fliA* mutants calculated from colonization of each infant mouse. Red line represents the average of CI.

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High infection dose of *V. cholerae* 

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Many killed by host antimicrobial agents

In intestines, low cell density represses quorum sensing Passage through mucosa breaks flagella to secrete FlgM FliA activation further represses *hapR V. cholerae* is primed for activation of virulence genes

Fig. S5. The proposed model of enhanced expression of virulence genes due to passage through the mucosal layer by V. cholerae during host colonization. See Discussion in the text for details.



Movie S1. The motility of bacteria collected from transwell without 1% mucin.

Movie S1(MOV)

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Movie S2(MOV)

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## Table S1. Locations and relevant phenotypes of transposon insertions that activate tcpA expression in luxO mutants

	Gene name	Gene function	Zeocin-resistance*	CT/TcpA production <sup>†</sup>	Lux activity <sup>‡</sup>
Controls	Wild type		+	++	++
	luxO		—	-	+++
	hapR		+	++	_
Tn results in constitutive expression <sup>§</sup>	VC2338 ( <i>lacZ</i> )	Express P <sub>tcpA-</sub> sh ble directly	+	—	+++
	VC2647(aphA)	Virulence gene regulator	+	++	+++
	VC0826( <i>tcpP</i> )	Virulence gene regulator	+	++	+++
	VC0838 ( <i>toxT</i> )	Virulence gene regulator	+	++	+++
	VC2066( <i>fliA</i> )	Late flagellar synthesis regulator	+	++	+
Tn results in disruption <sup>¶</sup>	VC0583(hapR)	Quorum sensing regulator	+	++	_
	VC1130(vicH)	H-NS-like protein	+	++	+++
	VC2305(ompK)	Outer membrane protein	+	++	+++
	VC2271(ribD) VC2198 (flgD)	Riboflavin biosynthesis	+	+	+++
	VC2199 (flgC) VC2200 (flgB)	Flagellar biosynthesis, rod proteins	+	+	+

All strains contain the P<sub>tcpA</sub>-sh ble reporter in the lacZ locus and a cosmid harboring luxCDABE from V. harveyi (pBB1).

\*Strains were grown in AKI and zeocin-resistant colonies were counted on LB plates.

<sup>†</sup>Strains were grown in AKI and cholera toxin ELISA and TcpA Western blots were performed.

<sup>+</sup>Strains were grown in LB at 30°C to midlog and luminescence was measured.

<sup>§</sup>All phenotypes were dependent on arabinose induction.

<sup>¶</sup>All phenotypes were independent of arabinose.

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