SUPPLEMENTARY MATERIALS

Supplementary Figure 1. Expression of outer border cells markers.

Expression of Slbo-lacZ (**a, b**) and E(spl)-LacZ (**c, d**) markers in which the two anterior polar cells were either heterozygous (**a, c**) or homozygous mutant for DralA (**b, d**). Markers expression is normal as is migration of the cluster (arrowheads) when cells are heterozygous (a, c). In contrast, the expression of both markers is absent when polar cells are mutant for DralA (b, d). Stars indicate the theoretical position of the cluster if it would have been wild type. **a**, **c**, stage 10B egg chambers; **b**, **d**, stage 9 egg chambers.

Supplementary Figure 2. Border cell migration phenotype in egg chambers overexpressing Rap1 dominant negative.

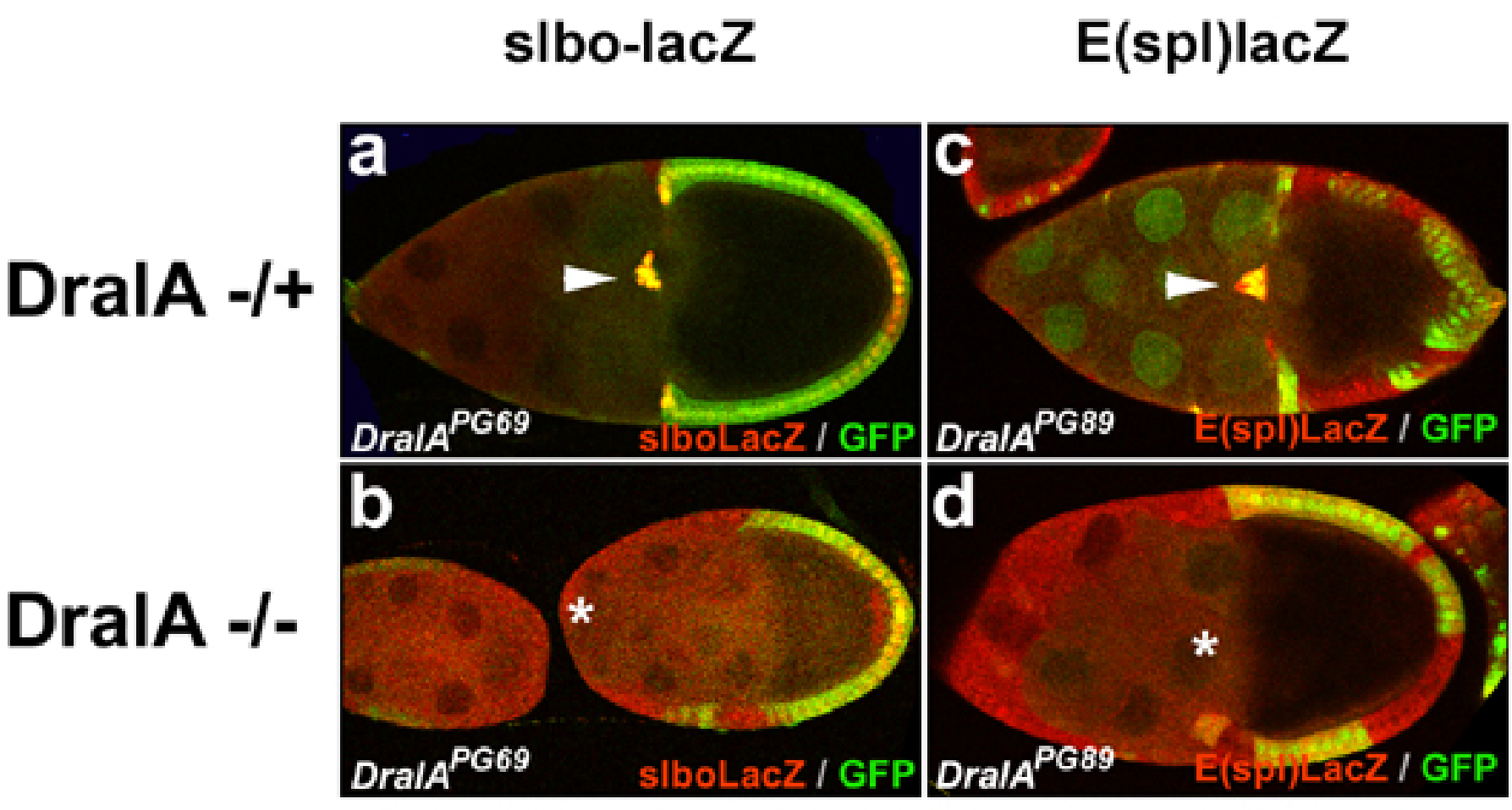
Expression of UAS-Rap1DN in border cells block their migration; stage 10 egg chamber (**a**). (**b**) percentage ofmigration defects in stage 10 egg chambers following expression of Rap1DN.

Supplementary Methods

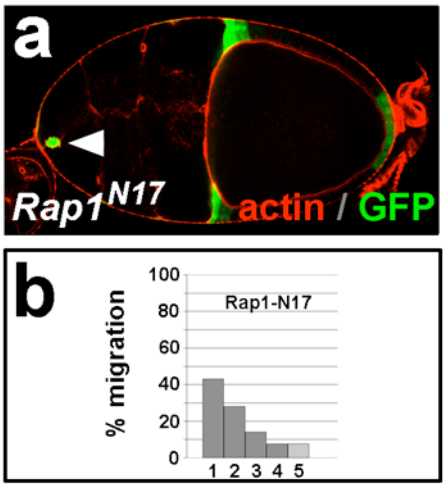
DNA cloning. pUAS-*DralA*-GFP was constructed by single-step ligation of a DNA fragment containing the 5'UTR and coding regions of *DralA*, amplified by PCR and cloned in the pUAS-(C)GFP plasmid. pUAS-GFP-*DralA* was constructed by single-step ligation of a *DralA* DNA fragment containing the coding region amplified by PCR and cloned in the pUAS-(N)GFP plasmid. GFP-*DralA*$^{\Delta CTLL}$ was made by introducing, using PCR, a stop codon at amino-acid 197.

The UAS constructs were introduced into w^{1118} flies by standard methods of P-element-mediated germline transformation, using the Δ2-3 helper plasmid. For each construct, several independent transgenic lines were generated and tested.

In situ hybridization. In situ hybridization to ovaries was performed as described (Queenan et al., 1997), using a digoxigenin-labeled antisense RNA probe encompassing the entire *DralA* cDNA. A sense probe was used in parallel as a control.



Supplemental fig. 1 (Ghiglione et al.)



suppl. fig. 2 (Ghiglione et al.)